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(57) Abstract

The invention provides human membrane transport proteins (MTRP) and polynucleotides which identify and encode MTRP. The invention also provides expression vectors, host cells, antibodies, agonists, and antagonists. The invention also provides methods for diagnosing, treating, or preventing disorders associated with expression of MTRP.

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MEMBRANE TRANSPORT PROTEINS

TECHNICAL FIELD

This invention relates to nucleic acid and amino acid sequences of membrane transport proteins and to the use of these sequences in the diagnosis, treatment, and prevention of membrane transport disorders; immune/inflammatory disorders; and cell proliferative disorders including cancer.

BACKGROUND OF THE INVENTION

Eukaryotic cells are bound by a lipid bilayer membrane and subdivided into functionally distinct, membrane bound compartments. The membranes maintain essential differences between the cytosol, the extracellular environment, and the contents of intracellular organelles such as the Golgi or the endoplasmic reticulum. As lipid membranes are highly impermeable to most polar molecules, transport of essential nutrients; metal ions such as K*, NH₄*, P_i, SO₄²⁻; sugars; vitamins; metabolic waste products; cell signaling molecules; drugs; peptides; and proteins and other macromolecules across lipid membranes and between organelles must be mediated by a variety of transport molecules. Many transport mechanisms are substrate specific, with each transport protein carrying particular members of a molecular class, such as ions, sugars, or amino acids, across membranes. For example, amino acids are imported into cells via specific amino acid permeases.

Transport proteins are multi-pass transmembrane proteins, which either actively transport molecules across the membrane or passively allow them to cross. Active transport involves directional pumping of a solute across the membrane, usually against an electrochemical gradient. Active transport is tightly coupled to a source of metabolic energy, such as ATP hydrolysis or an electrochemically favorable ion gradient. Passive transport involves the movement of a solute down its electrochemical gradient. Transport proteins can be further classified as either carrier proteins or channel proteins. Carrier proteins, which can function in active or passive transport, bind to a specific solute to be transported and undergo a conformational change which transfers the bound solute across the membrane. Channel proteins, which only function in passive transport, form hydrophilic pores across the membrane. When the pores open, specific solutes, such as inorganic ions, pass through the membrane and down the electrochemical gradient of the solute.

Transport proteins play roles in antibiotic resistance, toxin secretion, ion balance, synaptic neurotransmission, kidney function, intestinal absorption, tumor growth, and other diverse cell functions (Griffith, J. and C. Sansom (1998) The Transporter Facts Book, Academic Press, San Diego CA, pp. 3-29). A variety of human inherited diseases are caused by mutation of transport proteins. For example, cystinuria is an inherited disease that results from the inability to transport cystine, the

disulfide-linked dimer of cysteine, from the urine into the blood. Accumulation of cystine in the urine leads to the formation of cystine stones in the kidneys. Also, many transport proteins are composed of subunits that may confer specificity for the tissue in which the transport mechanism functions, and are therefore associated with tissue-specific disorders. Examples of transport proteins include facilitative transporters, the secondary active symporters and antiporters driven by ion gradients, and active ATP binding cassette transporters involved in multiple-drug resistance and targeting of antigenic peptides to MHC Class I molecules, and the E1-E2 cation transport ATPases.

Carrier proteins which transport a single solute from one side of the membrane to the other are called uniporters. In contrast, coupled transporters link the transfer of one solute with simultaneous or sequential transfer of a second solute, either in the same direction (symport) or in the opposite direction (antiport). For example, intestinal and kidney epithelium contains a variety of symporter systems wherein the movement of sodium into the cell down its electrochemical gradient co-transports a second solute into the cell. The sodium gradient that provides the driving force for solute uptake is maintained by the ubiquitous Na*/K* ATPase. Sodium-coupled transporters include the mammalian glucose transporter (SGLT1), iodide transporter (NIS), and multivitamin transporter (SMVT). These three transporters have twelve putative transmembrane segments, extracellular glycosylation sites, and cytoplasmically-oriented N- and C-termini. NIS plays a crucial role in the evaluation, diagnosis, and treatment of various thyroid pathologies because it is the molecular basis for radioiodide thyroid-imaging techniques and for specific targeting of radioisotopes to the thyroid gland (Levy, O. et al. (1997) Proc. Natl. Acad. Sci. USA 94:5568-5573). SMVT is expressed in the intestinal mucosa, kidney, and placenta, and is implicated in the transport of the water-soluble vitamins, e.g., biotin and pantothenate (Prasad, P.D. et al. (1998) J. Biol. Chem. 273:7501-7506).

The largest and most diverse family of transport proteins is the ATP-binding cassette (ABC) transporters. As a family, ABC transporters can transport substances that differ markedly in chemical structure and size, ranging from small molecules such as ions, sugars, amino acids, peptides, and phospholipids, to lipopeptides, large proteins, and complex hydrophobic drugs. Each ABC transporter consists of four modules: two nucleotide-binding domains (NBDs), which hydrolyze ATP to supply the energy required for transport; and two membrane-spanning domains (MSDs), which may form membrane channels. The NBDs consist of approximately two hundred conserved amino acid residues while the MSDs each contain six putative transmembrane segments. (See, e.g., Saurin, W. et al. (1994) Mol. Microbiol. 12:993-1004; Shani, N. et al. (1996) J. Biol. Chem. 271:8725-8730; Koster, W. and B. Bohm (1992) Mol. Gen. Genet. 232:399-407.) The four ABC transporter modules may be encoded by a single gene, as is the case for the cystic fibrosis transmembrane conductance regulator (CFTR), or by separate genes. When encoded by separate genes, each gene product

contains a single NBD and MSD. These "half-molecules" form homo- and heterodimers, such as Tap1 and Tap2, the endoplasmic reticulum-based major histocompatibility (MHC) peptide transport system associated with antigen processing (Androlewicz, M.J. et al. (1994) Proc. Natl. Acad. Sci. USA 91:12716-12720).

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Several genetic diseases are attributed to defects in ABC transporters, including the following diseases and their corresponding proteins: cystic fibrosis (CFTR, an ion channel; Welsh, M.J. and A.E. Smith (1993) Cell 73:1251-1254); X-linked adrenoleukodystrophy, an inborn error of peroxisomal β-oxidation of very long chain fatty acids (adrenoleukodystrophy protein, ALDP); Zellweger syndrome, an inborn error of peroxisome biogenesis (peroxisomal membrane protein-70, PMP70); and hyperinsulinemic hypoglycemia (sulfonylurea receptor, SUR). The ABC transporters known as P-glycoproteins, or multidrug resistance (MDR) proteins, are associated with resistance to a wide range of hydrophobic drugs (MDR1; Gottesman, M.M. and I. Pastan (1993) Annu. Rev. Biochem. 62:385-427) or with phosphatidylcholine transport (MDR2; Ruetz, S. and P. Gros (1994) Cell 77:1071-1081). MDR is common in cancer cells, and contributes to low efficacy or failure of chemotherapy (Taglight, D. and S. Michaelis (1998) Methods Enzymol. 292:131-163). MDR is mediated by transporters, e.g., P-glycoproteins or the multidrug resistance-associated protein MRP, that normally function in the liver, intestines, and kidney to move toxic substances from the cytosol into the bile, intestinal lumen, or urine. In cancerous cells, these transporters extrude chemotherapeutic agents into the extracellular space, thereby conferring drug resistance. Recently, an ABC transporter-type protein was isolated from a human leukemia cell line. This transporter, termed the anthracycline resistance associated protein (GI 1279457, SEQ ID NO:42), is overexpressed in a multidrug resistant leukemia cell sub-line, and has sequence homology with other multidrugresistance associated proteins including MRP (Longhurst, T.J. et al. (1996) Br. J. Cancer 74:1331-1335).

Transport of fatty acids across the plasma membrane can occur by diffusion, a high capacity, low affinity process. However, under normal physiological conditions a significant fraction of fatty acid transport appears to occur via a high affinity, low capacity protein-mediated transport process. Fatty acid transport protein (FATP), an integral membrane protein with four transmembrane segments, is expressed in tissues exhibiting high levels of plasma membrane fatty acid flux, such as muscle, heart, and adipose. Expression of FATP is upregulated in 3T3-L1 cells during adipose conversion, and expression in COS7 fibroblasts elevates the cells' uptake of long-chain fatty acids. Expression studies suggest a role for FATP in lipid metabolism, obesity, and type II diabetes mellitus (Hui, T.Y. et al. (1998) J. Biol. Chem. 273:27420-27429).

E1-E2 (or P-type) ATPases constitute a superfamily of cation transporters present in both

prokaryotes and eukaryotes that mediate membrane flux of all biologically relevant cations. These ATPases are postulated to exist in two different conformational states, designated E1 and E2, during the course of the ATP hydrolysis reaction, and to conserve the energy from ATP hydrolysis in the form of an acyl phosphate, primarily an aspartyl phosphate. Members of this family are divided into four major groups; the Ca⁺²-transporting ATPases, Na⁺/K⁺ -and gastric H⁺/K⁺-transporting ATPases, plasma membrane H⁺-transporting ATPases (proton pumps), and the bacterial P-type ATPases (BLOCKS: BL00154, P-type cation-transporting ATPase superfamily signature).

The metabolism of amino acids is complex and highly regulated. While cells are capable of creating most amino acids de novo, the import of amino acids into cells via specific amino acid permease proteins is vital for maintaining the appropriate and complete availability of all necessary amino acids. This is particularly important during cell proliferation and differentiation. In addition to their role as protein building blocks, amino acids also serve as precursors for a variety of other important macromolecules. For example, the hormone thyroxine, the pigment melanin, and the neurotransmitters histamine, epinephrine, and serotonin are produced from various amino acid precursors, including histidine, tyrosine, and tryptophan. A component of sphingolipid formation, sphingosine, is derived from serine. Porphyrin rings, which are components of heme molecules, use glycine as a nitrogen donor. Significant portions of the ring structures of purines and pyrimidines, components of nucleic acids, are formed from the breakdown of numerous amino acids. Amino acids are also important in energy metabolism. Unlike fatty acids and glucose, amino acids cannot be stored in the cell, so excess amino acids are fed into the citric acid cycle to produce energy molecules including fatty acids, ketone bodies, and glucose. Thus, precise control of amino acid metabolism is extremely important to both proliferating and non-proliferating cells.

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The E16 gene, cloned from human peripheral blood lymphocytes, encodes a 241 amino acid integral membrane protein with multiple predicted transmembrane domains (Gaugitsch, H.W. et al. (1992) J. Biol. Chem. 267:11267-11273). E16 gene expression is closely linked to cellular activation and division. In myeloid and lymphoid cells, E16 transcripts are rapidly induced and rapidly degraded after stimulation. This pattern of expression resembles the kinetics seen for proto-oncogenes and lymphokines in the T cell system. Elevated levels of E16 expression were detected in colonic, gastric, and breast adenocarcinomas, and in lymphoma, while little or no E16 expression was detected in normal (non-cancerous) human tissues such as adult brain, lung, liver, colon, esophagus, stomach, or kidney, nor in four-month fetal brain, lung, liver, or kidney (Wolf, D.A. et al. (1996) Cancer Res. 56:5012-5022; Gaugitsch et al., supra). E16 was detected in every cell line tested. Its presence in rapidly dividing cell lines and its absence in human tissues with low proliferative potential suggest that E16 is directly involved in the cell division process, where it helps provide important

building blocks for energy metabolism, biochemical synthetic pathways, and protein synthesis.

Post-translational modification of polypeptides occurs in the lumen of the Golgi apparatus. Such modifications include, for example, the addition of sugar molecules by enzymes such as N-acetylglucosaminyltransferase, to produce glycoproteins. The sugar-donating molecules in this reaction are typically nucleotide sugars, such as uridine diphosphate-galactose (UDP-Gal). UPD-Gal and other nucleotide sugars are transported from the cytosol into the Golgi apparatus by specific transporter molecules. The availability of these nucleotide sugars can regulate which glycoproteins are synthesized, and therefore has a significant impact on cellular function (Toma, L. et al. (1996) J. Biol. Chem. 271:3897-3901; Guillen, E. et al. (1998) Proc. Natl. Acad. Sci. USA 95:7888-7892).

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The discovery of new membrane transport proteins and the polynucleotides encoding them satisfies a need in the art by providing new compositions which are useful in the diagnosis, prevention, and treatment of membrane transport disorders; immune/inflammatory disorders; and cell proliferative disorders including cancer.

SUMMARY OF THE INVENTION

The invention features substantially purified polypeptides, membrane transport proteins, referred to collectively as "MTRP" and individually as "MTRP-1," "MTRP-2," "MTRP-3," "MTRP-4," "MTRP-5," "MTRP-6," "MTRP-7," "MTRP-8," "MTRP-9," "MTRP-10," "MTRP-11," "MTRP-12," "MTRP-13," "MTRP-14," "MTRP-15," "MTRP-16," and "MTRP-17." In one aspect, the invention provides a substantially purified polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-17 and fragments thereof. The invention also includes a polypeptide comprising an amino acid sequence that differs by one or more conservative amino acid substitutions from an amino acid sequence selected from the group consisting of SEQ ID NO:1-17.

The invention further provides a substantially purified variant having at least 90% amino acid identity to at least one of the amino acid sequences selected from the group consisting of SEQ ID NO:1-17 and fragments thereof. The invention also provides an isolated and purified polynucleotide encoding the polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-17 and fragments thereof. The invention also includes an isolated and purified polynucleotide variant having at least 90% polynucleotide sequence identity to the polynucleotide encoding the polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-17 and fragments thereof.

Additionally, the invention provides an isolated and purified polynucleotide which hybridizes under stringent conditions to the polynucleotide encoding the polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-17 and fragments thereof. The

invention also provides an isolated and purified polynucleotide having a sequence which is complementary to the polynucleotide encoding the polypeptide comprising the amino acid sequence selected from the group consisting of SEQ ID NO:1-17 and fragments thereof.

The invention also provides a method for detecting a polynucleotide in a sample containing nucleic acids, the method comprising the steps of: (a) hybridizing the complement of the polynucleotide sequence to at least one of the polynucleotides of the sample, thereby forming a hybridization complex; and (b) detecting the hybridization complex, wherein the presence of the hybridization complex correlates with the presence of a polynucleotide in the sample. In one aspect, the method further comprises amplifying the polynucleotide prior to hybridization.

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The invention also provides an isolated and purified polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:18-34 and fragments thereof. The invention further provides an isolated and purified polynucleotide variant having at least 90% polynucleotide sequence identity to the polynucleotide sequence selected from the group consisting of SEQ ID NO:18-34 and fragments thereof. The invention also provides an isolated and purified polynucleotide having a sequence which is complementary to the polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:18-34 and fragments thereof.

The invention further provides an expression vector containing at least a fragment of the polynucleotide encoding the polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-17. In another aspect, the expression vector is contained within a host cell.

The invention also provides a method for producing a polypeptide, the method comprising the steps of: (a) culturing the host cell containing an expression vector containing a polynucleotide of the invention under conditions suitable for the expression of the polypeptide; and (b) recovering the polypeptide from the host cell culture.

The invention also provides a pharmaceutical composition comprising a substantially purified polypeptide having the amino acid sequence selected from the group consisting of SEQ ID NO:1-17 and fragments thereof, in conjunction with a suitable pharmaceutical carrier.

The invention further includes a purified antibody which binds to a polypeptide selected from the group consisting of SEQ ID NO:1-17 and fragments thereof. The invention also provides a purified agonist and a purified antagonist to the polypeptide.

The invention also provides a method for treating or preventing a disorder associated with decreased expression or activity of MTRP, the method comprising administering to a subject in need of such treatment an effective amount of a pharmaceutical composition comprising a substantially

purified polypeptide having the amino acid sequence selected from the group consisting of SEQ ID NO:1-17 and fragments thereof, in conjunction with a suitable pharmaceutical carrier.

The invention also provides a method for treating or preventing a disorder associated with increased expression or activity of MTRP, the method comprising administering to a subject in need of such treatment an effective amount of an antagonist of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-17 and fragments thereof.

BRIEF DESCRIPTION OF THE FIGURES AND TABLES

Figures 1A, 1B, 1C, and 1D show the amino acid sequence alignment between MTRP-3

(Incyte Clone ID 1720440; SEQ ID NO:3) and mouse fatty acid transport protein (GI 2612939; SEQ ID NO:35), produced using the multisequence alignment program of LASERGENE software (DNASTAR, Madison WI).

Figures 2A, 2B, 2C, and 2D show the amino acid sequence alignment between MTRP-4 (Incyte Clone ID 2274290; SEQ ID NO:4) and <u>Schistosoma mansoni</u> SMDR1 (GI 425474; SEQ ID NO:36), produced using the multisequence alignment program of LASERGENE software (DNASTAR).

Figures 3A, 3B, 3C, and 3D show the amino acid sequence alignment between MTRP-5 (Incyte Clone ID 2740029; SEQ ID NO:5) and rat sodium-dependent multivitamin transporter (GI 3015617; SEQ ID NO:37), produced using the multisequence alignment program of LASERGENE software (DNASTAR).

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Table 1 shows polypeptide and nucleotide sequence identification numbers (SEQ ID NOs), clone identification numbers (clone IDs), cDNA libraries, and cDNA fragments used to assemble full-length sequences encoding MTRP.

Table 2 shows features of each polypeptide sequence, including potential motifs, homologous sequences, and methods, algorithms, and searchable databases used for analysis of MTRP.

Table 3 shows the tissue-specific expression patterns of each nucleic acid sequence as determined by northern analysis; diseases, disorders, or conditions associated with these tissues; and the vector into which each cDNA was cloned.

Table 4 describes the tissues used to construct the cDNA libraries from which cDNA clones encoding MTRP were isolated.

Table 5 shows the tools, programs, and algorithms used to analyze MTRP, along with applicable descriptions, references, and threshold parameters.

DESCRIPTION OF THE INVENTION

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Before the present proteins, nucleotide sequences, and methods are described, it is understood that this invention is not limited to the particular machines, materials and methods described, as these may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims.

It must be noted that as used herein and in the appended claims, the singular forms "a," "an," and "the" include plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to "a host cell" includes a plurality of such host cells, and a reference to "an antibody" is a reference to one or more antibodies and equivalents thereof known to those skilled in the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meanings as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any machines, materials, and methods similar or equivalent to those described herein can be used to practice or test the present invention, the preferred machines, materials and methods are now described. All publications mentioned herein are cited for the purpose of describing and disclosing the cell lines, protocols, reagents and vectors which are reported in the publications and which might be used in connection with the invention. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of prior invention.

DEFINITIONS

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"MTRP" refers to the amino acid sequences of substantially purified MTRP obtained from any species, particularly a mammalian species, including bovine, ovine, porcine, murine, equine, and human, and from any source, whether natural, synthetic, semi-synthetic, or recombinant.

The term "agonist" refers to a molecule which intensifies or mimics the biological activity of MTRP. Agonists may include proteins, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of MTRP either by directly interacting with MTRP or by acting on components of the biological pathway in which MTRP participates.

An "allelic variant" is an alternative form of the gene encoding MTRP. Allelic variants may result from at least one mutation in the nucleic acid sequence and may result in altered mRNAs or in polypeptides whose structure or function may or may not be altered. A gene may have none, or many allelic variants of its naturally occurring form. Common mutational changes which give rise to allelic variants are generally ascribed to natural deletions, additions, or substitutions of nucleotides. Each of these types of changes may occur alone, or in combination with the others, one or more times in a given sequence.

"Altered" nucleic acid sequences encoding MTRP include those sequences with deletions,

insertions, or substitutions of different nucleotides, resulting in a polypeptide the same as MTRP or a polypeptide with at least one functional characteristic of MTRP. Included within this definition are polymorphisms which may or may not be readily detectable using a particular oligonucleotide probe of the polynucleotide encoding MTRP, and improper or unexpected hybridization to allelic variants, with a locus other than the normal chromosomal locus for the polynucleotide sequence encoding MTRP. The encoded protein may also be "altered," and may contain deletions, insertions, or substitutions of amino acid residues which produce a silent change and result in a functionally equivalent MTRP. Deliberate amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues, as long as the biological or immunological activity of MTRP is retained. For example, negatively charged amino acids may include aspartic acid and glutamic acid, and positively charged amino acids may include lysine and arginine. Amino acids with uncharged polar side chains having similar hydrophilicity values may include: asparagine and glutamine; and serine and threonine. Amino acids with uncharged side chains having similar hydrophilicity values may include: leucine, isoleucine, and valine; glycine and alanine; and phenylalanine and tyrosine.

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The terms "amino acid" and "amino acid sequence" refer to an oligopeptide, peptide, polypeptide, or protein sequence, or a fragment of any of these, and to naturally occurring or synthetic molecules. Where "amino acid sequence" is recited to refer to an amino acid sequence of a naturally occurring protein molecule, "amino acid sequence" and like terms are not meant to limit the amino acid sequence to the complete native amino acid sequence associated with the recited protein molecule.

"Amplification" relates to the production of additional copies of a nucleic acid sequence.

Amplification is generally carried out using polymerase chain reaction (PCR) technologies well known in the art.

The term "antagonist" refers to a molecule which inhibits or attenuates the biological activity of MTRP. Antagonists may include proteins such as antibodies, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of MTRP either by directly interacting with MTRP or by acting on components of the biological pathway in which MTRP participates.

The term "antibody" refers to intact immunoglobulin molecules as well as to fragments thereof, such as Fab, F(ab')₂, and Fv fragments, which are capable of binding an epitopic determinant. Antibodies that bind MTRP polypeptides can be prepared using intact polypeptides or using fragments containing small peptides of interest as the immunizing antigen. The polypeptide or oligopeptide used to immunize an animal (e.g., a mouse, a rat, or a rabbit) can be derived from the

translation of RNA, or synthesized chemically, and can be conjugated to a carrier protein if desired. Commonly used carriers that are chemically coupled to peptides include bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin (KLH). The coupled peptide is then used to immunize the animal.

The term "antigenic determinant" refers to that region of a molecule (i.e., an epitope) that makes contact with a particular antibody. When a protein or a fragment of a protein is used to immunize a host animal, numerous regions of the protein may induce the production of antibodies which bind specifically to antigenic determinants (particular regions or three-dimensional structures on the protein). An antigenic determinant may compete with the intact antigen (i.e., the immunogen used to elicit the immune response) for binding to an antibody.

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The term "antisense" refers to any composition containing a nucleic acid sequence which is complementary to the "sense" strand of a specific nucleic acid sequence. Antisense molecules may be produced by any method including synthesis or transcription. Once introduced into a cell, the complementary nucleotides combine with natural sequences produced by the cell to form duplexes and to block either transcription or translation. The designation "negative" or "minus" can refer to the antisense strand, and the designation "positive" or "plus" can refer to the sense strand.

The term "biologically active" refers to a protein having structural, regulatory, or biochemical functions of a naturally occurring molecule. Likewise, "immunologically active" refers to the capability of the natural, recombinant, or synthetic MTRP, or of any oligopeptide thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies.

The terms "complementary" and "complementarity" refer to the natural binding of polynucleotides by base pairing. For example, the sequence "5' A-G-T 3'" bonds to the complementary sequence "3' T-C-A 5'." Complementarity between two single-stranded molecules may be "partial," such that only some of the nucleic acids bind, or it may be "complete," such that total complementarity exists between the single stranded molecules. The degree of complementarity between nucleic acid strands has significant effects on the efficiency and strength of the hybridization between the nucleic acid strands. This is of particular importance in amplification reactions, which depend upon binding between nucleic acid strands, and in the design and use of peptide nucleic acid (PNA) molecules.

A "composition comprising a given polynucleotide sequence" and a "composition comprising a given amino acid sequence" refer broadly to any composition containing the given polynucleotide or amino acid sequence. The composition may comprise a dry formulation or an aqueous solution.

Compositions comprising polynucleotide sequences encoding MTRP or fragments of MTRP may be employed as hybridization probes. The probes may be stored in freeze-dried form and may be

associated with a stabilizing agent such as a carbohydrate. In hybridizations, the probe may be deployed in an aqueous solution containing salts (e.g., NaCl), detergents (e.g., sodium dodecyl sulfate; SDS), and other components (e.g., Denhardt's solution, dry milk, salmon sperm DNA, etc.).

"Consensus sequence" refers to a nucleic acid sequence which has been resequenced to resolve uncalled bases, extended using the XL-PCR kit (Perkin-Elmer, Norwalk CT) in the 5' and/or the 3' direction, and resequenced, or which has been assembled from the overlapping sequences of one or more Incyte Clones and, in some cases, one or more public domain ESTs, using a computer program for fragment assembly, such as the GELVIEW fragment assembly system (GCG, Madison WI). Some sequences have been both extended and assembled to produce the consensus sequence.

"Conservative amino acid substitutions" are those substitutions that, when made, least interfere with the properties of the original protein, i.e., the structure and especially the function of the protein is conserved and not significantly changed by such substitutions. The table below shows amino acids which may be substituted for an original amino acid in a protein and which are regarded as conservative amino acid substitutions.

15	Original Residue	Conservative Substitution
•	Ala	Gly, Ser
	Arg	His, Lys
	Asn	Asp, Gln, His
•	Asp	Asn, Glu
20	Cys	Ala, Ser
	Gln	Asn, Glu, His
	Glu	Asp, Gln, His
	Gly	Ala
	His	Asn, Arg, Gln, Glu
25	Ile	Leu, Val
	Leu	Ile, Val
	Lys	Arg, Gln, Glu
	Met	Leu, Ile
	Phe	His, Met, Leu, Trp, Tyr
30	Ser	Cys, Thr
	Thr	Ser, Val
	Trp	Phe, Tyr
	Tyr	His, Phe, Trp
	Val	Ile, Leu, Thr
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Conservative amino acid substitutions generally maintain (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a beta sheet or alpha helical conformation, (b) the charge or hydrophobicity of the molecule at the site of the substitution, and/or (c) the bulk of the side chain.

A "deletion" refers to a change in the amino acid or nucleotide sequence that results in the absence of one or more amino acid residues or nucleotides.

The term "derivative" refers to the chemical modification of a polypeptide sequence, or a polynucleotide sequence. Chemical modifications of a polynucleotide sequence can include, for example, replacement of hydrogen by an alkyl, acyl, hydroxyl, or amino group. A derivative polynucleotide encodes a polypeptide which retains at least one biological or immunological function of the natural molecule. A derivative polypeptide is one modified by glycosylation, pegylation, or any similar process that retains at least one biological or immunological function of the polypeptide from which it was derived.

A "fragment" is a unique portion of MTRP or the polynucleotide encoding MTRP which is identical in sequence to but shorter in length than the parent sequence. A fragment may comprise up to the entire length of the defined sequence, minus one nucleotide/amino acid residue. For example, a fragment may comprise from 5 to 1000 contiguous nucleotides or amino acid residues. A fragment used as a probe, primer, antigen, therapeutic molecule, or for other purposes, may be at least 5, 10, 15, 20, 25, 30, 40, 50, 60, 75, 100, 150, 250 or at least 500 contiguous nucleotides or amino acid residues in length. Fragments may be preferentially selected from certain regions of a molecule. For example, a polypeptide fragment may comprise a certain length of contiguous amino acids selected from the first 250 or 500 amino acids (or first 25% or 50% of a polypeptide) as shown in a certain defined sequence. Clearly these lengths are exemplary, and any length that is supported by the specification, including the Sequence Listing, tables, and figures, may be encompassed by the present embodiments.

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A fragment of SEQ ID NO:18-34 comprises a region of unique polynucleotide sequence that specifically identifies SEQ ID NO:18-34, for example, as distinct from any other sequence in the same genome. A fragment of SEQ ID NO:18-34 is useful, for example, in hybridization and amplification technologies and in analogous methods that distinguish SEQ ID NO:18-34 from related polynucleotide sequences. The precise length of a fragment of SEQ ID NO:18-34 and the region of SEQ ID NO:18-34 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A fragment of SEQ ID NO:1-17 is encoded by a fragment of SEQ ID NO:18-34. A fragment of SEQ ID NO:1-17 comprises a region of unique amino acid sequence that specifically identifies SEQ ID NO:1-17. For example, a fragment of SEQ ID NO:1-17 is useful as an immunogenic peptide for the development of antibodies that specifically recognize SEQ ID NO:1-17. The precise length of a fragment of SEQ ID NO:1-17 and the region of SEQ ID NO:1-17 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

The term "similarity" refers to a degree of complementarity. There may be partial similarity or complete similarity. The word "identity" may substitute for the word "similarity." A partially

complementary sequence that at least partially inhibits an identical sequence from hybridizing to a target nucleic acid is referred to as "substantially similar." The inhibition of hybridization of the completely complementary sequence to the target sequence may be examined using a hybridization assay (Southern or northern blot, solution hybridization, and the like) under conditions of reduced stringency. A substantially similar sequence or hybridization probe will compete for and inhibit the binding of a completely similar (identical) sequence to the target sequence under conditions of reduced stringency. This is not to say that conditions of reduced stringency are such that non-specific binding is permitted, as reduced stringency conditions require that the binding of two sequences to one another be a specific (i.e., a selective) interaction. The absence of non-specific binding may be tested by the use of a second target sequence which lacks even a partial degree of complementarity (e.g., less than about 30% similarity or identity). In the absence of non-specific binding, the substantially similar sequence or probe will not hybridize to the second non-complementary target sequence.

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The phrases "percent identity" and "% identity," as applied to polynucleotide sequences, refer to the percentage of residue matches between at least two polynucleotide sequences aligned using a standardized algorithm. Such an algorithm may insert, in a standardized and reproducible way, gaps in the sequences being compared in order to optimize alignment between two sequences, and therefore achieve a more meaningful comparison of the two sequences.

Percent identity between polynucleotide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program. This program is part of the LASERGENE software package, a suite of molecular biological analysis programs (DNASTAR, Madison WI). CLUSTAL V is described in Higgins, D.G. and P.M. Sharp (1989) CABIOS 5:151-153 and in Higgins, D.G. et al. (1992) CABIOS 8:189-191. For pairwise alignments of polynucleotide sequences, the default parameters are set as follows: Ktuple=2, gap penalty=5, window=4, and "diagonals saved"=4. The "weighted" residue weight table is selected as the default. Percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polynucleotide sequence pairs.

Alternatively, a suite of commonly used and freely available sequence comparison algorithms is provided by the National Center for Biotechnology Information (NCBI) Basic Local Alignment Search Tool (BLAST) (Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410), which is available from several sources, including the NCBI, Bethesda, MD, and on the Internet at http://www.ncbi.nlm.nih.gov/BLAST/. The BLAST software suite includes various sequence analysis programs including "blastn," that is used to align a known polynucleotide sequence with other polynucleotide sequences from a variety of databases. Also available is a tool called "BLAST 2"

Sequences" that is used for direct pairwise comparison of two nucleotide sequences. "BLAST 2 Sequences" can be accessed and used interactively at http://www.ncbi.nlm.nih.gov/gorf/bl2.html. The "BLAST 2 Sequences" tool can be used for both blastn and blastp (discussed below). BLAST programs are commonly used with gap and other parameters set to default settings. For example, to compare two nucleotide sequences, one may use blastn with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Reward for match: 1

Penalty for mismatch: -2

Open Gap: 5 and Extension Gap: 2 penalties

Gap x drop-off: 50

Expect: 10

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Word Size: 11

Filter: on

Percent identity may be measured over the length of an entire defined sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined sequence, for instance, a fragment of at least 20, at least 30, at least 40, at least 50, at least 70, at least 100, or at least 200 contiguous nucleotides. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures, or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

Nucleic acid sequences that do not show a high degree of identity may nevertheless encode similar amino acid sequences due to the degeneracy of the genetic code. It is understood that changes in a nucleic acid sequence can be made using this degeneracy to produce multiple nucleic acid sequences that all encode substantially the same protein.

The phrases "percent identity" and "% identity," as applied to polypeptide sequences, refer to the percentage of residue matches between at least two polypeptide sequences aligned using a standardized algorithm. Methods of polypeptide sequence alignment are well-known. Some alignment methods take into account conservative amino acid substitutions. Such conservative substitutions, explained in more detail above, generally preserve the hydrophobicity and acidity at the site of substitution, thus preserving the structure (and therefore function) of the polypeptide.

Percent identity between polypeptide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program (described and referenced above). For pairwise alignments of

polypeptide sequences using CLUSTAL V, the default parameters are set as follows: Ktuple=1, gap penalty=3, window=5, and "diagonals saved"=5. The PAM250 matrix is selected as the default residue weight table. As with polynucleotide alignments, the percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polypeptide sequence pairs.

Alternatively the NCBI BLAST software suite may be used. For example, for a pairwise comparison of two polypeptide sequences, one may use the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) with blastp set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Open Gap: 11 and Extension Gap: 1 penalties

Gap x drop-off: 50

Expect: 10

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Word Size: 3

Filter: on

Percent identity may be measured over the length of an entire defined polypeptide sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined polypeptide sequence, for instance, a fragment of at least 15, at least 20, at least 30, at least 40, at least 50, at least 70 or at least 150 contiguous residues. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

"Human artificial chromosomes" (HACs) are linear microchromosomes which may contain DNA sequences of about 6 kb to 10 Mb in size, and which contain all of the elements required for stable mitotic chromosome segregation and maintenance.

The term "humanized antibody" refers to antibody molecules in which the amino acid sequence in the non-antigen binding regions has been altered so that the antibody more closely resembles a human antibody, and still retains its original binding ability.

"Hybridization" refers to the process by which a polynucleotide strand anneals with a complementary strand through base pairing under defined hybridization conditions. Specific hybridization is an indication that two nucleic acid sequences share a high degree of identity. Specific hybridization complexes form under permissive annealing conditions and remain hybridized after the "washing" step(s). The washing step(s) is particularly important in determining the stringency of the hybridization process, with more stringent conditions allowing less non-specific binding, i.e., binding between pairs of nucleic acid strands that are not perfectly matched. Permissive conditions for annealing of nucleic acid sequences are routinely determinable by one of ordinary skill in the art and

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may be consistent among hybridization experiments, whereas wash conditions may be varied among experiments to achieve the desired stringency, and therefore hybridization specificity. Permissive annealing conditions occur, for example, at 68° C in the presence of about $6 \times SSC$, about 1% (w/v) SDS, and about $100 \mu g/ml$ denatured salmon sperm DNA.

Generally, stringency of hybridization is expressed, in part, with reference to the temperature under which the wash step is carried out. Generally, such wash temperatures are selected to be about 5°C to 20°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. An equation for calculating T_m and conditions for nucleic acid hybridization are well known and can be found in Sambrook et al., 1989, Molecular Cloning: A Laboratory Manual, 2nd ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; specifically see volume 2, chapter 9.

High stringency conditions for hybridization between polynucleotides of the present invention include wash conditions of 68°C in the presence of about 0.2 x SSC and about 0.1% SDS, for 1 hour. Alternatively, temperatures of about 65°C, 60°C, 55°C, or 42°C may be used. SSC concentration may be varied from about 0.1 to 2 x SSC, with SDS being present at about 0.1%. Typically, blocking reagents are used to block non-specific hybridization. Such blocking reagents include, for instance, denatured salmon sperm DNA at about 100-200 µg/ml. Organic solvent, such as formamide at a concentration of about 35-50% v/v, may also be used under particular circumstances, such as for RNA:DNA hybridizations. Useful variations on these wash conditions will be readily apparent to those of ordinary skill in the art. Hybridization, particularly under high stringency conditions, may be suggestive of evolutionary similarity between the nucleotides. Such similarity is strongly indicative of a similar role for the nucleotides and their encoded polypeptides.

The term "hybridization complex" refers to a complex formed between two nucleic acid sequences by virtue of the formation of hydrogen bonds between complementary bases. A hybridization complex may be formed in solution (e.g., $C_0 t$ or $R_0 t$ analysis) or formed between one nucleic acid sequence present in solution and another nucleic acid sequence immobilized on a solid support (e.g., paper, membranes, filters, chips, pins or glass slides, or any other appropriate substrate to which cells or their nucleic acids have been fixed).

The words "insertion" and "addition" refer to changes in an amino acid or nucleotide sequence resulting in the addition of one or more amino acid residues or nucleotides, respectively.

"Immune response" can refer to conditions associated with inflammation, trauma, immune disorders, or infectious or genetic disease, etc. These conditions can be characterized by expression of various factors, e.g., cytokines, chemokines, and other signaling molecules, which may affect

cellular and systemic defense systems.

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The term "microarray" refers to an arrangement of distinct polynucleotides on a substrate.

The terms "element" and "array element" in a microarray context, refer to hybridizable polynucleotides arranged on the surface of a substrate.

The term "modulate" refers to a change in the activity of MTRP. For example, modulation may cause an increase or a decrease in protein activity, binding characteristics, or any other biological, functional, or immunological properties of MTRP.

The phrases "nucleic acid" and "nucleic acid sequence" refer to a nucleotide, oligonucleotide, polynucleotide, or any fragment thereof. These phrases also refer to DNA or RNA of genomic or synthetic origin which may be single-stranded or double-stranded and may represent the sense or the antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material.

"Operably linked" refers to the situation in which a first nucleic acid sequence is placed in a functional relationship with the second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding sequence. Generally, operably linked DNA sequences may be in close proximity or contiguous and, where necessary to join two protein coding regions, in the same reading frame.

"Peptide nucleic acid" (PNA) refers to an antisense molecule or anti-gene agent which comprises an oligonucleotide of at least about 5 nucleotides in length linked to a peptide backbone of amino acid residues ending in lysine. The terminal lysine confers solubility to the composition.

PNAs preferentially bind complementary single stranded DNA or RNA and stop transcript elongation, and may be pegylated to extend their lifespan in the cell.

"Probe" refers to nucleic acid sequences encoding MTRP, their complements, or fragments thereof, which are used to detect identical, allelic or related nucleic acid sequences. Probes are isolated oligonucleotides or polynucleotides attached to a detectable label or reporter molecule. Typical labels include radioactive isotopes, ligands, chemiluminescent agents, and enzymes. "Primers" are short nucleic acids, usually DNA oligonucleotides, which may be annealed to a target polynucleotide by complementary base-pairing. The primer may then be extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification (and

Probes and primers as used in the present invention typically comprise at least 15 contiguous nucleotides of a known sequence. In order to enhance specificity, longer probes and primers may also be employed, such as probes and primers that comprise at least 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, or at least 150 consecutive nucleotides of the disclosed nucleic acid sequences. Probes and primers may be considerably longer than these examples, and it is understood that any length supported by the

identification) of a nucleic acid sequence, e.g., by the polymerase chain reaction (PCR).

specification, including the tables, figures, and Sequence Listing, may be used.

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Methods for preparing and using probes and primers are described in the references, for example Sambrook et al., 1989, Molecular Cloning: A Laboratory Manual, 2nd ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; Ausubel et al., 1987, Current Protocols in Molecular Biology, Greene Publ. Assoc. & Wiley-Intersciences, New York NY; Innis et al., 1990, PCR Protocols, A Guide to Methods and Applications, Academic Press, San Diego CA. PCR primer pairs can be derived from a known sequence, for example, by using computer programs intended for that purpose such as Primer (Version 0.5, 1991, Whitehead Institute for Biomedical Research, Cambridge MA).

Oligonucleotides for use as primers are selected using software known in the art for such purpose. For example, OLIGO 4.06 software is useful for the selection of PCR primer pairs of up to 100 nucleotides each, and for the analysis of oligonucleotides and larger polynucleotides of up to 5,000 nucleotides from an input polynucleotide sequence of up to 32 kilobases. Similar primer selection programs have incorporated additional features for expanded capabilities. For example, the PrimOU primer selection program (available to the public from the Genome Center at University of Texas South West Medical Center, Dallas TX) is capable of choosing specific primers from megabase sequences and is thus useful for designing primers on a genome-wide scope. The Primer3 primer selection program (available to the public from the Whitehead Institute/MIT Center for Genome Research, Cambridge MA) allows the user to input a "mispriming library," in which sequences to avoid as primer binding sites are user-specified. Primer3 is useful, in particular, for the selection of oligonucleotides for microarrays. (The source code for the latter two primer selection programs may also be obtained from their respective sources and modified to meet the user's specific needs.) The PrimeGen program (available to the public from the UK Human Genome Mapping Project Resource Centre, Cambridge UK) designs primers based on multiple sequence alignments, thereby allowing selection of primers that hybridize to either the most conserved or least conserved regions of aligned nucleic acid sequences. Hence, this program is useful for identification of both unique and conserved oligonucleotides and polynucleotide fragments. The oligonucleotides and polynucleotide fragments identified by any of the above selection methods are useful in hybridization technologies, for example, as PCR or sequencing primers, microarray elements, or specific probes to identify fully or partially complementary polynucleotides in a sample of nucleic acids. Methods of oligonucleotide selection are not limited to those described above.

A "recombinant nucleic acid" is a sequence that is not naturally occurring or has a sequence that is made by an artificial combination of two or more otherwise separated segments of sequence. This artificial combination is often accomplished by chemical synthesis or, more commonly, by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques

such as those described in Sambrook, <u>supra</u>. The term recombinant includes nucleic acids that have been altered solely by addition, substitution, or deletion of a portion of the nucleic acid. Frequently, a recombinant nucleic acid may include a nucleic acid sequence operably linked to a promoter sequence. Such a recombinant nucleic acid may be part of a vector that is used, for example, to transform a cell.

Alternatively, such recombinant nucleic acids may be part of a viral vector, e.g., based on a vaccinia virus, that could be use to vaccinate a mammal wherein the recombinant nucleic acid is expressed, inducing a protective immunological response in the mammal.

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The term "sample" is used in its broadest sense. A sample suspected of containing nucleic acids encoding MTRP, or fragments thereof, or MTRP itself, may comprise a bodily fluid; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; a cell; genomic DNA, RNA, or cDNA, in solution or bound to a substrate; a tissue; a tissue print; etc.

The terms "specific binding" and "specifically binding" refer to that interaction between a protein or peptide and an agonist, an antibody, an antagonist, a small molecule, or any natural or synthetic binding composition. The interaction is dependent upon the presence of a particular structure of the protein, e.g., the antigenic determinant or epitope, recognized by the binding molecule. For example, if an antibody is specific for epitope "A," the presence of a polypeptide containing the epitope A, or the presence of free unlabeled A, in a reaction containing free labeled A and the antibody will reduce the amount of labeled A that binds to the antibody.

The term "substantially purified" refers to nucleic acid or amino acid sequences that are removed from their natural environment and are isolated or separated, and are at least about 60% free, preferably about 75% free, and most preferably about 90% free from other components with which they are naturally associated.

A "substitution" refers to the replacement of one or more amino acids or nucleotides by different amino acids or nucleotides, respectively.

"Substrate" refers to any suitable rigid or semi-rigid support including membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, plates, polymers, microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which polynucleotides or polypeptides are bound.

"Transformation" describes a process by which exogenous DNA enters and changes a recipient cell. Transformation may occur under natural or artificial conditions according to various methods well known in the art, and may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method for transformation is selected based on the type of host cell being transformed and may include, but is not limited to, viral infection,

electroporation, heat shock, lipofection, and particle bombardment. The term "transformed" cells includes stably transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome, as well as transiently transformed cells which express the inserted DNA or RNA for limited periods of time.

A "variant" of a particular nucleic acid sequence is defined as a nucleic acid sequence having at least 40% sequence identity to the particular nucleic acid sequence over a certain length of one of the nucleic acid sequences using blastn with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of nucleic acids may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 95% or at least 98% or greater sequence identity over a certain defined length. A variant may be described as, for example, an "allelic" (as defined above), "splice," "species," or "polymorphic" variant. A splice variant may have significant identity to a reference molecule, but will generally have a greater or lesser number of polynucleotides due to alternate splicing of exons during mRNA processing. The corresponding polypeptide may possess additional functional domains or lack domains that are present in the reference molecule. Species variants are polynucleotide sequences that vary from one species to another. The resulting polypeptides generally will have significant amino acid identity relative to each other. A polymorphic variant is a variation in the polynucleotide sequence of a particular gene between individuals of a given species. Polymorphic variants also may encompass "single nucleotide polymorphisms" (SNPs) in which the polynucleotide sequence varies by one nucleotide base. The presence of SNPs may be indicative of, for example, a certain population, a disease state, or a propensity for a disease state.

A "variant" of a particular polypeptide sequence is defined as a polypeptide sequence having at least 40% sequence identity to the particular polypeptide sequence over a certain length of one of the polypeptide sequences using blastp with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of polypeptides may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 95%, or at least 98% or greater sequence identity over a certain defined length of one of the polypeptides.

THE INVENTION

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The invention is based on the discovery of new human membrane transport proteins (MTRP), the polynucleotides encoding MTRP, and the use of these compositions for the diagnosis, treatment, or prevention of membrane transport disorders; immune/inflammatory disorders; and cell proliferative disorders including cancer.

Table 1 lists the Incyte clones used to assemble full length nucleotide sequences encoding MTRP. Columns 1 and 2 show the sequence identification numbers (SEQ ID NOs) of the polypeptide

and nucleotide sequences, respectively. Column 3 shows the clone IDs of the Incyte clones in which nucleic acids encoding each MTRP were identified, and column 4 shows the cDNA libraries from which these clones were isolated. Column 5 shows Incyte clones and their corresponding cDNA libraries. Clones for which cDNA libraries are not indicated were derived from pooled cDNA libraries. The Incyte clones in column 5 were used to assemble the consensus nucleotide sequence of each MTRP and are useful as fragments in hybridization technologies.

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The columns of Table 2 show various properties of each of the polypeptides of the invention: column 1 references the SEQ ID NO; column 2 shows the number of amino acid residues in each polypeptide; column 3 shows potential phosphorylation sites; column 4 shows potential glycosylation sites; column 5 shows the amino acid residues comprising signature sequences and motifs; column 6 shows homologous sequences as identified by BLAST analysis; and column 7 shows analytical methods and in some cases, searchable databases to which the analytical methods were applied. The methods of column 7 were used to characterize each polypeptide through sequence homology and protein motifs.

As shown in Figures 1A, 1B, 1C, and 1D, MTRP-3 has chemical and structural similarity with mouse fatty acid transport protein (FATP; Gl 2612939; SEQ ID NO:35). In particular, MTRP-3 and FATP share 65% identity. As shown in Figures 2A, 2B, 2C, and 2D, MTRP-4 has chemical and structural similarity with Schistosoma mansoni ATP-binding cassette family protein, SMDR-1 (GI 425474; SEQ ID NO:36). In particular, MTRP-4 and SMDR-1 share 38% identity. As shown in Figures 3A, 3B, 3C, and 3D, MTRP-5 has chemical and structural similarity with rat sodium-dependent multivitamin transporter (SMVT; Gl 3015617; SEQ ID NO:37). In particular, MTRP-5 and SMVT share 82% identity.

The columns of Table 3 show the tissue-specificity and diseases, disorders, or conditions associated with nucleotide sequences encoding MTRP. The first column of Table 3 lists the nucleotide SEQ ID NOs. Column 2 lists tissue categories which express MTRP as a fraction of total tissues expressing MTRP. Column 3 lists diseases, disorders, or conditions associated with those tissues expressing MTRP as a fraction of total tissues expressing MTRP. Column 4 lists the vectors used to subclone each cDNA library.

Of particular note are the expression patterns of SEQ ID NO:30 and SEQ ID NO:31. SEQ ID NO:30 is expressed in only five libraries, of which at least four (80%) are associated with cell proliferation and at least one (20%) with inflammation. Two (40%) of the five libraries are associated with cardiovascular tissue, and one each (20%) with gastrointestinal, nervous, and reproductive tissues. SEQ ID NO:31 is expressed in only four libraries, of which at least three (75%) are associated with cell proliferation and at least two (50%) with inflammation or the immune response.

Two (50%) of the four libraries are associated with hematopoietic/immune tissue, and one each (25%) with cardiovascular and reproductive tissues.

The following fragments of the nucleotide sequences encoding MTRP are useful, for example, in hybridization or amplification technologies to identify SEQ ID NO:18-34 and to distinguish between SEQ ID NO:18-34 and related polynucleotide sequences. The useful fragments include the fragment of SEQ ID NO:18 from about nucleotide 110 to about nucleotide 154; the fragment of SEQ ID NO:19 from about nucleotide 759 to about nucleotide 839; the fragment of SEQ ID NO:20 from about nucleotide 1531 to about nucleotide 1578; the fragment of SEQ ID NO:21 from about nucleotide 538 to about nucleotide 597; the fragment of SEQ ID NO:22 from about nucleotide 2241 to about nucleotide 2294; the fragment of SEQ ID NO:23 from about nucleotide 116 to about nucleotide 145; the fragment of SEQ ID NO:24 from about nucleotide 60 to about nucleotide 89; the fragment of SEQ ID NO:25 from about nucleotide 160 to about nucleotide 189; the fragment of SEQ ID NO:26 from about nucleotide 763 to about nucleotide 792; the fragment of SEQ ID NO:27 from about nucleotide 43 to about nucleotide 72; the fragment of SEQ ID NO:28 from about nucleotide 361 to about nucleotide 405; the fragment of SEQ ID NO:29 from about nucleotide 35 to about nucleotide 79; the fragment of SEQ ID NO:30 from about nucleotide 206 to about nucleotide 250; the fragment of SEQ ID NO:31 from about nucleotide 71 to about nucleotide 115; the fragment of SEQ ID NO:32 from about nucleotide 161 to about nucleotide 205; the fragment of SEQ ID NO:33 from about nucleotide 364 to about nucleotide 408; and the fragment of SEQ ID NO:34 from about nucleotide 18 to about nucleotide 62. The polypeptides encoded by the specified fragments of SEQ ID NO:20-30 and SEQ ID NO:32-34 are useful, for example, as immunogenic peptides.

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The columns of Table 4 show descriptions of the tissues used to construct the cDNA libraries from which cDNA clones encoding MTRP were isolated. Column 1 references the nucleotide SEQ ID NOs, column 2 shows the cDNA libraries from which these clones were isolated, and column 3 shows the tissue origins and other descriptive information relevant to the cDNA libraries in column 2.

The invention also encompasses MTRP variants. A preferred MTRP variant is one which has at least about 80%, or alternatively at least about 90%, or even at least about 95% amino acid sequence identity to the MTRP amino acid sequence, and which contains at least one functional or structural characteristic of MTRP.

The invention also encompasses polynucleotides which encode MTRP. In a particular embodiment, the invention encompasses a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:18-34, which encodes MTRP.

The invention also encompasses a variant of a polynucleotide sequence encoding MTRP. In particular, such a variant polynucleotide sequence will have at least about 75%, or alternatively at

least about 85%, or even at least about 95% polynucleotide sequence identity to the polynucleotide sequence encoding MTRP. A particular aspect of the invention encompasses a variant of a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:18-34 which has at least about 75%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to a nucleic acid sequence selected from the group consisting of SEQ ID NO:18-34. Any one of the polynucleotide variants described above can encode an amino acid sequence which contains at least one functional or structural characteristic of MTRP.

It will be appreciated by those skilled in the art that as a result of the degeneracy of the genetic code, a multitude of polynucleotide sequences encoding MTRP, some bearing minimal similarity to the polynucleotide sequences of any known and naturally occurring gene, may be produced. Thus, the invention contemplates each and every possible variation of polynucleotide sequence that could be made by selecting combinations based on possible codon choices. These combinations are made in accordance with the standard triplet genetic code as applied to the polynucleotide sequence of naturally occurring MTRP, and all such variations are to be considered as being specifically disclosed.

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Although nucleotide sequences which encode MTRP and its variants are generally capable of hybridizing to the nucleotide sequence of the naturally occurring MTRP under appropriately selected conditions of stringency, it may be advantageous to produce nucleotide sequences encoding MTRP or its derivatives possessing a substantially different codon usage, e.g., inclusion of non-naturally occurring codons. Codons may be selected to increase the rate at which expression of the peptide occurs in a particular prokaryotic or eukaryotic host in accordance with the frequency with which particular codons are utilized by the host. Other reasons for substantially altering the nucleotide sequence encoding MTRP and its derivatives without altering the encoded amino acid sequences include the production of RNA transcripts having more desirable properties, such as a greater half-life, than transcripts produced from the naturally occurring sequence.

The invention also encompasses production of DNA sequences which encode MTRP and MTRP derivatives, or fragments thereof, entirely by synthetic chemistry. After production, the synthetic sequence may be inserted into any of the many available expression vectors and cell systems using reagents well known in the art. Moreover, synthetic chemistry may be used to introduce mutations into a sequence encoding MTRP or any fragment thereof.

Also encompassed by the invention are polynucleotide sequences that are capable of hybridizing to the claimed polynucleotide sequences, and, in particular, to those shown in SEQ ID NO:18-34 and fragments thereof under various conditions of stringency. (See, e.g., Wahl, G.M. and S.L. Berger (1987) Methods Enzymol. 152:399-407; Kimmel, A.R. (1987) Methods Enzymol.

152:507-511.) Hybridization conditions, including annealing and wash conditions, are described in "Definitions."

Methods for DNA sequencing are well known in the art and may be used to practice any of the embodiments of the invention. The methods may employ such enzymes as the Klenow fragment of DNA polymerase I, SEQUENASE (US Biochemical, Cleveland OH), Taq polymerase (Perkin-Elmer), thermostable T7 polymerase (Amersham Pharmacia Biotech, Piscataway NJ), or combinations of polymerases and proofreading exonucleases such as those found in the ELONGASE amplification system (Life Technologies, Gaithersburg MD). Preferably, sequence preparation is automated with machines such as the MICROLAB 2200 liquid transfer system (Hamilton, Reno NV), PTC200 thermal cycler (MJ Research, Watertown MA) and ABI CATALYST 800 thermal cycler (Perkin-Elmer). Sequencing is then carried out using either the ABI 373 or 377 DNA sequencing system (Perkin-Elmer), the MEGABACE 1000 DNA sequencing system (Molecular Dynamics, Sunnyvale CA), or other systems known in the art. The resulting sequences are analyzed using a variety of algorithms which are well known in the art. (See, e.g., Ausubel, F.M. (1997) Short Protocols in Molecular Biology, John Wiley & Sons, New York NY, unit 7.7; Meyers, R.A. (1995) Molecular Biology and Biotechnology, Wiley VCH, New York NY, pp. 856-853.)

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The nucleic acid sequences encoding MTRP may be extended utilizing a partial nucleotide sequence and employing various PCR-based methods known in the art to detect upstream sequences, such as promoters and regulatory elements. For example, one method which may be employed, restriction-site PCR, uses universal and nested primers to amplify unknown sequence from genomic DNA within a cloning vector. (See, e.g., Sarkar, G. (1993) PCR Methods Applic. 2:318-322.) Another method, inverse PCR, uses primers that extend in divergent directions to amplify unknown sequence from a circularized template. The template is derived from restriction fragments comprising a known genomic locus and surrounding sequences. (See, e.g., Triglia, T. et al. (1988) Nucleic Acids Res. 16:8186.) A third method, capture PCR, involves PCR amplification of DNA fragments adjacent to known sequences in human and yeast artificial chromosome DNA. (See, e.g., Lagerstrom, M. et al. (1991) PCR Methods Applic. 1:111-119.) In this method, multiple restriction enzyme digestions and ligations may be used to insert an engineered double-stranded sequence into a region of unknown sequence before performing PCR. Other methods which may be used to retrieve unknown sequences are known in the art. (See, e.g., Parker, J.D. et al. (1991) Nucleic Acids Res. 19:3055-3060). Additionally, one may use PCR, nested primers, and PROMOTERFINDER libraries (Clontech, Palo Alto CA) to walk genomic DNA. This procedure avoids the need to screen libraries and is useful in finding intron/exon junctions. For all PCR-based methods, primers may be designed using commercially available software, such as OLIGO 4.06 Primer Analysis software (National

Biosciences, Plymouth MN) or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the template at temperatures of about 68°C to 72°C.

When screening for full-length cDNAs, it is preferable to use libraries that have been size-selected to include larger cDNAs. In addition, random-primed libraries, which often include sequences containing the 5' regions of genes, are preferable for situations in which an oligo d(T) library does not yield a full-length cDNA. Genomic libraries may be useful for extension of sequence into 5' non-transcribed regulatory regions.

Capillary electrophoresis systems which are commercially available may be used to analyze the size or confirm the nucleotide sequence of sequencing or PCR products. In particular, capillary sequencing may employ flowable polymers for electrophoretic separation, four different nucleotide-specific, laser-stimulated fluorescent dyes, and a charge coupled device camera for detection of the emitted wavelengths. Output/light intensity may be converted to electrical signal using appropriate software (e.g., GENOTYPER and SEQUENCE NAVIGATOR, Perkin-Elmer), and the entire process from loading of samples to computer analysis and electronic data display may be computer controlled. Capillary electrophoresis is especially preferable for sequencing small DNA fragments which may be present in limited amounts in a particular sample.

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In another embodiment of the invention, polynucleotide sequences or fragments thereof which encode MTRP may be cloned in recombinant DNA molecules that direct expression of MTRP, or fragments or functional equivalents thereof, in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences which encode substantially the same or a functionally equivalent amino acid sequence may be produced and used to express MTRP.

The nucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter MTRP-encoding sequences for a variety of purposes including, but not limited to, modification of the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. For example, oligonucleotide-mediated site-directed mutagenesis may be used to introduce mutations that create new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, and so forth.

In another embodiment, sequences encoding MTRP may be synthesized, in whole or in part, using chemical methods well known in the art. (See, e.g., Caruthers, M.H. et al. (1980) Nucleic Acids Symp. Ser. 7:215-223; and Horn, T. et al. (1980) Nucleic Acids Symp. Ser. 7:225-232.)

Alternatively, MTRP itself or a fragment thereof may be synthesized using chemical methods. For example, peptide synthesis can be performed using various solid-phase techniques. (See, e.g.,

Roberge, J.Y. et al. (1995) Science 269:202-204.) Automated synthesis may be achieved using the ABI 431A peptide synthesizer (Perkin-Elmer). Additionally, the amino acid sequence of MTRP, or any part thereof, may be altered during direct synthesis and/or combined with sequences from other proteins, or any part thereof, to produce a variant polypeptide.

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The peptide may be substantially purified by preparative high performance liquid chromatography. (See, e.g., Chiez, R.M. and F.Z. Regnier (1990) Methods Enzymol. 182:392-421.) The composition of the synthetic peptides may be confirmed by amino acid analysis or by sequencing. (See, e.g., Creighton, T. (1984) Proteins, Structures and Molecular Properties, WH Freeman, New York NY.)

In order to express a biologically active MTRP, the nucleotide sequences encoding MTRP or derivatives thereof may be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for transcriptional and translational control of the inserted coding sequence in a suitable host. These elements include regulatory sequences, such as enhancers, constitutive and inducible promoters, and 5' and 3' untranslated regions in the vector and in polynucleotide sequences encoding MTRP. Such elements may vary in their strength and specificity. Specific initiation signals may also be used to achieve more efficient translation of sequences encoding MTRP. Such signals include the ATG initiation codon and adjacent sequences, e.g. the Kozak sequence. In cases where sequences encoding MTRP and its initiation codon and upstream regulatory sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals including an in-frame ATG initiation codon should be provided by the vector. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate for the particular host cell system used. (See, e.g., Scharf, D. et al. (1994) Results Probl. Cell Differ. 20:125-162.)

Methods which are well known to those skilled in the art may be used to construct expression vectors containing sequences encoding MTRP and appropriate transcriptional and translational control elements. These methods include in vitro recombinant DNA techniques, synthetic techniques, and in vivo genetic recombination. (See, e.g., Sambrook, J. et al. (1989) Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Press, Plainview NY, ch. 4, 8, and 16-17; Ausubel, F.M. et al. (1995) Current Protocols in Molecular Biology, John Wiley & Sons, New York NY, ch. 9, 13, and 16.)

A variety of expression vector/host systems may be utilized to contain and express sequences encoding MTRP. These include, but are not limited to, microorganisms such as bacteria transformed

with recombinant bacteriophage, plasmid, or cosmid DNA.expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with viral expression vectors (e.g., baculovirus); plant cell systems transformed with viral expression vectors (e.g., cauliflower mosaic virus, CaMV, or tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or animal cell systems. The invention is not limited by the host cell employed.

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In bacterial systems, a number of cloning and expression vectors may be selected depending upon the use intended for polynucleotide sequences encoding MTRP. For example, routine cloning, subcloning, and propagation of polynucleotide sequences encoding MTRP can be achieved using a multifunctional E. coli vector such as PBLUESCRIPT (Stratagene, La Jolla CA) or PSPORT1 plasmid (Life Technologies). Ligation of sequences encoding MTRP into the vector's multiple cloning site disrupts the *lacZ* gene, allowing a colorimetric screening procedure for identification of transformed bacteria containing recombinant molecules. In addition, these vectors may be useful for in vitro transcription, dideoxy sequencing, single strand rescue with helper phage, and creation of nested deletions in the cloned sequence. (See, e.g., Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509.) When large quantities of MTRP are needed, e.g. for the production of antibodies, vectors which direct high level expression of MTRP may be used. For example, vectors containing the strong, inducible T5 or T7 bacteriophage promoter may be used.

Yeast expression systems may be used for production of MTRP. A number of vectors containing constitutive or inducible promoters, such as alpha factor, alcohol oxidase, and PGH promoters, may be used in the yeast <u>Saccharomyces cerevisiae</u> or <u>Pichia pastoris</u>. In addition, such vectors direct either the secretion or intracellular retention of expressed proteins and enable integration of foreign sequences into the host genome for stable propagation. (See, e.g., Ausubel, 1995, <u>supra</u>; Bitter, G.A. et al. (1987) Methods Enzymol. 153:516-544; and Scorer, C.A. et al. (1994) Bio/Technology 12:181-184.)

Plant systems may also be used for expression of MTRP. Transcription of sequences encoding MTRP may be driven viral promoters, e.g., the 35S and 19S promoters of CaMV used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) EMBO J. 6:307-311). Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used. (See, e.g., Coruzzi, G. et al. (1984) EMBO J. 3:1671-1680; Broglie, R. et al. (1984) Science 224:838-843; and Winter, J. et al. (1991) Results Probl. Cell Differ. 17:85-105.) These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. (See, e.g., The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196.)

In mammalian cells, a number of viral-based expression systems may be utilized. In cases

where an adenovirus is used as an expression vector, sequences encoding MTRP may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain infective virus which expresses MTRP in host cells. (See, e.g., Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659.) In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells. SV40 or EBV-based vectors may also be used for high-level protein expression.

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Human artificial chromosomes (HACs) may also be employed to deliver larger fragments of DNA than can be contained in and expressed from a plasmid. HACs of about 6 kb to 10 Mb are constructed and delivered via conventional delivery methods (liposomes, polycationic amino polymers, or vesicles) for therapeutic purposes. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.)

For long term production of recombinant proteins in mammalian systems, stable expression of MTRP in cell lines is preferred. For example, sequences encoding MTRP can be transformed into cell lines using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for about 1 to 2 days in enriched media before being switched to selective media. The purpose of the selectable marker is to confer resistance to a selective agent, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clones of stably transformed cells may be propagated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase and adenine phosphoribosyltransferase genes, for use in *tk* and *apr* cells, respectively. (See, e.g., Wigler, M. et al. (1977) Cell 11:223-232; Lowy, l. et al. (1980) Cell 22:817-823.) Also, antimetabolite, antibiotic, or herbicide resistance can be used as the basis for selection. For example, *dhfr* confers resistance to methotrexate; *neo* confers resistance to the aminoglycosides neomycin and G-418; and *als* and *pat* confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively. (See, e.g., Wigler, M. et al. (1980) Proc. Natl. Acad. Sci. USA 77:3567-3570; Colbere-Garapin, F. et al. (1981) J. Mol. Biol. 150:1-14.) Additional selectable genes have been described, e.g., *trpB* and *hisD*, which alter cellular requirements for metabolites. (See, e.g., Hartman, S.C. and R.C. Mulligan (1988) Proc. Natl. Acad. Sci. USA 85:8047-8051.) Visible markers, e.g., anthocyanins, green fluorescent proteins (GFP; Clontech), β glucuronidase and its substrate β-glucuronide, or luciferase and its substrate luciferin may be used. These markers can be used not only to identify transformants, but also to

quantify the amount of transient or stable protein expression attributable to a specific vector system. (See, e.g., Rhodes, C.A. (1995) Methods Mol. Biol. 55:121-131.)

Although the presence/absence of marker gene expression suggests that the gene of interest is also present, the presence and expression of the gene may need to be confirmed. For example, if the sequence encoding MTRP is inserted within a marker gene sequence, transformed cells containing sequences encoding MTRP can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a sequence encoding MTRP under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

In general, host cells that contain the nucleic acid sequence encoding MTRP and that express MTRP may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations, PCR amplification, and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein sequences.

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Immunological methods for detecting and measuring the expression of MTRP using either specific polyclonal or monoclonal antibodies are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on MTRP is preferred, but a competitive binding assay may be employed. These and other assays are well known in the art. (See, e.g., Hampton, R. et al. (1990) Serological Methods, a Laboratory Manual, APS Press, St. Paul MN, Sect. IV; Coligan, J.E. et al. (1997) Current Protocols in Immunology, Greene Pub. Associates and Wiley-Interscience, New York NY; and Pound, J.D. (1998) Immunochemical Protocols, Humana Press, Totowa NJ.)

A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides encoding MTRP include oligolabeling, nick translation, end-labeling, or PCR amplification using a labeled nucleotide. Alternatively, the sequences encoding MTRP, or any fragments thereof, may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits, such as those provided by Amersham Pharmacia Biotech, Promega (Madison WI), and US Biochemical. Suitable reporter molecules or labels which may be used for

ease of detection include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents, as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

Host cells transformed with nucleotide sequences encoding MTRP may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a transformed cell may be secreted or retained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides which encode MTRP may be designed to contain signal sequences which direct secretion of MTRP through a prokaryotic or eukaryotic cell membrane.

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In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" or "pro" form of the protein may also be used to specify protein targeting, folding, and/or activity. Different host cells which have specific cellular machinery and characteristic mechanisms for post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and Wl38) are available from the American Type Culture Collection (ATCC, Manassas VA) and may be chosen to ensure the correct modification and processing of the foreign protein.

In another embodiment of the invention, natural, modified, or recombinant nucleic acid sequences encoding MTRP may be ligated to a heterologous sequence resulting in translation of a fusion protein in any of the aforementioned host systems. For example, a chimeric MTRP protein containing a heterologous moiety that can be recognized by a commercially available antibody may facilitate the screening of peptide libraries for inhibitors of MTRP activity. Heterologous protein and peptide moieties may also facilitate purification of fusion proteins using commercially available affinity matrices. Such moieties include, but are not limited to, glutathione S-transferase (GST), maltose binding protein (MBP), thioredoxin (Trx), calmodulin binding peptide (CBP), 6-His, FLAG, c-myc, and hemagglutinin (HA). GST, MBP, Trx, CBP, and 6-His enable purification of their cognate fusion proteins on immobilized glutathione, maltose, phenylarsine oxide, calmodulin, and metalchelate resins, respectively. FLAG, c-myc, and hemagglutinin (HA) enable immunoaffinity purification of fusion proteins using commercially available monoclonal and polyclonal antibodies that specifically recognize these epitope tags. A fusion protein may also be engineered to contain a proteolytic cleavage site located between the MTRP encoding sequence and the heterologous protein sequence, so that MTRP may be cleaved away from the heterologous moiety following purification. Methods for fusion protein expression and purification are discussed in Ausubel (1995, supra, ch. 10). A variety of commercially available kits may also be used to facilitate expression and purification of

fusion proteins.

In a further embodiment of the invention, synthesis of radiolabeled MTRP may be achieved in vitro using the TNT rabbit reticulocyte lysate or wheat germ extract system (Promega). These systems couple transcription and translation of protein-coding sequences operably associated with the T7, T3, or SP6 promoters. Translation takes place in the presence of a radiolabeled amino acid precursor, for example, ³⁵S-methionine.

Fragments of MTRP may be produced not only by recombinant means, but also by direct peptide synthesis using solid-phase techniques. (See, e.g., Creighton, supra, pp. 55-60.) Protein synthesis may be performed by manual techniques or by automation. Automated synthesis may be achieved, for example, using the ABI 431A peptide synthesizer (Perkin-Elmer). Various fragments of MTRP may be synthesized separately and then combined to produce the full length molecule.

THERAPEUTICS

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Chemical and structural similarity, e.g., in the context of sequences and motifs, exists between regions of MTRP and membrane transport proteins, including amino acid transporters, ABC transporters, nucleotide-sugar transporters, transmembrane carrier proteins, and ATP-dependent transporter proteins. In addition, the expression of MTRP is closely associated with nervous, reproductive, and gastrointestinal tissues; cancer and other cell proliferative conditions; and with inflammation and the immune response. Therefore, MTRP appears to play a role in membrane transport disorders; immune/inflammatory disorders; and cell proliferative disorders including cancer. In the treatment of disorders associated with increased MTRP expression or activity, it is desirable to decrease the expression or activity of MTRP. In the treatment of disorders associated with decreased MTRP expression or activity, it is desirable to increase the expression or activity of MTRP.

Therefore, in one embodiment, MTRP or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of MTRP. Examples of such disorders include, but are not limited to, a membrane transport disorder such as cystinuria, dibasicaminoaciduria, hypercystinuria, lysinuria, hartnup disease, tryptophan malabsorption, methionine malabsorption, histidinuria, iminoglycinuria, dicarboxylicaminoaciduria, cystinosis, renal glycosuria, glucose-galactose malabsorption, familial hypercholesterolemia, hypouricemia, familial hypophophatemic rickets, congenital chloridorrhea, cystic fibrosis, familial goiter, distal renal tubular acidosis, Menkes' disease, lethal diarrhea, nephrogenic diabetes insipidus, juvenile pernicious anemia, folate malabsorption, adrenoleukodystrophy, hereditary myoglobinuria, Zellweger syndrome, hyperinsulinemic hypoglycemia, akinesia, amyotrophic lateral sclerosis, ataxia telangiectasia, cystic fibrosis, Becker's muscular dystrophy, Bell's palsy, Charcot-Marie Tooth disease, diabetes mellitus, diabetes insipidus,

diabetic neuropathy, Duchenne muscular dystrophy, hyperkalemic periodic paralysis, normokalemic periodic paralysis, Parkinson's disease, malignant hyperthermia, multidrug resistance, myasthenia gravis, myotonic dystrophy, catatonia, tardive dyskinesia, dystonias, peripheral neuropathy, cerebral neoplasms, and prostate cancer; a cardiac disorder associated with transport such as angina, bradyarrythmia, tachyarrythmia, hypertension, Long QT syndrome, myocarditis, cardiomyopathy, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, thyrotoxic myopathy, ethanol myopathy, dermatomyositis, inclusion body myositis, infectious myositis, and polymyositis; a neurological disorder associated with transport such as Alzheimer's disease, amnesia, bipolar disorder, dementia, depression, epilepsy, Tourette's disorder, paranoid psychoses, and schizophrenia; and an other disorder associated with transport such as neurofibromatosis, postherpetic neuralgia, trigeminal neuropathy, sarcoidosis, sickle cell anemia, Wilson's disease, cataracts, infertility, pulmonary artery stenosis, sensorineural autosomal deafness, hyperglycemia, hypoglycemia, Grave's disease, goiter, Cushing's disease, Addison's disease, glucose-galactose malabsorption syndrome, and hypercholesterolemia; an immune/inflammatory disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyenodocrinopathycandidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation; a viral, bacterial, fungal, parasitic, protozoal, or helminthic infection; and trauma; a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia; and a cancer including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma; and, in particular, a cancer of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis,

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thymus, thyroid, and uterus.

In another embodiment, a vector capable of expressing MTRP or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of MTRP including, but not limited to, those described above.

In a further embodiment, a pharmaceutical composition comprising a substantially purified MTRP in conjunction with a suitable pharmaceutical carrier may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of MTRP including, but not limited to, those provided above.

In still another embodiment, an agonist which modulates the activity of MTRP may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of MTRP including, but not limited to, those listed above.

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In a further embodiment, an antagonist of MTRP may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of MTRP. Examples of such disorders include, but are not limited to, those membrane transport disorders; immune/inflammatory disorders; and cell proliferative disorders including cancer described above. In one aspect, an antibody which specifically binds MTRP may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for bringing a pharmaceutical agent to cells or tissues which express MTRP.

In an additional embodiment, a vector expressing the complement of the polynucleotide encoding MTRP may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of MTRP including, but not limited to, those described above.

In other embodiments, any of the proteins, antagonists, antibodies, agonists, complementary sequences, or vectors of the invention may be administered in combination with other appropriate therapeutic agents. Selection of the appropriate agents for use in combination therapy may be made by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents may act synergistically to effect the treatment or prevention of the various disorders described above. Using this approach, one may be able to achieve therapeutic efficacy with lower dosages of each agent, thus reducing the potential for adverse side effects.

An antagonist of MTRP may be produced using methods which are generally known in the art. In particular, purified MTRP may be used to produce antibodies or to screen libraries of pharmaceutical agents to identify those which specifically bind MTRP. Antibodies to MTRP may also be generated using methods that are well known in the art. Such antibodies may include, but are not limited to, polyclonal, monoclonal, chimeric, and single chain antibodies, Fab fragments, and fragments produced by a Fab expression library. Neutralizing antibodies (i.e., those which inhibit dimer formation) are generally preferred for therapeutic use.

For the production of antibodies, various hosts including goats, rabbits, rats, mice, humans, and others may be immunized by injection with MTRP or with any fragment or oligopeptide thereof which has immunogenic properties. Depending on the host species, various adjuvants may be used to increase immunological response. Such adjuvants include, but are not limited to, Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, KLH, and dinitrophenol. Among adjuvants used in humans, BCG (bacilli Calmette-Guerin) and Corynebacterium parvum are especially preferable.

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It is preferred that the oligopeptides, peptides, or fragments used to induce antibodies to MTRP have an amino acid sequence consisting of at least about 5 amino acids, and generally will consist of at least about 10 amino acids. It is also preferable that these oligopeptides, peptides, or fragments are identical to a portion of the amino acid sequence of the natural protein and contain the entire amino acid sequence of a small, naturally occurring molecule. Short stretches of MTRP amino acids may be fused with those of another protein, such as KLH, and antibodies to the chimeric molecule may be produced.

Monoclonal antibodies to MTRP may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma technique. (See, e.g., Kohler, G. et al. (1975) Nature 256:495-497; Kozbor, D. et al. (1985) J. Immunol. Methods 81:31-42; Cote, R.J. et al. (1983) Proc. Natl. Acad. Sci. USA 80:2026-2030; and Cole, S.P. et al. (1984) Mol. Cell Biol. 62:109-120.)

In addition, techniques developed for the production of "chimeric antibodies," such as the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used. (See, e.g., Morrison, S.L. et al. (1984) Proc. Natl. Acad. Sci. USA 81:6851-6855; Neuberger, M.S. et al. (1984) Nature 312:604-608; and Takeda, S. et al. (1985) Nature 314:452-454.) Alternatively, techniques described for the production of single chain antibodies may be adapted, using methods known in the art, to produce MTRP-specific single chain antibodies. Antibodies with related specificity, but of distinct idiotypic composition, may be generated by chain shuffling from random combinatorial immunoglobulin libraries. (See, e.g., Burton, D.R. (1991) Proc. Natl. Acad. Sci. USA 88:10134-10137.)

Antibodies may also be produced by inducing in vivo production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as disclosed in the literature. (See, e.g., Orlandi, R. et al. (1989) Proc. Natl. Acad. Sci. USA 86:3833-3837; Winter, G. et al. (1991) Nature 349:293-299.)

Antibody fragments which contain specific binding sites for MTRP may also be generated.

For example, such fragments include, but are not limited to, $F(ab')_2$ fragments produced by pepsin digestion of the antibody molecule and Fab fragments generated by reducing the disulfide bridges of the $F(ab')_2$ fragments. Alternatively, Fab expression libraries may be constructed to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity. (See, e.g., Huse, W.D. et al. (1989) Science 246:1275-1281.)

Various immunoassays may be used for screening to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays using either polyclonal or monoclonal antibodies with established specificities are well known in the art. Such immunoassays typically involve the measurement of complex formation between MTRP and its specific antibody. A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering MTRP epitopes is generally used, but a competitive binding assay may also be employed (Pound, supra).

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Various methods such as Scatchard analysis in conjunction with radioimmunoassay techniques may be used to assess the affinity of antibodies for MTRP. Affinity is expressed as an association constant, K_a , which is defined as the molar concentration of MTRP-antibody complex divided by the molar concentrations of free antigen and free antibody under equilibrium conditions. The K_a determined for a preparation of polyclonal antibodies, which are heterogeneous in their affinities for multiple MTRP epitopes, represents the average affinity, or avidity, of the antibodies for MTRP. The K_a determined for a preparation of monoclonal antibodies, which are monospecific for a particular MTRP epitope, represents a true measure of affinity. High-affinity antibody preparations with K_a ranging from about 10° to 10^{12} L/mole are preferred for use in immunoassays in which the MTRP-antibody complex must withstand rigorous manipulations. Low-affinity antibody preparations with K_a ranging from about 10° to 10^7 L/mole are preferred for use in immunopurification and similar procedures which ultimately require dissociation of MTRP, preferably in active form, from the antibody (Catty, D. (1988) Antibodies. Volume 1: A Practical Approach, IRL Press, Washington, DC; Liddell, J.E. and Cryer, A. (1991) A Practical Guide to Monoclonal Antibodies, John Wiley & Sons, New York NY).

The titer and avidity of polyclonal antibody preparations may be further evaluated to determine the quality and suitability of such preparations for certain downstream applications. For example, a polyclonal antibody preparation containing at least 1-2 mg specific antibody/ml, preferably 5-10 mg specific antibody/ml, is generally employed in procedures requiring precipitation of MTRP-antibody complexes. Procedures for evaluating antibody specificity, titer, and avidity, and guidelines for antibody quality and usage in various applications, are generally available. (See, e.g., Catty, supra, and Coligan et al. supra.)

In another embodiment of the invention, the polynucleotides encoding MTRP, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, the complement of the polynucleotide encoding MTRP may be used in situations in which it would be desirable to block the transcription of the mRNA. In particular, cells may be transformed with sequences complementary to polynucleotides encoding MTRP. Thus, complementary molecules or fragments may be used to modulate MTRP activity, or to achieve regulation of gene function. Such technology is now well known in the art, and sense or antisense oligonucleotides or larger fragments can be designed from various locations along the coding or control regions of sequences encoding MTRP.

Expression vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from various bacterial plasmids, may be used for delivery of nucleotide sequences to the targeted organ, tissue, or cell population. Methods which are well known to those skilled in the art can be used to construct vectors to express nucleic acid sequences complementary to the polynucleotides encoding MTRP. (See, e.g., Sambrook, supra; Ausubel, 1995, supra.)

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Genes encoding MTRP can be turned off by transforming a cell or tissue with expression vectors which express high levels of a polynucleotide, or fragment thereof, encoding MTRP. Such constructs may be used to introduce untranslatable sense or antisense sequences into a cell. Even in the absence of integration into the DNA, such vectors may continue to transcribe RNA molecules until they are disabled by endogenous nucleases. Transient expression may last for a month or more with a non-replicating vector, and may last even longer if appropriate replication elements are part of the vector system.

As mentioned above, modifications of gene expression can be obtained by designing complementary sequences or antisense molecules (DNA, RNA, or PNA) to the control, 5', or regulatory regions of the gene encoding MTRP. Oligonucleotides derived from the transcription initiation site, e.g., between about positions -10 and +10 from the start site, may be employed. Similarly, inhibition can be achieved using triple helix base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the ability of the double helix to open sufficiently for the binding of polymerases, transcription factors, or regulatory molecules. Recent therapeutic advances using triplex DNA have been described in the literature. (See, e.g., Gee, J.E. et al. (1994) in Huber, B.E. and B.I. Carr, Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177.) A complementary sequence or antisense molecule may also be designed to block translation of mRNA by preventing the transcript from binding to ribosomes.

Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme

molecule to complementary target RNA, followed by endonucleolytic cleavage. For example, engineered hammerhead motif ribozyme molecules may specifically and efficiently catalyze endonucleolytic cleavage of sequences encoding MTRP.

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Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the target molecule for ribozyme cleavage sites, including the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides, corresponding to the region of the target gene containing the cleavage site, may be evaluated for secondary structural features which may render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing accessibility to hybridization with complementary oligonucleotides using ribonuclease protection assays.

Complementary ribonucleic acid molecules and ribozymes of the invention may be prepared by any method known in the art for the synthesis of nucleic acid molecules. These include techniques for chemically synthesizing oligonucleotides such as solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by in vitro and in vivo transcription of DNA sequences encoding MTRP. Such DNA sequences may be incorporated into a wide variety of vectors with suitable RNA polymerase promoters such as T7 or SP6. Alternatively, these cDNA constructs that synthesize complementary RNA, constitutively or inducibly, can be introduced into cell lines, cells, or tissues.

RNA molecules may be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends of the molecule, or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. This concept is inherent in the production of PNAs and can be extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, as well as acetyl-, methyl-, thio-, and similarly modified forms of adenine, cytidine, guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases.

Many methods for introducing vectors into cells or tissues are available and equally suitable for use in vivo, in vitro, and ex vivo. For ex vivo therapy, vectors may be introduced into stem cells taken from the patient and clonally propagated for autologous transplant back into that same patient. Delivery by transfection, by liposome injections, or by polycationic amino polymers may be achieved using methods which are well known in the art. (See, e.g., Goldman, C.K. et al. (1997) Nat. Biotechnol. 15:462-466.)

Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as humans, dogs, cats, cows, horses, rabbits, and monkeys.

An additional embodiment of the invention relates to the administration of a pharmaceutical or sterile composition, in conjunction with a pharmaceutically acceptable carrier, for any of the therapeutic effects discussed above. Such pharmaceutical compositions may consist of MTRP, antibodies to MTRP, and mimetics, agonists, antagonists, or inhibitors of MTRP. The compositions may be administered alone or in combination with at least one other agent, such as a stabilizing compound, which may be administered in any sterile, biocompatible pharmaceutical carrier including, but not limited to, saline, buffered saline, dextrose, and water. The compositions may be administered to a patient alone, or in combination with other agents, drugs, or hormones.

The pharmaceutical compositions utilized in this invention may be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical, sublingual, or rectal means.

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In addition to the active ingredients, these pharmaceutical compositions may contain suitable pharmaceutically-acceptable carriers comprising excipients and auxiliaries which facilitate processing of the active compounds into preparations which can be used pharmaceutically. Further details on techniques for formulation and administration may be found in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing, Easton PA).

Pharmaceutical compositions for oral administration can be formulated using pharmaceutically acceptable carriers well known in the art in dosages suitable for oral administration. Such carriers enable the pharmaceutical compositions to be formulated as tablets, pills, dragees, capsules, liquids, gels, syrups, slurries, suspensions, and the like, for ingestion by the patient.

Pharmaceutical preparations for oral use can be obtained through combining active compounds with solid excipient and processing the resultant mixture of granules (optionally, after grinding) to obtain tablets or dragee cores. Suitable auxiliaries can be added, if desired. Suitable excipients include carbohydrate or protein fillers, such as sugars, including lactose, sucrose, mannitol, and sorbitol; starch from corn, wheat, rice, potato, or other plants; cellulose, such as methyl cellulose, hydroxypropylmethyl-cellulose, or sodium carboxymethylcellulose; gums, including arabic and tragacanth; and proteins, such as gelatin and collagen. If desired, disintegrating or solubilizing agents may be added, such as the cross-linked polyvinyl pyrrolidone, agar, and alginic acid or a salt thereof, such as sodium alginate.

Dragee cores may be used in conjunction with suitable coatings, such as concentrated sugar solutions, which may also contain gum arabic, tale, polyvinylpyrrolidone, carbopol gel, polyethylene glycol, and/or titanium dioxide, lacquer solutions, and suitable organic solvents or solvent mixtures. Dyestuffs or pigments may be added to the tablets or dragee coatings for product identification or to

characterize the quantity of active compound, i.e., dosage.

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Pharmaceutical preparations which can be used orally include push-fit capsules made of gelatin, as well as soft, sealed capsules made of gelatin and a coating, such as glycerol or sorbitol. Push-fit capsules can contain active ingredients mixed with fillers or binders, such as lactose or starches, lubricants, such as talc or magnesium stearate, and, optionally, stabilizers. In soft capsules, the active compounds may be dissolved or suspended in suitable liquids, such as fatty oils, liquid, or liquid polyethylene glycol with or without stabilizers.

Pharmaceutical formulations suitable for parenteral administration may be formulated in aqueous solutions, preferably in physiologically compatible buffers such as Hanks' solution, Ringer's solution, or physiologically buffered saline. Aqueous injection suspensions may contain substances which increase the viscosity of the suspension, such as sodium carboxymethyl cellulose, sorbitol, or dextran. Additionally, suspensions of the active compounds may be prepared as appropriate oily injection suspensions. Suitable lipophilic solvents or vehicles include fatty oils, such as sesame oil, or synthetic fatty acid esters, such as ethyl oleate, triglycerides, or liposomes. Non-lipid polycationic amino polymers may also be used for delivery. Optionally, the suspension may also contain suitable stabilizers or agents to increase the solubility of the compounds and allow for the preparation of highly concentrated solutions.

For topical or nasal administration, penetrants appropriate to the particular barrier to be permeated are used in the formulation. Such penetrants are generally known in the art.

The pharmaceutical compositions of the present invention may be manufactured in a manner that is known in the art, e.g., by means of conventional mixing, dissolving, granulating, dragee-making, levigating, emulsifying, encapsulating, entrapping, or lyophilizing processes.

The pharmaceutical composition may be provided as a salt and can be formed with many acids, including but not limited to, hydrochloric, sulfuric, acetic, lactic, tartaric, malic, and succinic acids. Salts tend to be more soluble in aqueous or other protonic solvents than are the corresponding free base forms. In other cases, the preparation may be a lyophilized powder which may contain any or all of the following: 1 mM to 50 mM histidine, 0.1% to 2% sucrose, and 2% to 7% mannitol, at a pH range of 4.5 to 5.5, that is combined with buffer prior to use.

After pharmaceutical compositions have been prepared, they can be placed in an appropriate container and labeled for treatment of an indicated condition. For administration of MTRP, such labeling would include amount, frequency, and method of administration.

Pharmaceutical compositions suitable for use in the invention include compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose. The determination of an effective dose is well within the capability of those skilled in the art.

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., of neoplastic cells, or in animal models such as mice, rats, rabbits, dogs, or pigs. An animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of active ingredient, for example MTRP or fragments thereof, antibodies of MTRP, and agonists, antagonists or inhibitors of MTRP, which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity may be determined by standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the ED₅₀ (the dose therapeutically effective in 50% of the population) or LD₅₀ (the dose lethal to 50% of the population) statistics. The dose ratio of toxic to therapeutic effects is the therapeutic index, which can be expressed as the LD₅₀/ED₅₀ ratio. Pharmaceutical compositions which exhibit large therapeutic indices are preferred. The data obtained from cell culture assays and animal studies are used to formulate a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that includes the ED₅₀ with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, the sensitivity of the patient, and the route of administration.

The exact dosage will be determined by the practitioner, in light of factors related to the subject requiring treatment. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Factors which may be taken into account include the severity of the disease state, the general health of the subject, the age, weight, and gender of the subject, time and frequency of administration, drug combination(s), reaction sensitivities, and response to therapy. Long-acting pharmaceutical compositions may be administered every 3 to 4 days, every week, or biweekly depending on the half-life and clearance rate of the particular formulation.

Normal dosage amounts may vary from about $0.1~\mu g$ to $100,000~\mu g$, up to a total dose of about 1 gram, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

DIAGNOSTICS

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In another embodiment, antibodies which specifically bind MTRP may be used for the diagnosis of disorders characterized by expression of MTRP, or in assays to monitor patients being

treated with MTRP or agonists, antagonists, or inhibitors of MTRP. Antibodies useful for diagnostic purposes may be prepared in the same manner as described above for therapeutics. Diagnostic assays for MTRP include methods which utilize the antibody and a label to detect MTRP in human body fluids or in extracts of cells or tissues. The antibodies may be used with or without modification, and may be labeled by covalent or non-covalent attachment of a reporter molecule. A wide variety of reporter molecules, several of which are described above, are known in the art and may be used.

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A variety of protocols for measuring MTRP, including ELISAs, RIAs, and FACS, are known in the art and provide a basis for diagnosing altered or abnormal levels of MTRP expression. Normal or standard values for MTRP expression are established by combining body fluids or cell extracts taken from normal mammalian subjects, for example, human subjects, with antibody to MTRP under conditions suitable for complex formation. The amount of standard complex formation may be quantitated by various methods, such as photometric means. Quantities of MTRP expressed in subject, control, and disease samples from biopsied tissues are compared with the standard values. Deviation between standard and subject values establishes the parameters for diagnosing disease.

In another embodiment of the invention, the polynucleotides encoding MTRP may be used for diagnostic purposes. The polynucleotides which may be used include oligonucleotide sequences, complementary RNA and DNA molecules, and PNAs. The polynucleotides may be used to detect and quantify gene expression in biopsied tissues in which expression of MTRP may be correlated with disease. The diagnostic assay may be used to determine absence, presence, and excess expression of MTRP, and to monitor regulation of MTRP levels during therapeutic intervention.

In one aspect, hybridization with PCR probes which are capable of detecting polynucleotide sequences, including genomic sequences, encoding MTRP or closely related molecules may be used to identify nucleic acid sequences which encode MTRP. The specificity of the probe, whether it is made from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a conserved motif, and the stringency of the hybridization or amplification will determine whether the probe identifies only naturally occurring sequences encoding MTRP, allelic variants, or related sequences.

Probes may also be used for the detection of related sequences, and may have at least 50% sequence identity to any of the MTRP encoding sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NO:18-34 or from genomic sequences including promoters, enhancers, and introns of the MTRP gene.

Means for producing specific hybridization probes for DNAs encoding MTRP include the cloning of polynucleotide sequences encoding MTRP or MTRP derivatives into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available, and may

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be used to synthesize RNA probes <u>in vitro</u> by means of the addition of the appropriate RNA polymerases and the appropriate labeled nucleotides. Hybridization probes may be labeled by a variety of reporter groups, for example, by radionuclides such as ³²P or ³⁵S, or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems, and the like.

Polynucleotide sequences encoding MTRP may be used for the diagnosis of disorders associated with expression of MTRP. Examples of such disorders include, but are not limited to, a membrane transport disorder such as cystinuria, dibasicaminoaciduria, hypercystinuria, lysinuria, hartnup disease, tryptophan malabsorption, methionine malabsorption, histidinuria, iminoglycinuria, dicarboxylicaminoaciduria, cystinosis, renal glycosuria, glucose-galactose malabsorption, familial hypercholesterolemia, hypouricemia, familial hypophophatemic rickets, congenital chloridorrhea, cystic fibrosis, familial goiter, distal renal tubular acidosis, Menkes' disease, lethal diarrhea, nephrogenic diabetes insipidus, juvenile pernicious anemia, folate malabsorption, adrenoleukodystrophy, hereditary myoglobinuria, Zellweger syndrome, hyperinsulinemic hypoglycemia, akinesia, amyotrophic lateral sclerosis, ataxia telangiectasia, cystic fibrosis, Becker's muscular dystrophy, Bell's palsy, Charcot-Marie Tooth disease, diabetes mellitus, diabetes insipidus, diabetic neuropathy, Duchenne muscular dystrophy, hyperkalemic periodic paralysis, normokalemic periodic paralysis, Parkinson's disease, malignant hyperthermia, multidrug resistance, myasthenia gravis, myotonic dystrophy, catatonia, tardive dyskinesia, dystonias, peripheral neuropathy, cerebral neoplasms, and prostate cancer; a cardiac disorder associated with transport such as angina, bradyarrythmia, tachyarrythmia, hypertension, Long QT syndrome, myocarditis, cardiomyopathy, nemaline myopathy, centronuclear myopathy, lipid myopathy, mitochondrial myopathy, thyrotoxic myopathy, ethanol myopathy, dermatomyositis, inclusion body myositis, infectious myositis, and polymyositis; a neurological disorder associated with transport such as Alzheimer's disease, amnesia, bipolar disorder, dementia, depression, epilepsy, Tourette's disorder, paranoid psychoses, and schizophrenia; and an other disorder associated with transport such as neurofibromatosis, postherpetic neuralgia, trigeminal neuropathy, sarcoidosis, sickle cell anemia, Wilson's disease, cataracts, infertility, pulmonary artery stenosis, sensorineural autosomal deafness, hyperglycemia, hypoglycemia, Grave's disease, goiter, Cushing's disease, Addison's disease, glucose-galactose malabsorption syndrome, and hypercholesterolemia; an immune/inflammatory disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyenodocrinopathycandidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia

with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation; a viral, bacterial, fungal, parasitic, protozoal, or helminthic infection; and trauma; a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia; and a cancer including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma; and, in particular, a cancer of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus. The polynucleotide sequences encoding MTRP may be used in Southern or northern analysis, dot blot, or other membrane-based technologies; in PCR technologies; in dipstick, pin, and multiformat ELISA-like assays; and in microarrays utilizing fluids or tissues from patients to detect altered MTRP expression. Such qualitative or quantitative methods are well known in the art.

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In a particular aspect, the nucleotide sequences encoding MTRP may be useful in assays that detect the presence of associated disorders, particularly those mentioned above. The nucleotide sequences encoding MTRP may be labeled by standard methods and added to a fluid or tissue sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantified and compared with a standard value. If the amount of signal in the patient sample is significantly altered in comparison to a control sample then the presence of altered levels of nucleotide sequences encoding MTRP in the sample indicates the presence of the associated disorder. Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or to monitor the treatment of an individual patient.

In order to provide a basis for the diagnosis of a disorder associated with expression of MTRP, a normal or standard profile for expression is established. This may be accomplished by combining body fluids or cell extracts taken from normal subjects, either animal or human, with a sequence, or a fragment thereof, encoding MTRP, under conditions suitable for hybridization or amplification. Standard hybridization may be quantified by comparing the values obtained from

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normal subjects with values from an experiment in which a known amount of a substantially purified polynucleotide is used. Standard values obtained in this manner may be compared with values obtained from samples from patients who are symptomatic for a disorder. Deviation from standard values is used to establish the presence of a disorder.

Once the presence of a disorder is established and a treatment protocol is initiated, hybridization assays may be repeated on a regular basis to determine if the level of expression in the patient begins to approximate that which is observed in the normal subject. The results obtained from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

With respect to cancer, the presence of an abnormal amount of transcript (either under- or overexpressed) in biopsied tissue from an individual may indicate a predisposition for the development of the disease, or may provide a means for detecting the disease prior to the appearance of actual clinical symptoms. A more definitive diagnosis of this type may allow health professionals to employ preventative measures or aggressive treatment earlier thereby preventing the development or further progression of the cancer.

Additional diagnostic uses for oligonucleotides designed from the sequences encoding MTRP may involve the use of PCR. These oligomers may be chemically synthesized, generated enzymatically, or produced in vitro. Oligomers will preferably contain a fragment of a polynucleotide encoding MTRP, or a fragment of a polynucleotide complementary to the polynucleotide encoding MTRP, and will be employed under optimized conditions for identification of a specific gene or condition. Oligomers may also be employed under less stringent conditions for detection or quantification of closely related DNA or RNA sequences.

Methods which may also be used to quantify the expression of MTRP include radiolabeling or biotinylating nucleotides, coamplification of a control nucleic acid, and interpolating results from standard curves. (See, e.g., Melby, P.C. et al. (1993) J. Immunol. Methods 159:235-244; Duplaa, C. et al. (1993) Anal. Biochem. 212:229-236.) The speed of quantitation of multiple samples may be accelerated by running the assay in a high-throughput format where the oligomer of interest is presented in various dilutions and a spectrophotometric or colorimetric response gives rapid quantitation.

In further embodiments, oligonucleotides or longer fragments derived from any of the polynucleotide sequences described herein may be used as targets in a microarray. The microarray can be used to monitor the expression level of large numbers of genes simultaneously and to identify genetic variants, mutations, and polymorphisms. This information may be used to determine gene function, to understand the genetic basis of a disorder, to diagnose a disorder, and to develop and

monitor the activities of therapeutic agents.

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Microarrays may be prepared, used, and analyzed using methods known in the art. (See, e.g., Brennan, T.M. et al. (1995) U.S. Patent No. 5,474,796; Schena, M. et al. (1996) Proc. Natl. Acad. Sci. USA 93:10614-10619; Baldeschweiler et al. (1995) PCT application WO95/251116; Shalon, D. et al. (1995) PCT application WO95/35505; Heller, R.A. et al. (1997) Proc. Natl. Acad. Sci. USA 94:2150-2155; and Heller, M.J. et al. (1997) U.S. Patent No. 5,605,662.)

In another embodiment of the invention, nucleic acid sequences encoding MTRP may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. The sequences may be mapped to a particular chromosome, to a specific region of a chromosome, or to artificial chromosome constructions, e.g., human artificial chromosomes (HACs), yeast artificial chromosomes (YACs), bacterial artificial chromosomes (BACs), bacterial P1 constructions, or single chromosome cDNA libraries. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355; Price, C.M. (1993) Blood Rev. 7:127-134; and Trask, B.J. (1991) Trends Genet. 7:149-154.)

Fluorescent in situ hybridization (FISH) may be correlated with other physical chromosome mapping techniques and genetic map data. (See, e.g., Heinz-Ulrich, et al. (1995) in Meyers, supra, pp. 965-968.) Examples of genetic map data can be found in various scientific journals or at the Online Mendelian Inheritance in Man (OMIM) World Wide Web site. Correlation between the location of the gene encoding MTRP on a physical chromosomal map and a specific disorder, or a predisposition to a specific disorder, may help define the region of DNA associated with that disorder. The nucleotide sequences of the invention may be used to detect differences in gene sequences among normal, carrier, and affected individuals.

In situ hybridization of chromosomal preparations and physical mapping techniques, such as linkage analysis using established chromosomal markers, may be used for extending genetic maps. Often the placement of a gene on the chromosome of another mammalian species, such as mouse, may reveal associated markers even if the number or arm of a particular human chromosome is not known. New sequences can be assigned to chromosomal arms by physical mapping. This provides valuable information to investigators searching for disease genes using positional cloning or other gene discovery techniques. Once the disease or syndrome has been crudely localized by genetic linkage to a particular genomic region, e.g., ataxia-telangiectasia to 11q22-23, any sequences mapping to that area may represent associated or regulatory genes for further investigation. (See, e.g., Gatti, R.A. et al. (1988) Nature 336:577-580.) The nucleotide sequence of the subject invention may also be used to detect differences in the chromosomal location due to translocation, inversion, etc., among normal, carrier, or affected individuals.

In another embodiment of the invention, MTRP, its catalytic or immunogenic fragments, or

WO 00/26245

oligopeptides thereof can be used for screening libraries of compounds in any of a variety of drug screening techniques. The fragment employed in such screening may be free in solution, affixed to a solid support, borne on a cell surface, or located intracellularly. The formation of binding complexes between MTRP and the agent being tested may be measured.

Another technique for drug screening provides for high throughput screening of compounds having suitable binding affinity to the protein of interest. (See, e.g., Geysen, et al. (1984) PCT application WO84/03564.) In this method, large numbers of different small test compounds are synthesized on a solid substrate. The test compounds are reacted with MTRP, or fragments thereof, and washed. Bound MTRP is then detected by methods well known in the art. Purified MTRP can also be coated directly onto plates for use in the aforementioned drug screening techniques. Alternatively, non-neutralizing antibodies can be used to capture the peptide and immobilize it on a solid support.

In another embodiment, one may use competitive drug screening assays in which neutralizing antibodies capable of binding MTRP specifically compete with a test compound for binding MTRP. In this manner, antibodies can be used to detect the presence of any peptide which shares one or more antigenic determinants with MTRP.

In additional embodiments, the nucleotide sequences which encode MTRP may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleotide sequences that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following preferred specific embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

The disclosures of all patents, applications, and publications mentioned above and below, in particular U.S. Ser. No. [Attorney Docket No. PF-0633 P, filed November 4, 1998], U.S. Ser. No. [Attorney Docket No. PF-0645 P, filed November 24, 1998], U.S. Ser. No. [Attorney Docket No. PF-0657 P, filed December 22, 1998], and U.S. Ser. No. 60/121,896, are hereby expressly incorporated by reference.

EXAMPLES

I. Construction of cDNA Libraries

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RNA was purchased from Clontech or isolated from tissues described in Table 4. Some

PCT/US99/26048 WO 00/26245

tissues were homogenized and lysed in guanidinium isothiocyanate, while others were homogenized and lysed in phenol or in a suitable mixture of denaturants, such as TRIZOL (Life Technologies), a monophasic solution of phenol and guanidine isothiocyanate. The resulting lysates were centrifuged over CsCl cushions or extracted with chloroform. RNA was precipitated from the lysates with either isopropanol or sodium acetate and ethanol, or by other routine methods.

Phenol extraction and precipitation of RNA were repeated as necessary to increase RNA purity. In some cases, RNA was treated with DNase. For most libraries, poly(A+) RNA was isolated using oligo d(T)-coupled paramagnetic particles (Promega), OLIGOTEX latex particles (QIAGEN, Chatsworth CA), or an OLIGOTEX mRNA purification kit (QIAGEN). Alternatively, RNA was isolated directly from tissue lysates using other RNA isolation kits, e.g., the POLY(A)PURE mRNA purification kit (Ambion, Austin TX).

In some cases, Stratagene was provided with RNA and constructed the corresponding cDNA libraries. Otherwise, cDNA was synthesized and cDNA libraries were constructed with the UNIZAP vector system (Stratagene) or SUPERSCRIPT plasmid system (Life Technologies), using the recommended procedures or similar methods known in the art. (See, e.g., Ausubel, 1997, supra, units 5.1-6.6.) Reverse transcription was initiated using oligo d(T) or random primers. Synthetic oligonucleotide adapters were ligated to double stranded cDNA, and the cDNA was digested with the appropriate restriction enzyme or enzymes. For most libraries, the cDNA was size-selected (300-1000 bp) using SEPHACRYL S1000, SEPHAROSE CL2B, or SEPHAROSE CL4B column chromatography (Amersham Pharmacia Biotech) or preparative agarose gel electrophoresis. cDNAs were ligated into compatible restriction enzyme sites of the polylinker of a suitable plasmid, e.g., PBLUESCRIPT plasmid (Stratagene), PSPORT1 plasmid (Life Technologies), or pINCY (Incyte Pharmaceuticals, Palo Alto CA). Recombinant plasmids were transformed into competent E. coli cells including XL1-Blue, XL1-BlueMRF, or SOLR from Stratagene or DH5a, DH10B, or ElectroMAX DH10B from Life Technologies.

Isolation of cDNA Clones II.

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Plasmids were recovered from host cells by in vivo excision using the UNIZAP vector system (Stratagene) or by cell lysis. Plasmids were purified using at least one of the following: a Magic or WIZARD Minipreps DNA purification system (Promega); an AGTC Miniprep purification kit (Edge Biosystems, Gaithersburg MD); and QIAWELL 8 Plasmid, QIAWELL 8 Plus Plasmid, QIAWELL 8 Ultra Plasmid purification systems or the R.E.A.L. PREP 96 plasmid purification kit from QIAGEN. Following precipitation, plasmids were resuspended in 0.1 ml of distilled water and stored, with or without lyophilization, at 4°C.

Alternatively, plasmid DNA was amplified from host cell lysates using direct link PCR in a

high-throughput format (Rao, V.B. (1994) Anal. Biochem. 216:1-14). Host cell lysis and thermal cycling steps were carried out in a single reaction mixture. Samples were processed and stored in 384-well plates, and the concentration of amplified plasmid DNA was quantified fluorometrically using PICOGREEN dye (Molecular Probes, Eugene OR) and a FLUOROSKAN II fluorescence scanner (Labsystems Oy, Helsinki, Finland).

III. Sequencing and Analysis

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cDNA sequencing reactions were processed using standard methods or high-throughput instrumentation such as the ABI CATALYST 800 (Perkin-Elmer) thermal cycler or the PTC-200 thermal cycler (MJ Research) in conjunction with the HYDRA microdispenser (Robbins Scientific) or the MICROLAB 2200 (Hamilton) liquid transfer system. cDNA sequencing reactions were prepared using reagents provided by Amersham Pharmacia Biotech or supplied in ABI sequencing kits such as the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Perkin-Elmer). Electrophoretic separation of cDNA sequencing reactions and detection of labeled polynucleotides were carried out using the MEGABACE 1000 DNA sequencing system (Molecular Dynamics); the ABI PRISM 373 or 377 sequencing system (Perkin-Elmer) in conjunction with standard ABI protocols and base calling software; or other sequence analysis systems known in the art. Reading frames within the cDNA sequences were identified using standard methods (reviewed in Ausubel, 1997, supra, unit 7.7). Some of the cDNA sequences were selected for extension using the techniques disclosed in Example V.

20 The polynucleotide sequences derived from cDNA sequencing were assembled and analyzed using a combination of software programs which utilize algorithms well known to those skilled in the art. Table 5 summarizes the tools, programs, and algorithms used and provides applicable descriptions, references, and threshold parameters. The first column of Table 5 shows the tools, programs, and algorithms used, the second column provides brief descriptions thereof, the third column presents appropriate references, all of which are incorporated by reference herein in their entirety, and the fourth column presents, where applicable, the scores, probability values, and other parameters used to evaluate the strength of a match between two sequences (the higher the score, the greater the homology between two sequences). Sequences were analyzed using MACDNASIS PRO software (Hitachi Software Engineering, South San Francisco CA) and LASERGENE software (DNASTAR). Polynucleotide and polypeptide sequence alignments were generated using the default parameters specified by the clustal algorithm as incorporated into the MEGALIGN multisequence alignment program (DNASTAR), which also calculates the percent identity between aligned sequences.

The polynucleotide sequences were validated by removing vector, linker, and polyA

sequences and by masking ambiguous bases, using algorithms and programs based on BLAST, dynamic programing, and dinucleotide nearest neighbor analysis. The sequences were then queried against a selection of public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases, and BLOCKS, PRINTS, DOMO, PRODOM, and PFAM to acquire annotation using programs based on BLAST, FASTA, and BLIMPS. The sequences were assembled into full length polynucleotide sequences using programs based on Phred, Phrap, and Consed, and were screened for open reading frames using programs based on GeneMark, BLAST, and FASTA. The full length polynucleotide sequences were translated to derive the corresponding full length amino acid sequences, and these full length sequences were subsequently analyzed by querying against databases such as the GenBank databases (described above), SwissProt, BLOCKS, PRINTS, DOMO, PRODOM, Prosite, and Hidden Markov Model (HMM)-based protein family databases such as PFAM. HMM is a probabilistic approach which analyzes consensus primary structures of gene families. (See, e.g., Eddy, S.R. (1996) Curr. Opin. Struct. Biol. 6:361-365.)

The programs described above for the assembly and analysis of full length polynucleotide and amino acid sequences were also used to identify polynucleotide sequence fragments from SEQ ID NO:18-34. Fragments from about 20 to about 4000 nucleotides which are useful in hybridization and amplification technologies were described in The Invention section above.

IV. Northern Analysis

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Northern analysis is a laboratory technique used to detect the presence of a transcript of a gene and involves the hybridization of a labeled nucleotide sequence to a membrane on which RNAs from a particular cell type or tissue have been bound. (See, e.g., Sambrook, <u>supra</u>, ch. 7; Ausubel, 1995, <u>supra</u>, ch. 4 and 16.)

Analogous computer techniques applying BLAST were used to search for identical or related molecules in nucleotide databases such as GenBank or LIFESEQ (Incyte Pharmaceuticals). This analysis is much faster than multiple membrane-based hybridizations. In addition, the sensitivity of the computer search can be modified to determine whether any particular match is categorized as exact or similar. The basis of the search is the product score, which is defined as:

% sequence identity x % maximum BLAST score

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The product score takes into account both the degree of similarity between two sequences and the length of the sequence match. For example, with a product score of 40, the match will be exact within a 1% to 2% error, and, with a product score of 70, the match will be exact. Similar molecules are usually identified by selecting those which show product scores between 15 and 40, although lower scores may identify related molecules.

The results of northern analyses are reported as a percentage distribution of libraries in which the transcript encoding MTRP occurred. Analysis involved the categorization of cDNA libraries by organ/tissue and disease. The organ/tissue categories included cardiovascular, dermatologic, developmental, endocrine, gastrointestinal, hematopoietic/immune, musculoskeletal, nervous, reproductive, and urologic. The disease/condition categories included cancer, inflammation, trauma, cell proliferation, neurological, and pooled. For each category, the number of libraries expressing the sequence of interest was counted and divided by the total number of libraries across all categories. Percentage values of tissue-specific and disease- or condition-specific expression are reported in Table 3.

V. Extension of MTRP Encoding Polynucleotides

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The full length nucleic acid sequences of SEQ ID NO:18-34 were produced by extension of an appropriate fragment of the full length molecule using oligonucleotide primers designed from this fragment. One primer was synthesized to initiate 5' extension of the known fragment, and the other primer, to initiate 3' extension of the known fragment. The initial primers were designed using OLIGO 4.06 software (National Biosciences), or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the target sequence at temperatures of about 68°C to about 72°C. Any stretch of nucleotides which would result in hairpin structures and primer-primer dimerizations was avoided.

Selected human cDNA libraries were used to extend the sequence. If more than one extension was necessary or desired, additional or nested sets of primers were designed.

High fidelity amplification was obtained by PCR using methods well known in the art. PCR was performed in 96-well plates using the PTC-200 thermal cycler (MJ Research, Inc.). The reaction mix contained DNA template, 200 nmol of each primer, reaction buffer containing Mg²⁺, (NH₄)₂SO₄, and β-mercaptoethanol, Taq DNA polymerase (Amersham Pharmacia Biotech), ELONGASE enzyme (Life Technologies), and Pfu DNA polymerase (Stratagene), with the following parameters for primer pair PCI A and PCI B: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C. In the alternative, the parameters for primer pair T7 and SK+ were as follows: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 57°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C.

The concentration of DNA in each well was determined by dispensing 100 µl PICOGREEN quantitation reagent (0.25% (v/v) PICOGREEN; Molecular Probes, Eugene OR) dissolved in 1X TE and 0.5 µl of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar, Acton MA), allowing the DNA to bind to the reagent. The plate was scanned in a Fluoroskan II

(Labsystems Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantify the concentration of DNA. A 5 μ l to 10 μ l aliquot of the reaction mixture was analyzed by electrophoresis on a 1 % agarose mini-gel to determine which reactions were successful in extending the sequence.

The extended nucleotides were desalted and concentrated, transferred to 384-well plates, digested with CviJI cholera virus endonuclease (Molecular Biology Research, Madison WI), and sonicated or sheared prior to religation into pUC 18 vector (Amersham Pharmacia Biotech). For shotgun sequencing, the digested nucleotides were separated on low concentration (0.6 to 0.8%) agarose gels, fragments were excised, and agar digested with Agar ACE (Promega). Extended clones were religated using T4 ligase (New England Biolabs, Beverly MA) into pUC 18 vector (Amersham Pharmacia Biotech), treated with Pfu DNA polymerase (Stratagene) to fill-in restriction site overhangs, and transfected into competent <u>E. coli</u> cells. Transformed cells were selected on antibiotic-containing media, individual colonies were picked and cultured overnight at 37°C in 384-well plates in LB/2x carb liquid media.

The cells were lysed, and DNA was amplified by PCR using Taq DNA polymerase (Amersham Pharmacia Biotech) and Pfu DNA polymerase (Stratagene) with the following parameters: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 72°C, 2 min; Step 5: steps 2, 3, and 4 repeated 29 times; Step 6: 72°C, 5 min; Step 7: storage at 4°C. DNA was quantified by PICOGREEN reagent (Molecular Probes) as described above. Samples with low DNA recoveries were reamplified using the same conditions as described above. Samples were diluted with 20% dimethysulfoxide (1:2, v/v), and sequenced using DYENAMIC energy transfer sequencing primers and the DYENAMIC DIRECT kit (Amersham Pharmacia Biotech) or the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Perkin-Elmer).

In like manner, the nucleotide sequences of SEQ ID NO:18-34 are used to obtain 5'
regulatory sequences using the procedure above, oligonucleotides designed for such extension, and an appropriate genomic library.

VI. Labeling and Use of Individual Hybridization Probes

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Hybridization probes derived from SEQ ID NO:18-34 are employed to screen cDNAs, genomic DNAs, or mRNAs. Although the labeling of oligonucleotides, consisting of about 20 base pairs, is specifically described, essentially the same procedure is used with larger nucleotide fragments. Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 software (National Biosciences) and labeled by combining 50 pmol of each oligomer, 250 μ Ci of [γ -32P] adenosine triphosphate (Amersham Pharmacia Biotech), and T4 polynucleotide kinase (DuPont NEN, Boston MA). The labeled oligonucleotides are substantially purified using a

SEPHADEX G-25 superfine size exclusion dextran bead column (Amersham Pharmacia Biotech). An aliquot containing 10⁷ counts per minute of the labeled probe is used in a typical membrane-based hybridization analysis of human genomic DNA digested with one of the following endonucleases: Ase I, Bgl II, Eco RI, Pst I, Xba I, or Pvu II (DuPont NEN).

The DNA from each digest is fractionated on a 0.7% agarose gel and transferred to nylon membranes (Nytran Plus, Schleicher & Schuell, Durham NH). Hybridization is carried out for 16 hours at 40°C. To remove nonspecific signals, blots are sequentially washed at room temperature under conditions of up to, for example, 0.1 x saline sodium citrate and 0.5% sodium dodecyl sulfate. Hybridization patterns are visualized using autoradiography or an alternative imaging means and compared.

VII. Microarrays

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A chemical coupling procedure and an ink jet device can be used to synthesize array elements on the surface of a substrate. (See, e.g., Baldeschweiler, supra.) An array analogous to a dot or slot blot may also be used to arrange and link elements to the surface of a substrate using thermal, UV, chemical, or mechanical bonding procedures. A typical array may be produced by hand or using available methods and machines and contain any appropriate number of elements. After hybridization, nonhybridized probes are removed and a scanner used to determine the levels and patterns of fluorescence. The degree of complementarity and the relative abundance of each probe which hybridizes to an element on the microarray may be assessed through analysis of the scanned images.

Full-length cDNAs, Expressed Sequence Tags (ESTs), or fragments thereof may comprise the elements of the microarray. Fragments suitable for hybridization can be selected using software well known in the art such as LASERGENE software (DNASTAR). Full-length cDNAs, ESTs, or fragments thereof corresponding to one of the nucleotide sequences of the present invention, or selected at random from a cDNA library relevant to the present invention, are arranged on an appropriate substrate, e.g., a glass slide. The cDNA is fixed to the slide using, e.g., UV cross-linking followed by thermal and chemical treatments and subsequent drying. (See, e.g., Schena, M. et al. (1995) Science 270:467-470; Shalon, D. et al. (1996) Genome Res. 6:639-645.) Fluorescent probes are prepared and used for hybridization to the elements on the substrate. The substrate is analyzed by procedures described above.

VIII. Complementary Polynucleotides

Sequences complementary to the MTRP-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring MTRP. Although use of oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same

procedure is used with smaller or with larger sequence fragments. Appropriate oligonucleotides are designed using OLIGO 4.06 software (National Biosciences) and the coding sequence of MTRP. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a complementary oligonucleotide is designed to prevent ribosomal binding to the MTRP-encoding transcript.

IX. Expression of MTRP

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Expression and purification of MTRP is achieved using bacterial or virus-based expression systems. For expression of MTRP in bacteria, cDNA is subcloned into an appropriate vector containing an antibiotic resistance gene and an inducible promoter that directs high levels of cDNA transcription. Examples of such promoters include, but are not limited to, the trp-lac (tac) hybrid 10 promoter and the T5 or T7 bacteriophage promoter in conjunction with the lac operator regulatory element. Recombinant vectors are transformed into suitable bacterial hosts, e.g., BL21(DE3). Antibiotic resistant bacteria express MTRP upon induction with isopropyl beta-Dthiogalactopyranoside (IPTG). Expression of MTRP in eukaryotic cells is achieved by infecting insect or mammalian cell lines with recombinant Autographica californica nuclear polyhedrosis virus 15 (AcMNPV), commonly known as baculovirus. The nonessential polyhedrin gene of baculovirus is replaced with cDNA encoding MTRP by either homologous recombination or bacterial-mediated transposition involving transfer plasmid intermediates. Viral infectivity is maintained and the strong polyhedrin promoter drives high levels of cDNA transcription. Recombinant baculovirus is used to infect Spodoptera frugiperda (Sf9) insect cells in most cases, or human hepatocytes, in some cases. 20 Infection of the latter requires additional genetic modifications to baculovirus. (See Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945.)

In most expression systems, MTRP is synthesized as a fusion protein with, e.g., glutathione

S-transferase (GST) or a peptide epitope tag, such as FLAG or 6-His, permitting rapid, single-step, affinity-based purification of recombinant fusion protein from crude cell lysates. GST, a 26-kilodalton enzyme from Schistosoma japonicum, enables the purification of fusion proteins on immobilized glutathione under conditions that maintain protein activity and antigenicity (Amersham Pharmacia Biotech). Following purification, the GST moiety can be proteolytically cleaved from MTRP at specifically engineered sites. FLAG, an 8-amino acid peptide, enables immunoaffinity purification using commercially available monoclonal and polyclonal anti-FLAG antibodies (Eastman Kodak). 6-His, a stretch of six consecutive histidine residues, enables purification on metal-chelate resins (QIAGEN). Methods for protein expression and purification are discussed in Ausubel (1995, supra, ch. 10 and 16). Purified MTRP obtained by these methods can be used directly in the

following activity assay,

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X. Demonstration of MTRP Activity

ATPase activity associated with MTRP can be measured by hydrolysis of radiolabeled ATP- $[\gamma^{-32}P]$, separation of the hydrolysis products by chromatographic methods, and quantitation of the recovered ^{32}P using a scintillation counter. The reaction mixture contains ATP- $[\gamma^{-32}P]$ and varying amounts of MTRP in a suitable buffer incubated at 37°C for a suitable period of time. The reaction is terminated by acid precipitation with trichloroacetic acid and then neutralized with base, and an aliquot of the reaction mixture is subjected to membrane or filter paper-based chromatography to separate the reaction products. The amount of ^{32}P liberated is counted in a scintillation counter. The amount of radioactivity recovered is proportional to the ATPase activity of MTRP in the assay.

MTRP transport activity is assayed by measuring uptake of labeled substrates into <u>Xenopus</u> laevis oocytes. Oocytes at stages V and VI are injected with MTRP mRNA (10 ng per oocyte) and incubated for 3 days at 18°C in OR2 medium (82.5mM NaCl, 2.5 mM KCl, 1mM CaCl₂, 1mM MgCl₂, 1mM Na₂HPO₄, 5 mM Hepes, 3.8 mM NaOH, 50μg/ml gentamycin, pH 7.8) to allow expression of MTRP protein. Oocytes are then transferred to standard uptake medium (100mM NaCl, 2 mM KCl, 1mM CaCl₂, 1mM MgCl₂, 10 mM Hepes/Tris, pH 7.5). Uptake of various substrates (e.g., amino acids, sugars, drugs, and neurotransmitters) is initiated by adding a ³H-labeled substrate to the oocytes. After 30 minutes of incubation, uptake is terminated by washing the oocytes three times in Na[†]-free medium. Incorporation of ³H is measured, and compared with controls. MTRP transport activity is proportional to the level of internalized ³H-labeled substrate.

XI. Functional Assays

MTRP function is assessed by expressing the sequences encoding MTRP at physiologically elevated levels in mammalian cell culture systems. cDNA is subcloned into a mammalian expression vector containing a strong promoter that drives high levels of cDNA expression. Vectors of choice include pCMV SPORT (Life Technologies) and pCR3.1 (Invitrogen, Carlsbad CA), both of which contain the cytomegalovirus promoter. 5-10 μ g of recombinant vector are transiently transfected into a human cell line, for example, an endothelial or hematopoietic cell line, using either liposome formulations or electroporation. 1-2 μ g of an additional plasmid containing sequences encoding a marker protein are co-transfected. Expression of a marker protein provides a means to distinguish transfected cells from nontransfected cells and is a reliable predictor of cDNA expression from the recombinant vector. Marker proteins of choice include, e.g., Green Fluorescent Protein (GFP; Clontech), CD64, or a CD64-GFP fusion protein. Flow cytometry (FCM), an automated, laser optics-based technique, is used to identify transfected cells expressing GFP or CD64-GFP and to evaluate the apoptotic state of the cells and other cellular properties. FCM detects and quantifies the uptake of

fluorescent molecules that diagnose events preceding or coincident with cell death. These events include changes in nuclear DNA content as measured by staining of DNA with propidium iodide; changes in cell size and granularity as measured by forward light scatter and 90 degree side light scatter; down-regulation of DNA synthesis as measured by decrease in bromodeoxyuridine uptake; alterations in expression of cell surface and intracellular proteins as measured by reactivity with specific antibodies; and alterations in plasma membrane composition as measured by the binding of fluorescein-conjugated Annexin V protein to the cell surface. Methods in flow cytometry are discussed in Ormerod, M.G. (1994) Flow Cytometry, Oxford, New York NY.

The influence of MTRP on gene expression can be assessed using highly purified populations of cells transfected with sequences encoding MTRP and either CD64 or CD64-GFP. CD64 and CD64-GFP are expressed on the surface of transfected cells and bind to conserved regions of human immunoglobulin G (IgG). Transfected cells are efficiently separated from nontransfected cells using magnetic beads coated with either human IgG or antibody against CD64 (DYNAL, Lake Success NY). mRNA can be purified from the cells using methods well known by those of skill in the art. Expression of mRNA encoding MTRP and other genes of interest can be analyzed by northern analysis or microarray techniques.

XII. Production of MTRP Specific Antibodies

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MTRP substantially purified using polyacrylamide gel electrophoresis (PAGE; see, e.g., Harrington, M.G. (1990) Methods Enzymol. 182:488-495), or other purification techniques, is used to immunize rabbits and to produce antibodies using standard protocols.

Alternatively, the MTRP amino acid sequence is analyzed using LASERGENE software (DNASTAR) to determine regions of high immunogenicity, and a corresponding oligopeptide is synthesized and used to raise antibodies by means known to those of skill in the art. Methods for selection of appropriate epitopes, such as those near the C-terminus or in hydrophilic regions are well described in the art. (See, e.g., Ausubel, 1995, supra, ch. 11.)

Typically, oligopeptides of about 15 residues in length are synthesized using an ABI 431A peptide synthesizer (Perkin-Elmer) using fmoc-chemistry and coupled to KLH (Sigma-Aldrich, St. Louis MO) by reaction with N-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS) to increase immunogenicity. (See, e.g., Ausubel, 1995, supra.) Rabbits are immunized with the oligopeptide-KLH complex in complete Freund's adjuvant. Resulting antisera are tested for antipeptide and anti-MTRP activity by, for example, binding the peptide or MTRP to a substrate, blocking with 1% BSA, reacting with rabbit antisera, washing, and reacting with radio-iodinated goat anti-rabbit IgG.

XIII. Purification of Naturally Occurring MTRP Using Specific Antibodies

Naturally occurring or recombinant MTRP is substantially purified by immunoaffinity

chromatography using antibodies specific for MTRP. An immunoaffinity column is constructed by covalently coupling anti-MTRP antibody to an activated chromatographic resin, such as CNBr-activated SEPHAROSE (Amersham Pharmacia Biotech). After the coupling, the resin is blocked and washed according to the manufacturer's instructions.

Media containing MTRP are passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of MTRP (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt antibody/MTRP binding (e.g., a buffer of pH 2 to pH 3, or a high concentration of a chaotrope, such as urea or thiocyanate ion), and MTRP is collected.

XIV. Identification of Molecules Which Interact with MTRP

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MTRP, or biologically active fragments thereof, are labeled with ¹²⁵I Bolton-Hunter reagent. (See, e.g., Bolton A.E. and W.M. Hunter (1973) Biochem. J. 133:529-539.) Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled MTRP, washed, and any wells with labeled MTRP complex are assayed. Data obtained using different concentrations of MTRP are used to calculate values for the number, affinity, and association of MTRP with the candidate molecules.

Various modifications and variations of the described methods and systems of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention.

20 Although the invention has been described in connection with certain embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the described modes for carrying out the invention which are obvious to those skilled in molecular biology or related fields are intended to be within the scope of the following claims.

Table 1

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Fragments	642329R1 (BRSTNOT03), 961344H1 (BRSTTUT03), 3149970H1 (ADRENON04), 3255621H1 (OVARTUN01), 4265773H1 (KIDNNOT32), 4641320H1 (PROSTMT03), 4875558H1 (COLDNOT01)	148511X20R1 (FIBRNGT01), 580891H1 (BRAVTXT05), 1304328F1 (PLACNOT02), 1469890T6 (PANCTUT02), 1799816T6 (COLNNOT27), 1985910R6 (LUNGAST01), 2722244F6 (LUNGTUT10), 3128782F6 (LUNGTUT12), 3128782H1 (LUNGTUT12), 3276993F6 (PROSBPT06), SBMA03256F1	744485R1 (BRAITUT01), 859118R1 (BRAITUT03), 944049T1 (ADRENOT03), 1432755R1 (BEPINON01), 1720440H1 (BLADNOT06), 1808737X11C1 (PROSTUT12), 1812106F6 (PROSTUT12), 2192988F6 (THYRTUT03), 2192988X13F1 (THYRTUT03), 2192988X14F1 (THYRTUT03), 3384757H1 (ESOGNOT04)	1732422F6 (BRSTTUT08), 2098563H1 (BRAITUT02), 2274290H1 (PROSNON01), 2274290X326D2 (PROSNON01), 2598580F6 (UTRSNOT10), 2779864F6 (OVARTUT03), 2864759F6 (KIDNNOT20), 2864759T6 (KIDNNOT20), 3221871H1 (COLNNON03)	966363H1 (BRSTNOTO5), 1000112R1 (BRSTNOTO3), 1647057F6 (PROSTUT09), 2740029H1 (BRSTTUT14), 2740029X30F1 (BRSTTUT14), 2740029X321F1 (BRSTTUT14), 5068692H1 (PANCNOT23), 5193978H1 (LUNLTUT04), SBOA03895D1	1508631H1 (LUNGNOT14), 1596418F1 (BRAINOT14), 2414415F6 (HNT3AZT01), 2414415H1 (HNT3AZT01), 2414415X300D1 (HNT3AZT01), 2902794H1 (DRGCNOT01), 3080373H1 (BRAIUNT01), 3554689H1 (SYNONOT01), 3881949F6 (SPLNNOT11), 4996983H1 (MYEPTXT02)	627910R6 (KIDNNOTOS), 2358261R6 (LUNGFETOS), 2358464F6 (LUNGFETOS), 2466714H1 (THYRNOTO8), 2726053F6 (OVARTUTOS), 3845383H1 (DENDNOTO1), 4228854H1 (BRAMDITO1)
Library	BRSTTUT03	LUNGTUT12	BLADNOT06	PROSNON01	BRSTTUT14	HNT3AZT01	THYRNOT08
Clone ID	961344	3128782	1720440	2274290	2740029	2414415	2466714
Nucleotide SEQ ID NO:	18	19	20	21	22	23	24
Protein SEQ ID NO:	1	2	٣	4	Ω	9	٢

	T									7
Fragments	1311045F6 (COLNFET02), 1375061F6 (LUNGNOT10), 1580318F6 (DUODNOT01), 2016443F6 (ENDCNOT03), 2617942F6 (GBLANOT01), 2617942H1 (GBLANOT01), 2703625H1 (OVARTUT10)	782666T6 (MYOMNOT01), 2715384T6 (THYRNOT09), 2945431F6 (BRAITUT23), 2945431H1 (BRAITUT23), 4114919F6 (UTRSTUT07)	1754278T6 (LIVRTUT01), 1830341T6 (THP1AZT01), 2122328T6 (BRSTNOT07), 4074113F6 (KIDNNOT26), 4074113H1 (KIDNNOT26)	1413743H1 (BRAINOT12), 2643096F6 (LUNGTUT08), SBWA04580V1, SBWA04306V1, SBWA00027V1, SBWA00447V1, SBWA00075V1, SBWA03280V1, SBWA00778V1	1733477H1 (BRSTTUT08), 1733477F6 (BRSTTUT08), 3346594F6 (BRAITUT24), 1696051F6 (COLNUCT03), SAFC01922F1, 3234341F6 (COLNUCT03), 2634476T6 (COLNTUT15), SBCA03249F1	2641908H1 (LUNGTUT08), SASA01215F1, SASB01010F1, SASB01616F1, SASA03523F1	2656554H1 (LUNGTUT09), 2656554F6 (LUNGTUT09), 2656554T6 (LUNGTUT09)	2719228H1 (THYRNOT09), 2719228F6 (THYRNOT09), 532553H1 (BRAINOT03), 1626110F6 (COLNPOT01), 1626488F6 (COLNPOT01), 2100947R6 (BRAITUT02), SBGA05174F1	365782441 (ENDPNOTO2), 2450345F6 (ENDANOTO1), 2756551H1 (THPIAZSO8), SCBA01425V1, 2394519T6 (THPIAZTO1)	5378485H1 (BRAXNOT01), SCAA06347V1, 1570276F1 (UTRSNOT05)
Library	GBLANOT01	BRAITUT23	KIDNNOT26	BRAINOT12	BRSTTUT08	LUNGTUT08	LUNGTUT09	THYRNOT09	ENDPNOT02	BRAXNOT01
Clone ID	2617942	2945431	4074113	1413743	1733477	2641908	2656554	2719228	3657824	5378485
Nucleotide SEQ ID NO:	25	26	27	28	29	30	31	32	33	34
Protein SEQ ID NO:	ω ,	6	10	11	12	13	14	15	16	17
	otein Nucleotide Clone Library ID NO: SEQ ID NO: ID	Nucleotide Clone Library Fragments Fragments	Nucleotide Clone Library Fragments	Nucleotide	Nucleotide	Protein Nucleotide	Protein Nucleotide Clone Library Fragments	Protein Nucleotide Clone Library Fragments	Profein Nucleotide	SEQ ID NO: SEQ ID NO: Library Electrical Library

Table 2

Analytical Methods	BLAST BLOCKS MOTIFS PFAM PRINTS	BLAST BLOCKS MOTIFS PFAM PRINTS ProfileScan SPScan	BLAST MOTIFS PFAM PRINTS ProfileScan
Identification/ Homologies	E1-E2 ATPase [Mus musculus] (GI 2944187)	ABC transporter protein isoform [Homo sapiens] (GI 2522534)	Fatty acid transport protein [Homo sapiens] (GI 4206376) Fatty acid transport protein [Mus musculus] (GI 2612939)
Signature Sequences	E1-E2 ATPase signatures: R85-D115, T106-I128 P-type cation transporter signature: I109-I128 Transmembrane domains: I330-W351, L271-I288	ABC transporter signatures: F453-L467, G330-Q525, D652-Q816, L744-A758 ATP-binding (P-loop) domains: G337-T344, G659-S666	AMP-binding enzyme motif: R4-V404 AMP-binding domain signature: K91-M144 Lipocalin signature: E2-M14
Potential Glycosylation Sites		N301	N195 N238 N258 N383
Potential Phosphorylation Sites	S375 S34 T33 T89 T102 S161 S246 T298	T80 S106 S22 T40 T109 S110 S141 S229 S245 S255 S288 T408 T614 T679 T707 S819 S24 S274 S293 T386 T450 T536 S568 T581 T610 T673	S183 S99 S385 T387 T39 T106 S240 T376 T461 Y111 Y151 Y194 Y353
Amino Acid Residues	384	846	511
SEQ ID NO:	-1	2	ĸ

Analytical Methods	BLAST BLOCKS MOTIFS PFAM HMM	BLAST BLOCKS PFAM ProfileScan HMM
Identification/ Homologies	ABC transporter protein [Homo sapiens] (GI 4321407) SMDR1 [Schistosoma mansoni] (GI 425474)	Sodium:solute symporter [Homo sapiens] (GI 4884550)
Signature Sequences	ABC transporter domain: L595-L609 ABC transporter family signatures: G483-V499, L595-D626 ABC transporter transmembrane domain: L128-M408 ABC transporter nucleotide binding domain: G483-G668 ATP/GTP-binding site motif A: G490-T497 Transmembrane domain: F123-V144	Sodium:solute symporter family signature: M61-A467, N169-A216, T172-V217, G353-I383, I430-A467 Transmembrane domains: M27-Y45, F256-A276, I430-F448, V458-G480
Potential Glycosylation Sites	N176 N229 N410	N138 N489 N498 N534
Potential Phosphorylation Sites	S303 T224 T390 S501 T547 S560 S579 S623 T675 S248 S274 T326 S629 T655	S25 T55 S128 T172 S242 S284 T378 T140 S222 S283 T286 T549
Amino Acid Residues	718	635
SEQ ID NO:	4	

Analytical Methods	BLAST BLOCKS MOTIFS PFAM	BLAST BLOCKS MOTIFS	BLAST MOTIFS	BLAST BLOCKS , MOTIFS PFAM Profilescan
Identification/ Homologies	SLC7A8 [Homo sapiens] (GI 4581470) Amino acid transporter (GI 3639058, SEQ ID NO:38)	Aromatic amino acid transporter (GI 1840045, SEQ ID NO:39)	UDP-N-acetylglucosamine transporter [Canis familiaris] (GI 3298605)	ABC transporter (GI 3335175, SEQ ID NO:41)
Signature Sequences	Amino acid permeases signature: L36-K474 (from PFAM) Transmembrane domains: L76-L96, K111-Y131, S159-N176, I189-V207, N264-Y287, I422-F449, I451-W469	Aromatic amino acid permease signature: N53-S98 (from BLOCKS) Transmembrane domains: F47-V69, Y68-L88, T169-L191, L197-I216, S366-Y387, M330-I347, F432-I454	Transmembrane domains: I209-D227, I284-T307	ABC transporters signature: S2-G141 (from PFAM)
Potential Glycosylation Sites		·		
nino Potential Po cid Phosphorylation Glyc idues Sites	T11 S61 S103 S179 S292 T305 S337 T363 S487 S502 Y109	T106 S107 S215 S277 T306 T331 S419 S425	T23 T29 S65 T84 S106 S133 S160	S4 T22 S132 T151
Amino Acid Residues	535	456	325	178
SEQ ID NO:	9	7	ω	6

Analytical Methods	BLAST BLOCKS MOTIFS PFAM Profilescan	BLAST MOTIFS PRINTS	BLAST MOTIFS PFAM	BLAST HMM MOTIFS	BLAST HMM MOTIFS
Identification/ Homologies	Anthracycline resistance-associated B protein (ABC transporter; GI 1279457, P SEQ ID NO:42)	Transmembrane transporter [Schizosaccharomyces P	Transporter [Mus B musculus] (GI 5359730) M Transporter [Homo sapiens] (GI 291964)	Organic anion transporter OATP-B [Homo H sapiens] (GI 5006263)	Similar to Human Na+/H+ B exchanger 2 (A57644) H (Homo sapiens) (GI M
Signature Sequences	ABC transporters signature: Q45-G227 (from PFAM)	Aromatic amino acid permease signatures: F58-A104, A243-F265, L375-K394	Sulfate transporter signature: L198-T508	Transmembrane domains: L211-G230, A175-L196, W42-F61, F139-F159	Transmembrane domain: V22-F41
Potential . Glycosylation Sites			N614 N614	N116	N81
Potential Phosphorylation Sites	T89 S107 S214	T179 S11 S37 T114 S420 T174	T174 T251 S376 T615 S616 S633 S678 S713 S733 T138 S574 S603 T656 S687 T726 Y703	T32 S99 S254 T274 S103 T104 T133 S273	S9 S92 Y14
Amino Acid Residues	255	462	758	336	103
SEQ ID NO:	10	11	12	13	14

SEQ Amino Potential Potential ID Acid Phosphorylation Glycosylation Sites Sites	Potential Potentia Phosphorylation Glycosylat Sites Sites	Potentia Glycosylat Sites		Signature Sequences	Identification/ Homologies	Analytical Methods
123 S21 S118 T119 N39		N39	1	Signal peptide: M1-S21	Similar to Sugar transporter [Caenorhabditis elegans] (GI 3878537)	BLAST HMM MOTIFS
222 T83 S61 S74	T83 S61 S74				E. coli cation transport protein (GI495778)	BLAST MOTIFS
111 N100	N100	N100			Vacuolar proton-ATPase subunit M9.2 [Homo sapiens] (GI 2584789)	BLAST MOTIFS

Table 3

Nucleotide SEQ ID NO:	Tissue Expression (Fraction of Total)	Disease or Condition (Fraction of Total)	Vector
18	Reproductive (0.292) Nervous (0.167) Hematopoietic/Immune (0.125)	Cell proliferative (0.750) Inflammation (0.208)	PSPORT1
19	Reproductive (0.264) Nervous (0.155) Hematopoietic/Immune (0.145)	Cell proliferative (0.691) Inflammation (0.236)	pINCY
20	Reproductive Nervous Gastrointestinal	Cell proliferation (0.66) Inflammation and immune response (0.24)	pincy
21	Reproductive Nervous Gastrointestinal Hematopoietic/Immune	Cell proliferation (0.73) Inflammation and immune response (0.21)	PSPORT1
22	Nervous Reproductive	Cell proliferation (0.76) Inflammation and immune response (0.33)	pINCY
23	Reproductive (0.333) Nervous (0.296)	Cancer (0.519) Inflammation (0.185)	
24	Reproductive (0.250) Musculoskeletal (0.179) Hematopoietic/Immune (0.143)	Cancer (0.429) Inflammation (0.286)	
25	Reproductive (0.360) Gastrointestinal (0.200) Cardiovascular (0.120)	Cancer (0.520) Inflammation (0.160)	-
26	Nervous (0.267) Endocrine (0.133)	Cancer (0.533) Inflammation (0.333)	

Nucleotide SEQ ID NO:	Tissue Expression (Fraction of Total)	Disease or Condition (Fraction of Total)	Vector
27	Urologic (0.333) Cardiovascular (0.167) Developmental (0.167)	Cancer (0.667) Inflammation (0.167)	
28	Nervous (0.222) Hematopoietic/Immune (0.185) Reproductive (0.185)	Cell Proliferation (0.481) Inflammation (0.407)	pINCY
29	Gastrointestinal (0.286) Reproductive (0.257) Nervous (0.200)	Cell Proliferation (0.743) Inflammation (0.286)	pINCY
30	Cardiovascular (0.400) Nervous (0.200) Reproductive (0.200) Gastrointestinal (0.200)	Cell Proliferation (0.800) Inflammation (0.200)	pINCY
31	Hematopoietic/Immune (0.500) Reproductive (0.250) Cardiovascular (0.250)	Cell Proliferation (0.750) Inflammation (0.500)	pINCY
32	Gastrointestinal (0.304) Hematopoietic/Immune (0.174) Nervous (0.174)	Cell Proliferation (0.826) Inflammation (0.347)	pINCY
33	Cardiovascular (0.238) Nervous (0.190) Hematopoietic/Immune (0.143)	Cell Proliferation (0.571) Inflammation (0.473)	PINCY
34	Nervous (0.510) Cardiovascular (0.102)	Cell Proliferation (0.612) Inflammation (0.266)	pINCY

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PF-0633 PCT

Nucleotide SEQ ID NO:	Library	Library Comment
	BRSTTUT03	Library was constructed using RNA isolated from breast tumor tissue removed from a 58-year-old Caucasian female during a unilateral extended simple mastectomy. Pathology indicated multicentric invasive grade 4 lobular carcinoma. The mass was identified in the upper outer quadrant, and three separate nodules were found in the lower outer quadrant of the left breast. Patient history included skin cancer, rheumatic heart disease, osteoarthritis, and tuberculosis. Family history included cerebrovascular disease, coronary artery aneurysm, breast cancer, prostate cancer, atherosclerotic coronary artery disease, and type I diabetes.
19	LUNGTUT12	Library was constructed using RNA isolated from tumorous lung tissue removed from a 70-year-old Caucasian female during a lung lobectomy of the left upper lobe. Pathology indicated grade 3 (of 4) adenocarcinoma and vascular invasion. Patient history included tobacco abuse, depressive disorder, anxiety state, and skin cancer. Family history included cerebrovascular disease, congestive heart failure, colon cancer, depressive disorder, and primary liver cancer.
20	BLADNOT06	Library was constructed using RNA isolated from posterior wall bladder tissue removed from a 66-year-old Caucasian male during a radical prostatectomy, radical cystectomy, and urinary diversion. Pathology for the associated tumor tissue indicated grade 3 transitional cell carcinoma on the anterior wall of the bladder and urothelium. Patient history included lung neoplasm. Family history included a malignant breast neoplasm, tuberculosis, cerebrovascular disease, atherosclerotic coronary artery disease, and lung cancer.
21	PROSNON01	Normalized prostate library was constructed from 4.4 million independent clones from a prostate library. Starting RNA was made from prostate tissue removed from a 28-year-old, Caucasian male. The normalization and hybridization conditions were adapted from Soares, M.B. et al. (1994) Proc. Natl. Acad. Sci. USA 91:9228-9232, using a longer (19 hour) reannealing hybridization period.

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Nucleotide	Library	Library Comment
SEQ ID NO:		
22	BRSTIUT14	Library was constructed using RNA isolated from breast tumor tissue removed from a 62-year-old Caucasian female during a unilateral extended simple mastectomy. Pathology indicated an invasive grade 3 (of 4), nuclear grade 3 (of 3) adenocarcinoma, ductal type. Ductal carcinoma in situ, comedo type, comprised 60% of the tumor mass. Metastatic adenocarcinoma was identified in one (of 14) axillary lymph nodes with no perinodal extension. Tumor cells were strongly positive for estrogen receptors and weakly positive for progesterone receptors. Patient history included benign colon neoplasm, hyperlipidemia, cardiac dysrhythmia, and obesity. Family history included atherosclerotic coronary artery disease, myocardial infarction, colon cancer, ovarian cancer, lung cancer, and cerebrovascular disease.
23	HNT3AZT01	Library was constructed using RNA isolated from the hNT2 cell line (derived from a human teratocarcinoma that exhibited properties characteristic of a committed neuronal precursor). Cells were treated for three days with 0.35 micromolar 5-aza-2'-deoxycytidine (AZ).
24	THYRNOT08	Library was constructed using RNA isolated from the diseased left thyroid tissue removed from a 13-year-old Caucasian female during a complete thyroidectomy. Pathology indicated lymphocytic thyroiditis.
25	GBLANOT01	Library was constructed using RNA isolated from diseased gallbladder tissue removed from a 53-year-old Caucasian female during a cholecystectomy. Pathology indicated mild chronic cholecystitis and cholelithiasis with approximately 150 mixed gallstones. Family history included benign hypertension.
26	BRAITUT23	Library was constructed using RNA isolated from left posterior brain tumor tissue removed from a 36-year-old male during a cerebral meninges lesion excision. Pathology indicated, meningioma. Family history included malignant skin melanoma, atherosclerotic coronary artery disease, hyperlipidemia, Huntington's chorea, and rheumatoid arthritic

Nucleotide SEQ ID NO:	Library	Library Comment
27	KIDNNOT26	Library was constructed using RNA isolated from left kidney medulla and cortex tissue removed from a 53-year-old Caucasian female during a nephroureterectomy. Patient history included hyperlipidemia, cardiac dysrhythmia, metrorrhagia, cerebrovascular disease, and atherosclerotic coronary artery disease.
28	BRAINOT12	Library was constructed using RNA isolated from brain tissue removed from the right frontal lobe of a 5-year-old Caucasian male during a hemispherectomy. Pathology indicated extensive polymicrogyria and mild to moderate gliosis (predominantly subpial and subcortical), which are consistent with chronic seizure disorder. The patient presented with intractable convulsive epilepsy. Family history included a cervical neoplasm.
29	BRSTIUT08	Library was constructed using RNA isolated from breast tumor tissue removed from a 45-year-old Caucasian female during unilateral extended simple mastectomy. Pathology indicated invasive nuclear grade 2-3 adenocarcinoma (ductal type), with 3 of 23 lymph nodes positive for metastatic disease. Greater than 50% of the tumor volume was in-situ, both comedo and non-comedo types. There were also positive estrogen/progesterone receptors and uninvolved tissue showing proliferative changes. Patient history included valvuloplasty of mitral valve without replacement, rheumatic mitral insufficiency, and rheumatic heart disease. Family history included acute myocardial infarction and atherosclerotic coronary artery disease and type II diabetes.

Nucleotide Library LUNGTUTO8 Library was constructed using RNA isolated from lung tumor tissue removed 63-year-old Caucasian male during a right upper lobectomy with fiberoptic Pathology indicated a grade 3 adenocarcinoma, forming a mass penetrating the right upper lung lobe. The bronchial margh of the resection was free of tissue from the superior segment of the right lower lobe lung showed multiple lymph nodes were negative for tumor. In add yeast forms resembling histoplasma identified in two lymph nodes stained yeast forms resembling histoplasma identified in two lymph nodes stained yeast forms resembling history included atherosclerotic coronary artery disease myocardial infarction, rectal cancer, an asymtomatic abdominal aortic aneu abuse, and cardiac dysrbythmia. Family history included congestive heart for myocardial infarction. 31 LUNGTUTO9 Library was constructed RNA isolated from lung tumor tissue removed from a squamous cell carcinoma in the right upper lobe, forming an infiltrating mithe bronchus and the surrounding parenchyma. One (of 4) intrapulmonary per lymph nodes contained a metastatic tumor. An apical cap was identified. On paratracheal lower lymph nodes contained a metastatic tumor. Permanent supmediastinal sections revealed metastatic squamous cell carcinoms in the lymph Patient history included of type II diabetes, thyvoid disorder. deriversive			
LUNGTUT08	Nucleotide SEQ ID NO:	Library	Library Comment
LUNGTUT09	30	LUNGTUTO8	Library was constructed using RNA isolated from lung tumor tissue removed from a 63-year-old Caucasian male during a right upper lobectomy with fiberoptic bronchoscopy. Pathology indicated a grade 3 adenocarcinoma, forming a mass penetrating the pleura of the right upper lung lobe. The bronchial margin of the resection was free of tumor. The tissue from the superior segment of the right lower lobe lung showed multiple (2) calcified granulomas. Multiple lymph nodes were negative for tumor. In addition, budding yeast forms resembling histoplasma identified in two lymph nodes stained with GMS (silver). Patient history included atherosclerotic coronary artery disease, an acute myocardial infarction, rectal cancer, an asymtomatic abdominal aortic aneurysm, tobacco abuse, and cardiac dysrhythmia. Family history included congestive heart failure, stomach cancer, lung cancer, type II diabetes, atherosclerotic coronary artery disease, and acute myocardial infarction.
hyperlipidemia, esophageal ulcer, and tobacco use. Family history included and atherosclerotic coronary artery disease	31	LUNGTUT09	Library was constructed RNA isolated from lung tumor tissue removed from a 68-year-old Caucasian male during segmental lung resection. Pathology indicated invasive grade 3 squamous cell carcinoma in the right upper lobe, forming an infiltrating mass involving the bronchus and the surrounding parenchyma. One (of 4) intrapulmonary peribronchial lymph nodes contained a metastatic tumor. An apical cap was identified. One (of 15) right paratracheal lower lymph nodes contained a metastatic tumor. Permanent superior mediastinal sections revealed metastatic squamous cell carcinoma in the lymph nodes. Patient history included of type II diabetes, thyroid disorder, depressive disorder, hyperlipidemia, esophageal ulcer, and tobacco use. Family history included brain cancer and atherosclerotic coronary artery, disease

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	Nucleotide SEQ ID NO:	Library	Library Comment
	32	THYRNOT09	Library was constructed using RNA isolated from diseased thyroid tissue removed from an 18-year-old Caucasian female during an unilateral thyroid lobectomy and regional lymph node excision. Pathology indicated adenomatous goiter. This was associated with a follicular adenoma of the thyroid. The right neck lymph nodes displayed reactive follicular hyperplasia. The patient presented with hypophosphatemia. Patient history included compression of the brain, headache, iron deficiency anemia, active rickets, epidermal nevus syndrome (treated with lasers), and osteitis deformans. Family history included thyroid cancer, type II diabetes, benign hypertension, and nodular lymphoma.
7(33	ENDPNOT02	Library was constructed using RNA isolated from pulmonary artery endothelial cells removed from a 10-year-old Caucasian male. The cells were treated with TNF alpha and IL-1 beta 10ng/ml each for 20 hours.
)	34	BRAXNOT01	Library was constructed using RNA isolated from cerebellar tissue removed from a 70-year-old male. Patient history included chronic obstructive airways disease and left ventricular failure.

Table ?

Program	Description	Reference	Parameter Threehold
ABI FACTURA	A program that removes vector sequences and masks ambiguous bases in nucleic acid sequences.	Perkin-Elmer Applied Biosystems, Foster City, CA.	
ABI/PARACEL FDF	A Fast Data Finder useful in comparing and annotating amino acid or nucleic acid sequences.	Perkin-Elmer Applied Biosystems, Foster City, CA; Paracel Inc., Pasadena, CA	Mismatch <50%
ABI Auto Assembler	A program that assembles nucleic acid sequences.	Perkin-Elmer Applied Biosystems, Foster City, CA.	
BLAST	A Basic Local Alignment Search Tool useful in sequence similarity search for amino acid and nucleic acid sequences. BLAST includes five functions: blastp, blastn, blastx, tblastn, and tblastx.	Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410; Altschul, S.F. et al. (1997) Nucleic Acids Res. 25: 3389-3402.	ESTs: Probability value= 1.0E-8 or less Full Length sequences: Probability value= 1.0E-10 or less
FASTA	A Pearson and Lipman algorithm that searches for similarity between a query sequence and a group of sequences of the same type. FASTA comprises as least five functions: fasta, fasta, fastx, and ssearch.	Pearson, W.R. and D.J. Lipman (1988) Proc. Natl. Acad Sci. 85:2444-2448; Pearson, W.R. (1990) Methods Enzymol. 183: 63-98; and Smith, T.F. and M. S. Waterman (1981) Adv. Appl. Math. 2:482-489.	ESTs: fasta E value=1.06E-6 Assembled ESTs: fasta Identity= 95% or greater and Match length=200 bases or greater; fastx E value=1.0E-8 or less Full Length sequences: fastx score=100 or greater
BLIMPS	A BLocks IMProved Searcher that matches a sequence against those in BLOCKS, PRINTS, DOMO, PRODOM, and PFAM databases to search for gene families, sequence homology, and structural fingerprint regions.	Henikoff, S and J.G. Henikoff, Nucl. Acid Res., 19:6565-72, 1991. J.G. Henikoff and S. Henikoff (1996) Methods Enzymol. 266:88-105; and Attwood, T.K. et al. (1997) J. Chem. Inf. Comput. Sci. 37: 417-424.	Score=1000 or greater; Ratio of Score/Strength = 0.75 or larger, and, if applicable, Probability value= 1.0E-3 or less
HMMER	An algorithm for searching a query sequence against hidden Markov model (HMM)-based databases of protein family consensus sequences, such as PFAM.	Krogh, A. et al. (1994) J. Mol. Biol., 235:1501-1531; Sonnhammer, E.L.L. et al. (1988) Nucleic Acids Res. 26:320-322.	Score=10-50 bits for PFAM hits, depending on individual protein families

	Table 5 (cont.)	cont.)	
Program	Description	Reference	Parameter Threshold
ProfileScan	An algorithm that searches for structural and sequence motifs in protein sequences that match sequence patterns defined in Prosite.	Gribskov, M. et al. (1988) CABIOS 4:61-66; Gribskov, et al. (1989) Methods Enzymol. 183:146-159; Bairoch, A. et al. (1997) Nucleic Acids Res. 25: 217-221.	Normalized quality score GCG-specified "HIGH" value for that particular Prosite motif. Generally, score=1.4-2.1.
Phred	A base-calling algorithm that examines automated sequencer traces with high sensitivity and probability.	Ewing, B. et al. (1998) Genome Res. 8:175-185; Ewing, B. and P. Green (1998) Genome Res. 8:186- 194.	
Phrap	A Phils Revised Assembly Program including SWAT and CrossMatch, programs based on efficient implementation of the Smith-Waterman algorithm, useful in searching sequence homology and assembling DNA sequences.	Smith, T.F. and M. S. Waterman (1981) Adv. Appl. Math. 2:482-489; Smith, T.F. and M. S. Waterman (1981) J. Mol. Biol. 147:195-197; and Green, P., University of Washington, Seattle, WA.	Score= 120 or greater; Match length= 56 or greater
Consed	A graphical tool for viewing and editing Phrap assemblies	Gordon, D. et al. (1998) Genome Res. 8:195-202.	
SPScan	A weight matrix analysis program that scans protein sequences for the presence of secretory signal peptides.	Nielson, H. et al. (1997) Protein Engineering 10:1-6; Claverie, J.M. and S. Audic (1997) CABIOS 12: 431-439.	Score=3.5 or greater
Motifs	A program that searches amino acid sequences for patterns that matched those defined in Prosite.	Bairoch et al. <u>supra</u> ; Wisconsin Package Program Manual, version 9, page M51-59, Genetics Computer Group, Madison, WI.	

What is claimed is:

1. A substantially purified polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-17 and fragments thereof.

5

- 2. A substantially purified variant having at least 90% amino acid sequence identity to the amino acid sequence of claim 1.
 - 3. An isolated and purified polynucleotide encoding the polypeptide of claim 1.

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- 4. An isolated and purified polynucleotide variant having at least 90% polynucleotide sequence identity to the polynucleotide of claim 3.
- 5. An isolated and purified polynucleotide which hybridizes under stringent conditions to the polynucleotide of claim 3.
 - 6. An isolated and purified polynucleotide having a sequence which is complementary to the polynucleotide of claim 3.

20

- 7. A method for detecting a polynucleotide, the method comprising the steps of:
- (a) hybridizing the polynucleotide of claim 6 to at least one nucleic acid in a sample, thereby forming a hybridization complex; and
- (b) detecting the hybridization complex, wherein the presence of the hybridization complex correlates with the presence of the polynucleotide in the sample.

- 8. The method of claim 7 further comprising amplifying the polynucleotide prior to hybridization.
- 9. An isolated and purified polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:18-34 and fragments thereof.
 - 10. An isolated and purified polynucleotide variant having at least 90% polynucleotide sequence identity to the polynucleotide of claim 9.

11. An isolated and purified polynucleotide having a sequence which is complementary to the polynucleotide of claim 9.

12. An expression vector comprising at least a fragment of the polynucleotide of claim 3.

5

- 13. A host cell comprising the expression vector of claim 12.
- 14. A method for producing a polypeptide, the method comprising the steps of:
- a) culturing the host cell of claim 13 under conditions suitable for the expression of the polypeptide; and
 - b) recovering the polypeptide from the host cell culture.
 - 15. A pharmaceutical composition comprising the polypeptide of claim 1 in conjunction with a suitable pharmaceutical carrier.

15

- 16. A purified antibody which specifically binds to the polypeptide of claim 1.
- 17. A purified agonist of the polypeptide of claim 1.

20

18. A purified antagonist of the polypeptide of claim 1.

19. A method for treating or preventing a disorder associated with decreased expression or activity of MTRP, the method comprising administering to a subject in need of such treatment an effective amount of the pharmaceutical composition of claim 15.

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20. A method for treating or preventing a disorder associated with increased expression or activity of MTRP, the method comprising administering to a subject in need of such treatment an effective amount of the antagonist of claim 18.

FIGURE 1A

FIGURE 1B

GI 2612939 GI 2612939 GI 2612939 GI 2612939 GI 2612939 GI 2612939 1720440 1720440 1720440 1720440 1720440 1720440 V R M V R L M M A IA Q Q I K Y N V K Y N G 표 요 면 Ŀ ָט RV DI R ĸ QAPGK W Z \succ Z S PPREAENOH PVRDVEQRH $\overline{\mathcal{O}}$ C G Ü C \succ × G Q Ω 区 Ø Ŋ \approx K S Ω О Ŋ S w w \mathcal{O} \mathcal{O} Ø Ø 3 3 ᄺ Щ H 工 H Ŀ П > SLLK > ᅜᅜ × Ц Д 民 > > 耳 口 α S H Ω 1 н н Ø Ø S K E K K C ی Ω LG K Д Д Д Ø K 00 E X X П ſΞι Щ L R N L Д Д \succ ADVLY X × Ц Ŏ LA ¥ > X S Ц Ц 口 Ц α <u>ი</u> K K × PM Ŋ HH П H > 田田 民 民 > H Z Д RA QIIQ > > v<u></u> ტ Д HH Н H 口 Ω AAAVA α 田田 Σ Z L H Σ 田 Ŋ ט 0 0 Ŋ α 回 HH 田 н н \succ S H А Н 니 K S တ M A Ø O L P > 니 × TH TH Д, 出 TIV j च च C Ŋ 田 $\alpha \alpha$ Ö 又 Н G Ω U Ŋ r ഥ Д S K C 回 166 196 106 241 136 331 301 211 181

FIGURE 1C

FIGURE 1D

LDQEAYSRIQAGEEKL LDERVHARICAGDFSL
KTELRKEGFDPAIVKDPLFYLDAQKGRYVP 1720440 KTRLQREGFDPRQTSDRLFFLDLKQGRYVP GI 2612939
VLEKELPLYARPIFLRLLPELHKTGTYKFQ 1720440 ELQKVLASYARPIFLRLLPQVDTTGTFKIQ GI 2612939
GVEVPGTEGRAGMAAVASPTGNCDLERFAQ 1720440 GVAVPGVEGRAGMAAIADPHSQLDPNSMYQ GI 2612939

GI 425474

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GI 425474 GI 425474 GI 425474 GI 425474 GI 425474 GI 425474 2274290 2274290 2274290 2274290 2274290 2274290 2274290 K K \triangleright Ø X S H 1 S × 民 VY 出 3 ш > Н Д K $\frac{1}{Y}$ Д > Σ Д Z Z N N Ц AAVMN H Ø O 口 H ł \succ \simeq S G Q H 四 又 口 S >H Ц ш K S > G S 团 Ø R Ŋ C α ⋈ П \mathcal{O} Ø Е > S П ſτι > Ø \Box \gt > \mathbf{Z} \gt Ξ > Ø Ę Д OI. \succ G Ŀ [I Ŋ \mathbf{Z} Ţ Ц 召 দ S S 耳 Ø ł Д Д GA 召 ď Ü Ŋ 口 P \gt α C Д 口 > K П Ц Ы H> Т S H 니 Z \mathcal{O} H H 召 Ø H \Box H Ω E \mathcal{O} > \succ K Ø Ø W Ø Н > CD S Ø Д C A Ŋ α G 伍 > G H \gt K Д × щ LF × ď C \mathcal{O} H R Д Λ \succ LYGVOGLLS [z, 口 ŀ 3 C ı Ц 又 뇌 \succ H \mathcal{O} \mathcal{O} 1 S Н Ø > Д Ц 3 又 A \mathbf{H} 又 S C S 口 S X K 口 Ø S Д Ö 口 耳 Ç 回 О S HS 团 S > RR Д > H Д Ø H Д 又 \vdash Ø 口 田 ഥ 又 S Ö × C C o a C × Н 口 Н ſτι \succ Z ָם ט L G Ы П Ŋ S K X ፲ ¥ Ω Ŋ K U Z 出 S Q Д I > S Y > Ø 二 3 区 田 니디 H \mathbb{H} ഥ <u>[1</u> Д Д 149 120 119 179 149 ದ 8 32 33 **は** 路

FIGURE 2B

GI 425474

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GI 425474 GI 425474 GI 425474 425474 GI 425474 2274290 2274290 2274290 2274290 2274290 2274290 GI Λδ O V 口 C V K Ŏ DL NGDKL ł Z > Ŋ I M E E $\overline{\mathcal{O}}$ ALVG \mathfrak{O} Ω NLSVLFG QLSLLYG Z G S > 区 Ŋ G ß Ø 됴 S Ŋ ſτι Ø П J S L 团 > Н 口 Д S Σ > \gt HŊ Ц X S S Н 又 > Ŀ Н Ŋ Z 口 Д G G Ŋ Ö Ċ K Ø Д ı \mathcal{Q} Н L P Σ S Ŋ $\boldsymbol{\mathsf{H}}$ Д K Д ц SÖ \succ ı 田田 오 요 ਸ਼ Д Ø \vdash [I Д Д 2 Д \mathbf{Z} S Ω Ω T 니 Ŋ 口 S × \succ HHH Z Y S Ц ഥ H ĹΤΙ 0 Σ Д 田 U \mathbb{R} 召 H L N S > \succ 又 闰 Е Д 口 Ø 团 Ц П > K 耳 \gt > L L Q ᅜ Н \vdash 口 Ω C > 씸 召 K > ഥ G Z Е 召 Ŀ ഥ L R > 口 Ø Z S S 口 > C 召 C > Σ Σ 니 Ъ 3 Д A Ц H K R E S O 又 S GH Ø L S 田 K X Д ᆈ × X C Ъ X Ŋ \mathcal{O} О Д S C O A C **P** A \succ Д > K α α S G E > \mathcal{O} H > Ŀ Ŋ S Ы 359 389 389 445 462 449 492

GI 425474 GI 425474 GI 425474 GI 425474 GI 425474 GI 425474 2274290 2274290 2274290 2274290 2274290 DL 口 П A D ALDA 田田 R ILVMSK ONA 되[작 Ø K ı Z V V MLA 0 V V S E O Ø > 저 O R O R LVOR K RKADL Ω **~** 口 OR 工 F 出 H 田 耳 K Ω Ø Ø Ŋ P YYN G च च G Z LAY S K R G H G α Ω > × Ω > 田 S 田 السا AK <u>G</u> H NHGY П 0 \mathbb{R} ڻ ت Ø ß Ø 田 ø S Ŀ K Ø Ø S G S S ĸ × 田 H H α \mathbb{R} C E H Ы L M A 团 Т K A 二 \succ H 디디 G Ö 国 Ø K > > Ŋ Ω Ω 田 H Н 回 团 回 O D O LM 团 Ħ H 印 Д α S Ω Д 田 X 团 DAT > Ø YDTLVG N N H 二 G 又 H > Н ¥ Д Д R ථ ථ ĸ \triangleright 田 Д 日区 G Ü Ŀ Д Ø Д KN d A NAMK ĸ 口 N Ø C ß 3 G Ø U ALI ø > H ш Ċ | | | PY Ø 区 田 召 U 629 869 599 638 899 685 539 569 552 581

FIGURE 2D

\vdash	SVGVSTSA
\leftarrow I	-STAAPSYTTSDTNRVISTFSVVDYV
31	VLLLVLS
99	FGLLLVLSLVIGLYHACRGWGRHTVGELL
61	ADRKMGCL
8	
75	RFGTQYW
8	TOYWFLGCSYFLGLLIPAHIFIPV
(
121	RLHLTSAYEYLELRFNKTVRVCGTVTF
170	YRLHLTSAY
•	
151	VVLYA
150	OMVVYMGV

FIGURE 3A

FIGURE 3B

GI 3015617 GI 3015617 GI 3015617 GI 3015617 GI 3015617 GI 3015617 2740029 2740029 2740029 2740029 2740029 2740029 ΙV OM SHT S M G M S 口 Ö LMPL Ŀı $\overline{\mathcal{O}}$ 团 Ø 4 Ъ LM > Д Д ທັ TH TH > S Ω H H 压 ĮΤι Ŀ 压 ΔX G G > Σ E H 3 3 H Д Д Ö Ŋ H Ŋ HHĤ MA Z 区 Ø Ø 口 Ω Z ט ט \mathbf{Z} Ŋ 出 田 ŋ H E U Ø 口 Ů Ţ, Ŀ Ø Н \vdash FGYGLLCL FAYGLVCL Д Ц П Ŋ Ŋ \mathcal{O} Ŋ Ŋ Ή Ŋ L 口 H \succ \succ HTNL N L TM Ц Н 3 3 ≥ Σ П Ц ri ri GLVMAGLTMA Д \succ > \Rightarrow L P J. P. Ŋ S S E G ηн Ø A Ü G S S N F Ц Ц > > RGLA RSLA လ လ ကြ လ \succ $\mathbf{\Sigma}$ × S Ø Σ H G G A Гч O O F Z Z O 도 ᆸ S Ŀ 压 ſτι 口 Ч Ŋ Ŋ K Ø S P S N P P L N S A ηц GAVG Ω Ø Ω S பப் Ŋ Ŋ S GAI 다 다 다 A Ξ Σ ø A A \vdash S N N Ø Ø K Д Ŋ Ŋ E A R E T R PL T \ AA Д 口 Д, STL Σ Ŋ Ŋ A N S Ø Ŋ Z C ď 450 420 480 360 390 451 361 391 421 481

FIGURE 3C

 $|\Lambda|$ I

628

GI 3015617 GI 3015617 GI 3015617 GI 3015617 2740029 2740029 2740029 2740029 SSSTC CSPTY Ŋ I I H 口 Д Д ᄪᄪ ᆸ TUL O G O P ΛΛ GLVH \succ 田一 A Д Н E H Ω ග A D Ω Ω $\alpha \alpha$ Д AL Z 口 Z Z <u>ග</u> AM A H Ц >ທັນ **M M** 民民 X α च च N C 1 G G ¥ $\overline{\mathcal{O}}$ X K 民 Ω G W ı Σ 口 니디 2 民 民 Ω Ö 区 召 Ċ X X Ω E E ALQ 00 0 ن \mathcal{O} Ц വ വ Ω Ω C E E Ŋ 口 П म म Z Д MGN Д $\alpha \alpha$ ద Ц Ы Ç Ц Ö 599 598 629 569 539 541 571

SEQUENCE LISTING

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<110> INCYTE PHARMACEUTICALS, INC.
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      YUE, Henry
      TANG, Y. Tom
      LAL, Preeti
      CORLEY, Neil C.
      GUEGLER, Karl J.
      BAUGHN, Mariah R.
      AZIMZAI, Yalda
      LU, Dyung Aina M.
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Cys Gln Cys Pro Ala Val Val Cys Cys Arg Cys Ser Pro Thr Gln
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                 95
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Gly Arg Leu Leu Met Val His Gly Arg Asn Ser Tyr Lys Arg Ser
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                                    175
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Pro Leu Tyr Gln Gly Phe Leu Met Val Gly Tyr Ala Thr Ile Tyr
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Thr Met Phe Pro Val Phe Ser Leu Val Leu Asp Gln Asp Val Lys
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Pro Glu Met Ala Met Leu Tyr Pro Glu Leu Tyr Lys Asp Leu Thr
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Leu Phe Glu Ser Glu Phe Val His Val Val Ala Ile Ser Phe Thr
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Gly Arg Val Ser Phe Gly Ala Phe Leu Asp Val Ala Phe Ile Thr
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			Glu	Glu 95					TOO					103
			Thr	Ser					115					120
			Gly	Lys					130					100
			Asp	Gln					145					130
			Pro	Ala					160					103
			Glu	Gln					175					100
			Ser	Lys					190					100
			Asn	Glu					205					210
			Pro	Pro					220					227
			ser	Glu 230					235					240
			Glu	Ser					250					233
			Lys	260					265					20,0
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			His	250					355					200
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				200					385					Lys 390
				205					400					Gly 405
				Ala					415					Leu 420
				Ala	Ala				430					Ile 435
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Gln Gl Phe Le Arg Le Tyr Gl Glu Ly	ly Ti eu As eu H:	rp Arg	Leu 485 Lys 500	Thr			Ile	Trp	T 011				
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Arg Le	eu H	100	Val			Leu	Ile	Val 505	Ser	His	Asp	Gln	Gly 510
Tyr Gl		is Tyr	515	Cys	Thr	Asp	Ile	Ile 520	His	Leu	Asp	Ala	Gln 525
Glu Ly	ln G			Arg	Gly	Asn	Tyr		Thr	Phe	Lys	Lys	Met 540
		ln Lys		Lys	Glu	Leu	Leu		Gln	Tyr	Glu	Lys	Gln 555
	ys Ly	ys Leu		Glu	Leu	Lys	Ala	Gly 565	Gly	Lys	Ser	Thr	Lys 570
Gln Al	la G	lu Lys		Thr	Lys	Glu	Ala	Leu 580	Thr	Arg	Lys	Gln	Gln 585
Lys Cy	ys Ai	rg Arg	Lys 590	Asn	Gln	Asp	Glu	Glu 595	Ser	Gln	Glu	Ala	Pro 600
Glu Le	eu Le	eu Lys	Arg 605	Pro	Lys	Glu	Tyr	Thr 610	Val	Arg	Phe	Thr	Phe 615
Pro As			620					625					630
Val Th			635					640					645
Asp Pl			650					655					660
Asn G			665					670	,				6/5
Leu Tì			680					685					690
Ile G			695					700					/05
Glu Tì			710					715					720
Gln As			725					730					/35
Ala H			740					745					750
Arg Va			755					760					765
Ile L			770					775				:	780
Ala L			785					790					795
Val S			800					805					810
Trp V			815					820					825
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WO 00/26245

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Ser Leu Asp Pro Ser Leu Ser Leu Phe Cys Ser Gly Ser Trp Glu
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Tyr Asp Cys Leu Pro Leu Tyr His Ser Ala Gly Asn Ile Val Gly
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Ile Gly Gln Cys Leu Leu His Gly Met Thr Val Val Ile Arg Lys
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Lys Phe Ser Ala Ser Arg Phe Trp Asp Asp Cys Ile Lys Tyr Asn
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Cys Thr Ile Val Gln Tyr Ile Gly Glu Leu Cys Arg Tyr Leu Leu
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 Ser Arg Phe His Ile Pro Gln Val Ala Glu Phe Tyr Gly Ala Thr
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PCT/US99/26048 WO 00/26245

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Val Leu Glu Lys Glu Leu Pro Leu Tyr Ala Arg Pro Ile Phe Leu
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Arg Leu Leu Pro Glu Leu His Lys Thr Gly Thr Tyr Lys Phe Gln
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Lys Thr Glu Leu Arg Lys Glu Gly Phe Asp Pro Ala Ile Val Lys
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Pro Leu Ala Pro Arg Trp Ser Pro Ser Ala Trp Cys Trp Val Gly
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Gly Ala Leu Leu Gly Pro Met Val Leu Ser Lys His Pro His Leu
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                  80
Cys Leu Val Ala Leu Cys Glu Ala Glu Glu Ala Pro Pro Ala Ser
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 Trp Gln Phe Leu His Pro His Leu Leu Val Leu Gly Val Ala Val
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Val Leu Ala Leu Gly Ala Ala Leu Val Asn Val Gln Ile Pro Leu

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PCT/US99/26048 WO 00/26245

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Leu	Leu	Gly	Gln	Leu	Val	Glu	Val	Val	ALA	Lys	TYL	1111	A- 9	165
				7 5 5					160					
His	Val	Gly	Ser	Phe	Met	Thr	Glu	Ser	GIII	Asn	пец	261		180
				170	_	3	~ 1	03	175	T.an	ጥኮዮ	Dhe	Glv	Tyr
Leu	Leu	Ile	Leu	Tyr	Gly	Val	GIn	GIA	Ten	Leu	1111	1110	0-1	195
				185			~ 1	~1	190	Mat	Δla	Val	Asp	
Leu	Val	Leu	Leu	Ser	His	Val	GIY	GIU	205	Met	AIU	V W _		210
				200			T 011	T 011	203	Gln	Asp	Tle	Thr	Phe
Arg	Arg	Ala	Leu		Ser	Ser	Leu	пеп	220	Gln				225
				215	m1	~ 1	C15	T.011	Val	Ser	Arg	Leu	Thr	Thr
Phe	Asp	Ala	Asn		Thr	GIY	GIII	пец	235		5			240
				230	T	Car	Ser	Phe	Lvs	Leu	Val	Ile	Ser	Gln
Asp	Val	Gln	GIu		гуз	Ser	261	1110	250					255
		_		245	mb~	Gln	Val	Ala	Glv	Cys	Leu	Val	Ser	Leu
				200					∠o⊃					
		_	0	260	7. ~~	T.em	Thr	Leu	Leu	Leu	Met	Val	Ala	Thr
				275					200					
		•	14ab	275	1721	Glv	Thr	Leu	Met	Gly	Ser	Gly	Leu	Arg
				~~~					233					-
_		C ~ ~	ስ አላር	Gln	Cvs	Gln	Glu	Gln	Ile	Ala	Arg	Ala	Met	Gly
				2 A E					$3 \pm 0$					
17 7	77.	λαn	Glu	Δla	Leu	Gly	Asn	Val	Arg	Thr	Val	Arg	Ala	Phe
									323					-
תות	Mat	Glu	Gln	Arg	Glu	Glu	Glu	Arg	Tyr	Gly	Ala	Glu	Leu	Glu
				275										
λla	Cvs	Ara	Cvs	Arg	Ala	Glu	Glu	Leu	Gly	Arg	Gly	Ile	Ala	Leu
				12 E A					322					
Phe	Gln	Glv	Leu	Ser	Asn	Ile	Ala	Phe	Asn	Cys	Met	Vai	Leu	GIA
				200					3/0					
Thr	Leu	Phe	Ile	Gly	Gly	Ser	Leu	Val	Ala	Gly	GIn	GIN	ьеи	390
				200					302					
Gly	Gly	Asp	Leu	Met	Ser	Phe	Leu	Val	Ala	Ser	GIII	1111	vai	Gln 405
				~ ^ ~					- 400					
Arg	Ser	Met	Ala	Asn	Leu	Ser	Val	Leu	Pne	СТУ	6111	V CL _		Arg 420
Gly	, Leu	Ser	Ala	Gly	, Ala	Arg	vai	Pne	430	. Iy-	1.00			Asn 435
														Gln 450
Pro	Cys	: Ile	Pro	Let	ı ser	GIY	GIY	Cys	445	;	_	•		450
			_	440	) 271 han	. Dho	Gln	a Asr	Val	Cvs	Phe	Ser	Tyr	Pro 465
Let	ı Arg	; Gly	/ Ser	. Val	T'ILI	PILE	GII.	, AU	460	)			_	465
				455	) . al.	. 1/21	f.er	i Tivs	Ast	Phe	Thr	Lei	Thr	Leu 480
					`					,				
		3	<b>.</b>	470	) - 1777	בות ו	Lei	. Val	Gly	, Glr	Ser	Gly	/ Gly	Gly 495
				401	_				437	,				
		-1		485	) . Sai	- T.e.1	Lei	ı Glu	ı Arc	Phe	туг	Ası	Pro	Thr 510
					٠				. JU.	,				
		<b>.</b>	770	JUC 1 Mail	J - T.e.1	ı Ast	Gly	Arc	, Ası	Lei	ı Arg	Thi	r Lev	1 Asp 525
					_				22	,				
_	- 0	. M.	n T.A1	יבכ מאמיני	- - Gla	v Glr	ı Val	l Va	l Gly	y Phe	e Ile	se:	r Gli	1 Glu 540
					^				23.					
<b>D</b>	o 170'	1 10	n Dh	رور ۱۳۱۶ م	- v Th:	r Thi	c Ile	e Met	c Gl	ASI	n Ile	e Ar	g Phe	e Gly 555
Pr	o va.	r ne	C FII	54	, <b>-</b> 5				55	0				555
				J 1	-									

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WO 00/26245
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Val Phe Val Leu Leu Val Leu Ser Leu Ala Ile Gly Leu Tyr 40 His Ala Cys Arg Gly Trp Gly Arg His Thr Val Gly Glu Leu Leu 55 Met Ala Asp Arg Lys Met Gly Cys Leu Pro Val Ala Leu Ser Leu 70 Leu Ala Thr Phe Gln Ser Ala Val Ala Ile Leu Gly Val Pro Ser 80 Glu Ile Tyr Arg Phe Gly Thr Gln Tyr Trp Phe Leu Gly Cys Cys 100 Tyr Phe Leu Gly Leu Leu Ile Pro Ala His Ile Phe Ile Pro Val 115 Phe Tyr Arg Leu His Leu Thr Ser Ala Tyr Glu Tyr Leu Glu Leu 125 Arg Phe Asn Lys Thr Val Arg Val Cys Gly Thr Val Thr Phe Ile 145 140 Phe Gln Met Val Ile Tyr Met Gly Val Val Leu Tyr Ala Pro Ser

														165
				155			<b>01</b>	Db a	160	Leu		Leu	Ser	165 Val
Leu	Ala	Leu	Asn	Ala	Val	Thr	GIY	Phe	175	пеи	עבי			180
		_	<b>~1</b>	170	37-7	Cure	Thr	Val	Tvr	Thr	Ala	Leu	Gly	Gly
				105					T 20					
T	T 1.0	ת דת	172 Î	Tle	Trn	Thr	Asp	Val	Phe	Gln	Thr	Leu	Val	Met
				200					∠∪⊃					
Phe	Leu	Glv	Gln	Leu	Ala	Val	Ile	Ile	Val	Gly	Ser	Ala	Lys	Val
				215					220			1000		
Gly	Gly	Leu	Gly	Arg	Val	Trp	Ala	Val	Ala	Ser	Gln	His	GLY	Arg
				230					230					
Ile	Ser	Gly	Phe	Glu	Leu	Asp	Pro	Asp	Pro	pne	Val	Arg	1115	255
				245	_,	<b>~</b> 7	~1.r	7727	250 Dhe	Met	Met	Leu	Ser	
Phe	Trp	Thr	Leu		Pne	GIY	GIY	Val	265	1100				270
_	<b>~</b> 3	**- 1	7 ~~	260	תות	Gln	Val	Gln		Tyr	Leu.	Ser	Ser	Arg
				つフに					280					200
	Glu	Tare	Δla	Ala	Val	Leu	Ser	Cys	Tyr	Ala	Val	Phe	Pro	Phe
				290					295					500
Gln	Gln	Val	Ser	Leu	Cys	Val	Gly	Cys	Leu	Ile	Gly	Leu	Val	Met
				205					3 I U					
Phe	Ala	Tyr	Tyr	Gln	Glu	Tyr	Pro	Met	Ser	Ile	GIn	GIN	Ala	330
				220					323					
Ala	Ala	Pro	Asp	Gln	Phe	Val	Leu	Tyr	340	vaı	Mec	vab	шса	345
				335	•	D	C3.11	Leu	Dhe	Tle	Ala	Cvs	Leu	Phe
Lys	Gly	Leu	Pro		Leu	PLO	GIY	ьец	355			- 4		360
	<b>a</b> 1	0.00	T 011	350	Thr	Tle	Ser	Ser	Ala	Phe	Asn	Ser	Leu	Ala
				265					.3/0					• -
Thr	Val	Thr	Met	Glu	Asp	Leu	Ile	Arg	Pro	Trp	Phe	Pro	Glu	Phe
				200					385					
Ser	Glu	Ala	Arg	Ala	Ile	Met	Leu	Ser	Arg	Gly	Leu	Ala	Pne	G19 405
				205					400					
Tyr	Gly	Leu	Leu			Gly	Met	Ala	415	TIE	261	361	0111	420
			_	410	71.	ת ד ת	Tlo	Ser		Phe	Glv	Met	Val	Gly
Gly	Pro	Val	Leu	GIN 425		ALA	TTE	361	430		1			435
	D	7 011	Ton	425 Glv	Len	Phe	Cvs	Leu	Gly	Met	Phe	Phe	Pro	Cys
				440					44.5					
Δla	Asn	Pro	Pro	Gly	Ala	Val	Val	Gly	Leu	Leu	Ala	Gly	Leu	Val 465
				4 5 5					460					
Met	Ala	Phe	Trp	Ile	Gly	Ile	Gly	Ser	Ile	Val	Thr	Ser	Met	Gly 480
				470					44/2					
Ser	Ser	Met	Pro	Pro	Ser	Pro	Ser	Asn	GIY	Ser	ser	Pile	361	Leu 495
				405					490					
Pro	Thr	Asn	Leu			. Ala	rnr	vai	505	1111	1100			Leu 510
			_	500	) . Dwo	ምኮፖ	- G1v	r T.e.11	Gln	Ara	Phe	Tyr	Ser	Leu 525
				C15	:				520					
<b>C</b> =	. m	. 101	, т~~	OTO TUY	Ser	- Ala	His	Asn	Ser	Thr	Thr	Val	Ile	Val 540
				E 2 C	١				232					
₹ <i>11</i> ⊃ 7	ദ്വം	Len	ı Ile	. Val	. Ser	Leu	Leu	Thr	Gly	Arg	Met	Arg	Gly	Arg
				E 1 E					550					
Ser	Leu	. Asr	n Pro	Ala	th:	: Ile	туг	Pro	Val	Leu	Pro	Lys	Lev	Leu 570
				560					565	i				570

PCT/US99/26048 WO 00/26245

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Ser Leu Leu Pro Leu Ser Cys Gln Lys Arg Leu His Cys Arg Ser
                                    580
                575
Tyr Gly Gln Asp His Leu Asp Thr Gly Leu Phe Pro Glu Lys Pro
                                    595
                590
Arg Asn Gly Val Leu Gly Asp Ser Arg Asp Lys Glu Ala Met Ala
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Leu Asp Gly Thr Ala Tyr Gln Gly Ser Ser Ser Thr Cys Ile Leu
                                    625
Gln Glu Thr Ser Leu
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Pro Gly Gly Glu Ser Asp Ala Ser Pro Glu Ala Gly Ser Gly
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Gly Gly Val Ala Leu Lys Lys Glu Ile Gly Leu Val Ser Ala
                                     40
Cys Gly Ile Ile Val Gly Asn Ile Ile Gly Ser Gly Ile Phe Val
Ser Pro Lys Gly Val Leu Glu Asn Ala Gly Ser Val Gly Leu Ala
                                     70
Leu Ile Val Trp Ile Val Thr Gly Phe Ile Thr Val Val Gly Ala
```

Leu Cys Tyr Ala Glu Leu Gly Val Thr Ile Pro Lys Ser Gly Gly

Asp Tyr Ser Tyr Val Lys Asp Ile Phe Gly Gly Leu Ala Gly Phe

Leu Arg Leu Trp Ile Ala Val Leu Val Ile Tyr Pro Thr Asn Gln

Ala Val Ile Ala Leu Thr Phe Ser Asn Tyr Val Leu Gln Pro Leu

Phe Pro Thr Cys Phe Pro Pro Glu Ser Gly Leu Arg Leu Leu Ala

Ala Ile Cys Leu Leu Leu Thr Trp Val Asn Cys Ser Ser Val

Arg Trp Ala Thr Arg Val Gln Asp Ile Phe Thr Ala Gly Lys Leu

Leu Ala Leu Ala Leu Ile Ile Ile Met Gly Ile Val Gln Ile Cys

Lys Gly Glu Tyr Phe Trp Leu Glu Pro Lys Asn Ala Phe Glu Asn

Phe Gln Glu Pro Asp Ile Gly Leu Val Ala Leu Ala Phe Leu Gln

Gly Ser Phe Ala Tyr Gly Gly Trp Asn Phe Leu Asn Tyr Val Thr

110

125

140

155

170

185

200

215

10

85

100

115

130

145

160

175

190

220

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250
                245
Glu Glu Leu Val Asp Pro Tyr Lys Asn Leu Pro Arg Ala Ile Phe
                                    265
                260
Ile Ser Ile Pro Leu Val Thr Phe Val Tyr Val Phe Ala Asn Val
                                    280
                275
Ala Tyr Val Thr Ala Met Ser Pro Gln Glu Leu Leu Ala Ser Asn
                                    295
Ala Val Ala Val Thr Phe Gly Glu Lys Leu Leu Gly Val Met Ala
                                    310
Trp Ile Met Pro Ile Ser Val Ala Leu Ser Thr Phe Gly Gly Val
                                    325
                320
Asn Gly Ser Leu Phe Thr Ser Ser Arg Leu Phe Phe Ala Gly Ala
                                    340
Arg Glu Gly His Leu Pro Ser Val Leu Ala Met Ile His Val Lys
                                    355
                350
Arg Cys Thr Pro Ile Pro Ala Leu Leu Phe Thr Cys Ile Ser Thr
                                    370
                365
Leu Leu Met Leu Val Thr Ser Asp Met Tyr Thr Leu Ile Asn Tyr
                                    385
                380
Val Gly Phe Ile Asn Tyr Leu Phe Tyr Gly Val Thr Val Ala Gly
                                    400
Gln Ile Val Leu Arg Trp Lys Lys Pro Asp Ile Pro Arg Pro Ile
                                    415
                410
Lys Ile Asn Leu Leu Phe Pro Ile Ile Tyr Leu Leu Phe Trp Ala
                                    430
                425
Phe Leu Leu Val Phe Ser Leu Trp Ser Glu Pro Val Val Cys Gly
                                    445
Ile Gly Leu Ala Ile Met Leu Thr Gly Val Pro Val Tyr Phe Leu
                                    460
                455
Gly Val Tyr Trp Gln His Lys Pro Lys Cys Phe Ser Asp Phe Ile
                                    475
                470
Glu Leu Leu Thr Leu Val Ser Gln Lys Met Cys Val Val Val Tyr
                                    490
                485
Pro Glu Val Glu Arg Gly Ser Gly Thr Glu Glu Ala Asn Glu Asp
                                    505
Met Glu Glu Gln Gln Pro Met Tyr Gln Pro Thr Pro Thr Lys
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Asp Lys Asp Val Ala Gly Gln Pro Gln Pro
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Ser	Phe	Gly	Leu	Ser 50	Val	Phe	Asn	Leu	Met 55	Asn	Ala	Ile	Met	Gly 60
Ser	Gly	Ile	Leu	Gly 65	Leu	Ala	Tyr	Val	Met 70	Ala	Asn	Thr	Gly	Val 75
Phe	Gly	Phe	Ser	Phe 80	Leu	Leu	Leu	Thr	Val 85	Ala	Leu	Leu	Ala	Ser 90
Tyr	Ser	Val	His	Leu					Cys			Thr	Ala	Val
Thr	Ser	Tyr	Glu		Leu	Gly	Leu	Phe	Ala 115		Gly	Leu	Pro	Gly 120
Lys	Leu	Val	Val		Gly	Thr	Ile	Ile	Ile 130	Gln	Asn	Ile	Gly	Ala 135
Met	Ser	Ser	Tyr		Leu	Ile	Ile	Lys	Thr 145	Glu	Leu	Pro	Ala	Ala 150
Ile	Ala	Glu	Phe		Thr	Gly	Asp	Tyr	Asn 160	Arg	Tyr	Trp	Tyr	
Asp	Gly	Gln	Thr		Leu	Ile	Ile	Ile	Cys 175	Val	Gly	Ile	Val	Phe 180
Pro	Leu	Ala	Leu		Pro	Lys	Ile	Gly	Phe 190	Leu	Gly	Tyr	Thr	Ser 195
Ser	Leu	Ser	Phe		Phe	Met	Met	Phe	Phe 205	Ala	Leu	Val	Val	Ile 210
Ile	Lys	Lys	Trp		Ile	Pro	Cys	Pro	Leu 220	Thr	Leu	Asn	Tyr	Val 225
Glu	Lys	Gly	Phe		Ile	Ser	Asn	Val	Thr 235	Asp	Asp	Cys	Lys	Pro 240
Lys	Leu	Phe	His		Ser	Lys	Glu	Ser	Ala 250	Tyr	Ala	Leu	Pro	Thr 255
Met	Ala	Phe	Ser		Leu	Cys	His	Thr	Ser 265	Ile	Leu	Pro	Ile	Tyr 270
Cys	Glu	Leu	Gln		Pro	Ser	Lys	Lys	Arg 280	Met	Gln	Asn	Val	Thr 285
Asn	Thr	Ala	Ile			Ser	Phe	Leu	Ile 295	Tyr	Phe	Ile	Ser	Ala 300
Leu	Phe	Gly	Tyr		Thr	Phe	Tyr	Asp	Lys 310	Val	Glu	Ser	Glu	Leu 315
Leu	Lys	Gly	Tyr		Lys	Tyr	Leu	Ser	His 325	Asp	Val	Val	Val	Met 330
Thr	Val	Lys	Leu		Ile	Leu	Phe	Ala	Val 340	Leu	Leu	Thr	Val	Pro 345
Leu	Ile	His	Phe		Ala	Arg	Lys	Ala	Val 355	Thr	Met	Met	Phe	Phe 360
Ser	Asn	Phe	Pro		Ser	Trp	Ile	Arg	His 370	Phe	Leu	Ile	Thr	Leu 375
Ala	Leu	Asn	lle		Ile	Val	Leu	Leu	Ala 385	Ile	Tyr	Val	Pro	Asp 390
Ile	Arg	Asn	Val		Gly	Val	Val	Gly	Ala 400	Ser	Thr	Ser	Thr	Cys 405
Leu	Ile	Phe	· Ile		Pro	Gly	Leu	Phe		Leu	Lys	Leu	Ser	Arg 420
Glu	. Asp	Phe	. Leu		Trp	Lys	Lys	Leu		Ala	Phe	Val	Leu	Leu 435
Ile	Phe	Gly	, Ile	425 Leu	v ı Val	Gly	Asn	Phe			Ala	Leu	Ile	Ile

240 445 450

Phe Asp Trp Ile Asn Lys
455

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Gly Ala Ile Leu Val Ile Thr Ala Thr Phe Leu Tyr Gly Tyr Asp
                                    310
                305
Pro Lys Pro Ala Gly Asn Pro Thr Lys Ala
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                                     10
Pro Phe Leu Phe Ser Gly Thr Val Arg Glu Asn Leu Asp Pro Gln
Gly Leu His Lys Asp Arg Ala Leu Trp Gln Ala Leu Lys Gln Cys
                                     40
                 35
His Leu Ser Glu Val Ile Thr Ser Met Gly Gly Leu Asp Gly Glu
                 50
Leu Gly Glu Gly Gly Arg Ser Leu Ser Leu Gly Gln Arg Gln Leu
Leu Cys Leu Ala Arg Ala Leu Leu Thr Asp Ala Lys Ile Leu Cys
                 80
Ile Asp Glu Ala Thr Ala Ser Val Asp Gln Lys Thr Asp Gln Leu
                                    100
                 95
Leu Gln Gln Thr Ile Cys Lys Arg Phe Ala Asn Lys Thr Val Leu
                                    115
                110
Thr Ile Ala His Arg Leu Asn Thr Ile Leu Asn Ser Asp Arg Val
                                    130
Leu Val Leu Gln Ala Gly Arg Val Val Glu Leu Asp Ser Pro Ala
                                    145
                140
Thr Leu Arg Asn Gln Pro His Ser Leu Phe Gln Gln Leu Leu Gln
                155
                                    160
Ser Ser Gln Gln Gly Val Pro Ala Ser Leu Gly Gly Pro
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Gln Val Gly Ile Val Gly Arg Thr Gly Ala Gly Lys Ser Ser Leu
Ala Ser Gly Leu Leu Arg Leu Pro Glu Ala Ala Glu Gly Gly Ile
Trp Ile Asp Gly Val Pro Ile Ala His Val Gly Leu His Thr Leu
                 80
Arg Ser Arg Ile Ser Ile Ile Pro Gln Asp Pro Ile Leu Phe Pro
                                    100
                 95
Gly Ser Leu Arg Met Asn Leu Asp Leu Leu Gln Glu His Ser Asp
                                    115
Glu Ala Ile Trp Ala Ala Leu Glu Thr Val Gln Leu Lys Ala Leu
                                     130
                125
Val Ala Ser Leu Pro Gly Gln Leu Gln Tyr Lys Cys Ala Asp Arg
                                    145
                140
Gly Glu Asp Leu Ser Val Gly Gln Lys Gln Leu Leu Cys Leu Ala
                                    160
                155
Arg Ala Leu Leu Arg Lys Thr Gln Ile Leu Ile Leu Asp Glu Ala
                                    175
                170
Thr Ala Ala Val Asp Pro Gly Thr Glu Leu Gln Met Gln Ala Met
                                    190
                185
Leu Gly Ser Trp Phe Ala Gln Cys Thr Val Leu Leu Ile Ala His
                                    205
                200
Arg Leu Arg Ser Val Met Asp Cys Ala Arg Val Leu Val Met Asp
                                    220
Lys Gly Gln Val Ala Glu Ser Gly Ser Pro Ala Gln Leu Leu Ala
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Gln Lys Gly Leu Phe Tyr Arg Leu Ala Gln Glu Ser Gly Leu Val
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                                    250
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Ser	Thr	Asp	Ala	Gly	Glu	Arg	Ala	Arg	Leu	Leu	Gln	Ser	Pro	Cys
		-		20					25					30
Val	Asp	Thr	Ala	Pro	Lys	Ser	Glu	Trp	Glu	Ala	Ser	Pro	Gly	Gly
	•			35	-				40					45
Leu	Asp	Arg	Gly	Thr	Thr	Ser	Thr	Leu	Gly	Ala	Ile	Phe	Ile	Val
	1-		- 4	50					55					60
Val	Asn	Ala	Cvs	Leu	Gly	Ala	Gly	Leu	Leu	Asn	Phe	Pro	Ala	Ala
			-1	65	_		-		70					75
Phe	Ser	Thr	Ala	Glv	Gly	Val	Ala	Ala	Gly	Ile	Ala	Leu	Gln	Met
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Cys Ser Gln Ala Ser Asn Glu Arg Thr Tyr Gln Glu Val Val Trp
                 110
Ala Val Cys Gly Lys Leu Thr Gly Val Leu Cys Glu Val Ala Ile
                                     130
                 125
Ala Val Tyr Thr Phe Gly Thr Cys Ile Ala Phe Leu Ile Ile Ile
                 140.
                                     145
Gly Asp Gln Gln Asp Lys Ile Ile Ala Val Met Ala Lys Glu Pro
                                     160
Glu Gly Ala Ser Gly Pro Trp Tyr Thr Asp Arg Lys Phe Thr Ile
                 170
Ser Leu Thr Ala Phe Leu Phe Ile Leu Pro Leu Ser Ile Pro Arg
                                     190
                 185
Glu Ile Gly Phe Gln Lys Tyr Ala Ser Phe Leu Ser Val Val Gly
                                     205
                 200
Thr Trp Tyr Val Thr Ala Ile Val Ile Ile Lys Tyr Ile Trp Pro
                                     220
                 215
Asp Lys Glu Met Thr Pro Gly Asn Ile Leu Thr Arg Pro Ala Ser
                                     235
                230
Trp Met Ala Val Phe Asn Ala Met Pro Thr Ile Cys Phe Gly Phe
                 245
                                     250
Gln Cys His Val Ser Ser Val Pro Val Phe Asn Ser Met Gln Gln
                                     265
Pro Glu Val Lys Thr Trp Gly Gly Val Val Thr Ala Ala Met Val
                 275
Ile Ala Leu Ala Val Tyr Met Gly Thr Gly Ile Cys Gly Phe Leu
                                     295
                290
Thr Phe Gly Ala Ala Val Asp Pro Asp Val Leu Leu Ser Tyr Pro
                                     310
                305
Ser Glu Asp Met Ala Val Ala Val Ala Arg Ala Phe Ile Ile Leu
                                     325
                320
Ser Val Leu Thr Ser Tyr Pro Ile Leu His Phe Cys Gly Arg Ala
                                     340
                335
Val Val Glu Gly Leu Trp Leu Arg Tyr Gln Gly Val Pro Val Glu
                                     355
                350
Glu Asp Val Gly Arg Glu Arg Arg Arg Val Leu Gln Thr Leu
                                     370
Val Trp Phe Leu Leu Thr Leu Leu Leu Ala Leu Phe Ile Pro Asp
                380
Ile Gly Lys Val Ile Ser Val Ile Gly Gly Leu Ala Ala Cys Phe
                395
                                     400
Ile Phe Val Phe Pro Gly Leu Cys Leu Ile Gln Ala Lys Leu Ser
                                     415
                410
Glu Met Glu Glu Val Lys Pro Ala Ser Trp Trp Val Leu Val Ser
                                     430
                425
Tyr Gly Val Leu Leu Val Thr Leu Gly Ala Phe Ile Phe Gly Gln
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                                     445
Thr Thr Ala Asn Ala Ile Phe Val Asp Leu Leu Ala
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Ser	Leu	Gly	Lys	Ile 365		Ala	Leu	Arg	His	•	туг	Arg	y Val	. Asp 375
Ser	Asn	Gln	Glu	Let 380		Ala	Leu	Gly	Leu 385		Asr	Leu	lle	Gly 390
Gly	Ile	Phe	Gln	Cys 395		Pro	Val	Ser	Cys 400		Met	Ser	Arg	Ser 405
Leu	Val	Gln	Glu	Ser		Gly	Gly	Asn	Ser 415		Val	. Ala	Gly	Ala 420
Ile	Ser	Ser		Phe 425		Leu	Leu	Ile			-		_	Glu 435
Leu	Phe	His	Asp	Leu 440	Pro	Lys	Ala	Val	Leu 445		Ala	Ile	Ile	Ile 450
			_	455					460		_			465
		_		470	Arg				475					480
				485	Leu				490					495
				500	Leu				505					510
				515	Leu	-			520					525
	_			530	Tyr				535					540
_				545	Ser Leu				550					555
	_		_	560	Lys	_		_	565	_				570
				575	Leu				580					585
	_		_	590	Gly				595					600
				605	Arg				610					615
				620	Asp				625					630
				635	Ala				640					645
		_		650	Asp		-		655					660
_				665	Asp				670					675
				680	Arg				685	_				690
				695	Val				700					705
				710	Thr				715					720
				725					730					735
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JUL		val		755	TIT	-3- Y	- Lou							

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Phe Arg Phe Leu His Glu Thr Gly Gly Ala Met Val Tyr Asp Lys
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Pro Pro Lys Phe Ala Met Ser Arg Glu Gln Met Ser Gln Ser Cys
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Asn Ile Gln Asp Leu Ala Pro Ser Cys Ala Gly Phe Leu Phe Gly
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Gln Ala Leu Trp Ile Phe Gly Tyr Gly Ser Leu Val Trp Arg Pro 35 40 45

Asp Phe Ala Tyr Ser Asp Ser Arg Val Gly Phe Val Arg Gly Tyr

Ser Arg Arg Phe Trp Gln Gly Asp Thr Phe His Arg Gly Ser Asp

Lys Met Pro Gly Arg Val Val Thr Leu Leu Glu Asp His Glu Gly
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Cys Thr Trp Gly Val Ala Tyr Gln Val Gln Gly Glu Gln Val Ser 95 100 105

Lys Ala Leu Lys Tyr Leu Asn Val Arg Glu Ala Val Leu Gly Gly
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Tyr Asp Thr Lys Glu Val Thr Phe Tyr Pro Gln Asp Ala Pro Asp 125 130 135

Gln Pro Leu Lys Ala Leu Ala Tyr Val Ala Thr Pro Gln Asn Pro 140 145 150

Gly Tyr Leu Gly Pro Ala Pro Glu Glu Ala Ile Ala Thr Gln Ile 155 160 165

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Arg Leu Ala Asp Phe Met Gln Leu Cys Gly Pro Gln Ala Gln Asp 185 190 195

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tgcaccaaac ctgactgtga tccagttcat taaagtcttc cccagggtgc tgctgcaqac 600
Cctacgccac cccatcttcc tgctggtggt cctgtcccag gtatgcttgt catccatggc 660
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tgatggtgca tagtccttca gaagccagcc aggcaccacc tgggcctgag agcccttcca 1260
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tgcatcagcc tccatcggcc tccagacctt caaccacagt ggcatttctg ttaacatcca 300
ggacttggcc ccgtcctgcg ccggctttct gtttggtgtg gccaacacag ccggggcctt 360
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tggacaggct cagagggtgg acctgagctc tacccatgag gacctctagc teccaacece 540
acagcetete caaggaeeca ggegeeagea geeeegggae acaggggaet cagtgtgtga 600
gacttggtca ctccatgtca gacacacgag cagagaggaa cacaaaccac tgtggagcct 660
gaagctcctt aagaagagtc cacaacagct ggtgggaggg tggggtgggc ctgggtccag 720
accaggeteg etgetetetg ggeeteagtt tececacetg ecageggget eggeeetgte 780
ctcctcacag gctggtgtgg ccgtcagggt gggtggggtt attgttagta ggcgcagcct 840
catteceace acgatetgtt ecgegtggtt ecegecaaac eteceteggt egeegtgtte 900
teegeaagee teetgeageg eeegeetgee aatgtgagge tggeaecagg etgeageete 960
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<221> misc_feature

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tegecetece ggteateate tteaceaegt tetggggeet egteggeate geegggeeet 180
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tetgetgtta cetettetgg etcategeca teetggegea getgaacece etgtteggge 300
cccagctgaa gaatgagacc atctggtacg tgcgcttcct gtgggagtga cccgccgccc 360
ccgacccagg tgcccagctc tcggaatgac tgtggctcca ctgtccctga caaccccttc 420
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Ile Val Cys Lys Thr Ala Arg Arg Asp Leu Phe Gly Leu Ser Val
Leu Ile Arg Val Arg Leu Glu Leu Arg Arg His Arg Arg Ala Gly
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Asp Thr Ile Pro Cys Ile Phe Gln Ala Val Ala Arg Arg Gln Pro
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Glu Arg Leu Ala Leu Val Asp Ala Ser Ser Gly Ile Cys Trp Thr
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100

105

95

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Gl	u Gly	⁄ Arg	Pro	Glu 140	Phe	Val	Gly	Leu	Trp 145	Leu	Gly	Leu	Ala	Lys 150
Al	a Gly	v Val	Val	Ala 155	Ala	Leu	Leu	Asn	Val 160	Asn	Leu	Arg	Arg	Glu 165
Pr	o Lei	ı Ala	Phe	Cys	Leu			Ser		Ala	Lys	Ala	Leu	Ile 180
ту	r Gly	gly	Glu	Met 185	Ala	Ala	Ala			Glu	Val	Ser	Glu	Gln 195
Le	u Gly	/ Lys	Ser	Leu 200	Leu	Lys	Phe	Cys		Gly	Asp	Leu	Gly	Pro 210
Gl	u Ser	lle	Leu	Pro 215	Asp	Thr	Gln	Leu		Asp	Pro	Met	Leu	Ala 225
Gl	u Ala	a Pro	Thr	Thr 230	Pro	Leu	Ala	Gln		Pro	Gly	Lys	Gly	Met 240
As	p Asp	Arg	Leu	Phe 245	Tyr	Ile	Tyr	Thr	Ser 250	Gly	Thr	Thr	Gly	Leu 255
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Al	a Phe	e Gly	His	His 275	Ser	Tyr	Ser	Met	Arg 280	Ala	Ala	Asp	Val	Leu 285
ту	r Asp	Cys	Leu	Pro 290	Leu	Tyr	His	Ser	Ala 295	Gly	Asn	Ile	Met	Gly 300
Va	l Gly	/ Gln	Cys	Val 305	Ile	Tyr	Gly	Leu	Thr 310	Val	Val	Leu	Arg	Lys 315
_	s Phe			Ser					325			·		330
	s Thi			Gln					340					345
	g Glr			Arg					355					360
	a Vai			365					370					3/3
	n Arg			Val					385					390
	u Cys			395					400					405
	s Gl			410					415					420
	u Va.			425					430					435
	u Gl			440					445					450
	1 Gl			455					460					465
	r Va			470					475					480
	e Arg			485					490					495
	t As			500					505				•	210
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Ile Ala Asp Pro His Ser Gln Leu Asp Pro Asn Ser Met Tyr Gln
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                                     565
Glu Leu Gln Lys Val Leu Ala Ser Tyr Ala Arg Pro Ile Phe Leu
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                                     580
Arg Leu Leu Pro Gln Val Asp Thr Thr Gly Thr Phe Lys Ile Gln
                590
                                     595
Lys Thr Arg Leu Gln Arg Glu Gly Phe Asp Pro Arg Gln Thr Ser
                605
                                     610
Asp Arg Leu Phe Phe Leu Asp Leu Lys Gln Gly Arg Tyr Val Pro
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Leu Asp Glu Arg Val His Ala Arg Ile Cys Ala Gly Asp Phe Ser
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Leu
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Phe	Asp	Val	His	Ser 200	Ser	Gly	Lys	Leu	Val 205	Glu	Ile	Ile	Gly	Ser 210
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Gly	Leu	Arg	Asn		Ile	Gln	Val	Val	Gly 235	Ser	Val	Phe	Ala	Leu 240
Leu	Ser	Ile	Ser		Thr	Leu	Thr	Ala		Leu	Ile	Gly	Cys	Leu 255
Pro	Cys	Val	Phe		Ile	Gly	Ser	Leu			Thr	Glu	Leu	
His	Ile	Ser	Arg		Val	Gln	Ser	Gln				Phe	Ala	
Leu	Ile	Asp	Glu		Phe	Ser	His	Ile		Thr	Val	Lys	Ser	
Ala	Met	Glu	Asp		Leu	Ile	Asn	Lys	-	Asn	Tyr	Asn	Val	
Lys	Ala	Lys	Met		Ser	Glu	Lys	Leu		Phe	Gly	Ile	Gly	
Phe	Gln	Gly	Leu		Asn	Leu	Thr	Leu		Gly	Val	Val	Leu	Gly 345
Val	Leu	Tyr	Val		Gly	His	Leu	Met		Arg	Gly	Glu	Leu	
Ala	Gly	His	Leu	Met 365	Ser	Phe	Leu	Ala		Thr	Gln	Thr	Leu	Gln 375
Arg	Ser	Leu	Thr		Leu	Ser	Leu	Leu		Gly	Gln	Val	Val	Arg 390
Glv	Tvr	Thr	Ala		Lvs	Arq	Ile	His		Ile	Leu	Ala	Leu	Pro
2	- 4 -				•	_			400					405
		Ile		395 Ser					400					405
Ser	Gly		Gly	395 Ser 410 Val	Ile	Pro	Ser	Ser	400 Ser 415	Ser	Ser	Leu	Val	405 Val 420
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Ser Ser Ile Tyr	Gly Lys Tyr Pro	Ile Gln Ser	Gly His Ala Arg	395 Ser 410 Val 425 Pro 440 Pro 455 Gly	Ile Asn Ser Glu	Pro Asn Ile Thr	Ser Ile Glu Ile	Ser Lys Phe Val	400 Ser 415 Glu 430 Ser 445 Leu 460	Ser Leu Asp Asn	Ser Pro Val Glu	Leu Ser Lys Leu	Val Ser Phe Ser	405 Val 420 Ser 435 Ala 450 Met 465
Ser Ser Ile Tyr	Gly Lys Tyr Pro Leu	Ile Gln Ser Asn	Gly His Ala Arg Gly	395 Ser 410 Val 425 Pro 440 Pro 455 Gly 470 Thr	Ile Asn Ser Glu Lys	Pro Asn Ile Thr	Ser Ile Glu Ile Ile	Ser Lys Phe Val	400 Ser 415 Glu 430 Ser 445 Leu 460 Leu 475	Ser Leu Asp Asn Val	Ser Pro Val Glu Gly	Leu Ser Lys Leu Gln	Val Ser Phe Ser	405 Val 420 Ser 435 Ala 450 Met 465 Gly 480
Ser Ser Ile Tyr Phe Ala	Gly Lys Tyr Pro Leu Gly	Ile Gln Ser Asn Pro	Gly His Ala Arg Gly Ser	395 Ser 410 Val 425 Pro 440 Pro 455 Gly 470 Thr 485 Glu	Ile Asn Ser Glu Lys Val	Pro Asn Ile Thr Val Val	Ser Ile Glu Ile Ile Ser	Ser Lys Phe Val Ala Leu	400 Ser 415 Glu 430 Ser 445 Leu 460 Leu 475 Leu 490	Ser Leu Asp Asn Val Glu	Ser Pro Val Glu Gly Arg	Leu Ser Lys Leu Gln Phe	Val Ser Phe Ser Ser	405 Val 420 Ser 435 Ala 450 Met 465 Gly 480 Asp 495
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Ser Ser Ile Tyr Phe Ala Pro	Gly Lys Tyr Pro Leu Gly Ile Asn	Ile Gln Ser Asn Pro Lys Ser	Gly His Ala Arg Gly Ser Gly Asn	395 Ser 410 Val 425 Pro 440 Pro 455 Gly 470 Thr 485 Glu 500 Tyr 515	Ile Asn Ser Glu Lys Val Ile Leu	Pro Asn Ile Thr Val Val Leu Arg	Ser Ile Glu Ile Ile Ser Leu Ser	Ser Lys Phe Val Ala Leu Asn Lys	400 Ser 415 Glu 430 Ser 445 Leu 475 Leu 490 Gly 505 Leu 520	Ser Leu Asp Asn Val Glu Asp Ile	Ser Pro Val Glu Gly Arg Lys	Leu Ser Lys Leu Gln Phe Leu	Val Ser Phe Ser Tyr Thr	405 Val 420 Ser 435 Ala 450 Met 465 Gly 480 Asp 495 Asn 510 Ser 525
Ser Ser Ile Tyr Phe Ala Pro Phe Gln	Cly Lys Tyr Pro Leu Cly Ile Asn Clu	Ile Gln Ser Asn Pro Lys Ser Val	Gly His Ala Arg Gly Ser Gly Asn Gln	395 Ser 410 Val 425 Pro 440 Pro 455 Gly 470 Thr 485 Glu 500 Tyr 515 Ile 530 Asp	Ile Asn Ser Glu Lys Val Ile Leu Phe	Pro Asn Ile Thr Val Val Leu Arg	Ser Ile Glu Ile Ile Ser Leu Ser Ala	Ser Lys Phe Val Ala Leu Asn Lys Ser	400 Ser 415 Glu 430 Ser 445 Leu 460 Leu 475 Leu 490 Gly 505 Leu 520 Ile 535	Ser Leu Asp Asn Val Glu Asp Ile Arg	Ser Pro Val Glu Gly Arg Lys Gly	Leu Ser Lys Leu Gln Phe Leu Tyr	Val Ser Phe Ser Tyr Thr Ile Ile	405 Val 420 Ser 435 Ala 450 Met 465 Gly 480 Asp 495 Asn 510 Ser 525 Arg 540
Ser Ser Ile Tyr Phe Ala Pro Phe Gln Phe	Cly Lys Tyr Pro Leu Cly Ile Asn Clu Cly	Ile Gln Ser Asn Pro Lys Ser Val	Gly His Ala Arg Gly Ser Gly Asn Gln Phe	395 Ser 410 Val 425 Pro 440 Pro 455 Gly 470 Thr 485 Glu 500 Tyr 515 Ile 530 Asp 545 Ala	Ile Asn Ser Glu Lys Val Ile Leu Phe Ala	Pro Asn Ile Thr Val Val Leu Arg Asn Thr	Ser Ile Glu Ile Ile Ser Leu Ser Ala Asp	Ser Lys Phe Val Ala Leu Asn Lys Ser Glu	400 Ser 415 Glu 430 Ser 445 Leu 475 Leu 490 Gly 505 Leu 535 Glu 550	Ser Leu Asp Asn Val Glu Asp Ile Arg Val	Ser Pro Val Glu Gly Arg Lys Gly Glu Glu	Leu Ser Lys Leu Gln Phe Leu Tyr Asn Glu	Val Ser Phe Ser Ser Tyr Thr Ile Ile Ala	405 Val 420 Ser 435 Ala 450 Met 465 Gly 480 Asp 510 Ser 525 Arg 540 Ala 555
Ser Ser Ile Tyr Phe Ala Pro Phe Gln Phe Lys	Gly Lys Tyr Pro Leu Gly Ile Asn Glu Gly Leu	Ile Gln Ser Asn Pro Lys Ser Val Pro Arg	Gly His Ala Arg Gly Ser Gly Asn Gln Phe	395 Ser 410 Val 425 Pro 440 Pro 455 Gly 470 Thr 485 Glu 500 Tyr 515 Ile 530 Asp 545 Ala 560 Leu	Ile Asn Ser Glu Lys Val Ile Leu Phe Ala His	Pro Asn Ile Thr Val Val Leu Arg Asn Thr	Ser Ile Glu Ile Ile Ser Leu Ser Ala Asp Phe	Ser Lys Phe Val Ala Leu Asn Lys Ser Glu Ile	400 Ser 415 Glu 430 Ser 445 Leu 460 Leu 475 Leu 490 Gly 505 Leu 520 Ser 565	Ser Leu Asp Asn Val Glu Asp Ile Arg Val Asn	Ser Pro Val Glu Gly Arg Lys Gly Glu Glu Asp	Leu Ser Lys Leu Gln Phe Leu Tyr Asn Glu Leu	Val Ser Phe Ser Tyr Thr Ile Ile Ala Pro	405 Val 420 Ser 435 Ala 450 Met 465 Gly 480 Asp 495 Asn 510 Ser 525 Arg 540 Ala 555 Tyr 570
Ser Ser Ile Tyr Phe Ala Pro Phe Gln Phe Lys Gly	Cly Lys Tyr Pro Leu Gly Ile Asn Glu Gly Leu Tyr	Ile Gln Ser Asn Pro Lys Ser Val Pro Arg	Gly His Ala Arg Gly Ser Gly Asn Gln Phe Tyr	395 Ser 410 Val 425 Pro 440 Pro 455 Gly 470 Thr 485 Glu 500 Tyr 515 Asp 545 Ala 560 Leu 575	Ile Asn Ser Glu Lys Val Ile Leu Phe Ala His	Pro Asn Ile Thr Val Val Leu Arg Asn Thr Asp	Ser Ile Glu Ile Ile Ser Leu Ser Ala Asp Phe Gln	Ser Lys Phe Val Ala Leu Asn Lys Ser Glu Ile Gly	400 Ser 415 Glu 430 Ser 445 Leu 460 Leu 475 Leu 505 Leu 516 510 565 Thr 580	Ser Leu Asp Asn Val Glu Asp Ile Arg Val Asn Gly	Ser Pro Val Glu Gly Arg Lys Gly Glu Glu Asp	Leu Ser Lys Leu Gln Phe Leu Tyr Asn Glu Leu Ile	Val Ser Phe Ser Tyr Thr Ile Ile Ala Pro Ala	405 Val 420 Ser 435 Ala 450 Met 465 Gly 480 Asp 510 Ser 525 Arg 540 Ala 555 Tyr 570 Gly 585

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605
                                       610
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  Leu Asp Thr Glu Ser Glu Ala Lys Val Gln Asn Ala Leu Asn Asn
                  620
                                       625
  Ala Met Lys Gly Arg Thr Val Leu Ile Ile Ala His Arg Leu Ser
                  635
                                      640
  Thr Val Arg Lys Ala Asp Leu Ile Leu Val Met Ser Lys Gly Gln
                  650
                                      655
  Ile Val Glu Lys Gly Thr His Ser Glu Leu Met Ala Asn His Gly
                  665
                                      670
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  Tyr Tyr Tyr Asn Leu Val Gln Arg Gln Glu Gly Cys Asp Val Phe
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 Thr Asn Arg Val Ile Ser Thr Phe Ser Val Val Asp Tyr Val Val
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                                       25
 Phe Gly Leu Leu Val Leu Ser Leu Val Ile Gly Leu Tyr His
Ala Cys Arg Gly Trp Gly Arg His Thr Val Gly Glu Leu Leu Met
                                      55
Ala Asp Arg Lys Met Gly Cys Leu Pro Val Ala Leu Ser Leu Leu
                  65
                                      70
Ala Thr Phe Gln Ser Ala Val Ala Ile Leu Gly Gly Pro Ala Glu
                  80
Ile Tyr Arg Phe Gly Thr Gln Tyr Trp Phe Leu Gly Cys Ser Tyr
                                     100
Phe Leu Gly Leu Leu Ile Pro Ala His Ile Phe Ile Pro Val Phe
                 110
                                     115
Tyr Arg Leu His Leu Thr Ser Ala Tyr Glu Tyr Leu Glu Leu Arg
                 125
                                     130
Phe Asn Lys Ala Val Arg Ile Cys Gly Thr Val Thr Phe Ile Phe
                                     145
Gln Met Val Val Tyr Met Gly Val Ala Leu Tyr Ala Pro Ser Leu
                 155
                                     160
Ala Leu Asn Ala Val Thr Gly Phe Asp Leu Trp Leu Ser Val Leu
                170
                                     175
Ala Leu Gly Ile Val Cys Asn Ile Tyr Thr Ala Leu Gly Gly Leu
                185
Lys Ala Val Ile Trp Thr Asp Val Phe Gln Thr Leu Ile Met Phe
                                    205
Leu Gly Gln Leu Val Val Ile Ile Val Gly Ala Ala Lys Val Gly
                215
                                    220
Gly Leu Gly His Val Trp Ala Val Ala Ser Gln His Gly Leu Ile
                230
                                    235
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•••	00,20	-45												·
Ser	Gly	Ile	Glu	Leu 245	Asp	Pro	Asp	Pro	Phe 250	Val	Arg	His	Thr	Phe 255
Trp	Thr	Leu	Ala	Phe	Gly	Gly	Val	Phe	Met	Met	Leu	Ser	Leu	Tyr - 270
Gly	Val	Asn	Gln	260 Ala	Gln	Val	Gln	Arg	265 Tyr	Leu	Ser	Ser	His	Ser
				275					280					285
Glu	Lys	Ala	Ala	Val 290	Leu	Ser	Cys	Tyr	Ala 295	Val	Phe	Pro	Cys	Gln 300
din	17 = 1	Δla	Leu	Cvs	Met	Ser	Cvs	Leu	Ile	Gly	Leu	Val	Met	Phe
GIII	٠٠٠			305	7.7	-			310					315
בומ	ር። ጥኒፖን	Tyr	Lvs	Lvs	Tvr	Ser	Met	Ser	Pro	Gln	Gln	Glu	Gln	Ala
	-1-	-1-	2	320	•				325					330
Δla	Pro	Asp	Gln		Val	Leu	Tyr	Phe	Val	Met	Asp	Leu	Leu	Lys
				335			-		340					345
Asn	Met	Pro	Glv	Leu	Pro	Gly	Leu	Phe	Val	Ala	Cys	Leu	Phe	Ser
тыр		110		350		•			355					360
Glv	Ser	Leu	Ser		Ile	Ser	Ser	Ala	Phe	Asn	Ser	Leu	Ala	Thr
0.7	DC1			365					370					375
Val	Thr	Met.	Glu		Leu	Ile	Gln	Pro	Trp	Phe	Pro	Gln	Leu	Thr
VUI	****			380					385					390
Glu	Thr	Ara	Ala		Met	Leu	Ser	Arg	Ser	Leu	Ala	Phe	Ala	Tyr
014		••- 9		395				_	400					405
Glv	Leu	Val	Cvs	Leu	Gly	Met	Ala	Tyr	Val	Ser	Ser	His	Leu	Gly
Cry	200		-1-	410				-	415					420
Ser	Va1	Leu	Gln	Ala	Ala	Leu	Ser	Ile	Phe	Gly	Met	Val	Gly	Gly
0-1	V41			425					430	-				435
Pro	Leu	Leu	Glv	Leu	Phe	Cys	Leu	Gly	Met	Phe	Phe	Pro	Cys	Ala
110	1100	200		440		•		_	445					450
Δεη	Pro	Leu	Glv	Ala	Ile	Val	Gly	Leu	Leu	Thr	Gly	Leu	Thr	Met
11011				455			•		460					465
Δla	Phe	Trn	Ile		Ile	Gly	Ser	Ile	Val	Ser	Arg	Met	Ser	Ser
73.4.4				470		•			475					480
Δla	Ala	Ala	Ser	Pro	Pro	Leu	Asn	Gly	Ser	Ser	Ser	Phe	Leu	Pro
AIU				485				_	490					495
Ser	Asn	Leu	Thr	Val	Ala	Thr	Val	Thr	Thr	Leu	Met	Pro	Ser	Thr
				500					505					510
Len	Ser	Lvs	Pro	Thr	Gly	Leu	Gln	Gln	Phe	Tyr	Ser	Leu	Ser	Tyr
				515					520					525
Len	Tro	Tvr	Ser	Ala	His	Asn	Ser	Thr	Thr	Val	Ile	Ala	Val	Gly
	F	-,,		530					535					540
Leu	Ile	Val	Ser	Leu	Leu	Thr	Gly	Gly	Met	Arg	Gly	Arg	Ser	Leu
				545					550					555
Asn	Pro	Glv	Thr	Ile	Tyr	Pro	Val	Leu	Pro	Lys	Leu	Leu	Ala	Leu
				560					565	100				570
Leu	Pro	Leu	Ser		Gln	Lys	Arg	Leu	Cys	Trp	Arg	Ser	His	Asn
				575					580					585
Gln	azĀ	Ile	Pro	Val	Val	Thr	Asn	Leu	Phe	Pro	Glu	Lys	Met	Gly
				590					595					600
Asn	Glv	Ala	Leu	Gln	Asp	Ser	Arg	Asp	Lys	Glu	Arg	Met	Ala	Glu
				605					610					612
Asp	Glv	Leu	Val	His	Gln	Pro	Cys	Ser	Pro	Thr	Tyr	Ile	Val	Gln
	- 2			620			-		625					630
Glu	Thr	Ser	Leu											

<210> 38 <211> 507 <212> PRT <213> Homo sapiens

<300>
<308> GenBank ID No: g3639058

320

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Val Gly Leu Ser Cys Phe Gly Ser Val Asn Gly Ser Leu Phe Thr

325

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340
                335
Ser Ser Arg Leu Phe Phe Val Gly Ser Arg Glu Gly His Leu Pro
                                     355
Ser Ile Leu Ser Met Ile His Pro Gln Leu Leu Thr Pro Val Pro
                365
Ser Leu Val Phe Thr Cys Val Met Thr Leu Leu Tyr Ala Phe Ser
                                     385
                380
Lys Asp Ile Phe Ser Val Ile Asn Phe Phe Ser Phe Phe Asn Trp
                                    400
                                                         405
Leu Cys Val Ala Leu Ala Ile Ile Gly Met Ile Trp Leu Arg His
                410
                                     415
Arg Lys Pro Glu Leu Glu Arg Pro Ile Lys Val Asn Leu Ala Leu
                                     430
                425
Pro Val Phe Phe Ile Leu Ala Cys Leu Phe Leu Ile Ala Val Ser
                440
                                     445
Phe Trp Lys Thr Pro Val Glu Cys Gly Ile Gly Phe Thr Ile Ile
                 455
Leu Ser Gly Leu Pro Val Tyr Phe Phe Gly Val Trp Trp Lys Asn
                                     475
                470
Lys Pro Lys Trp Leu Leu Gln Gly Ile Phe Ser Thr Thr Val Leu
                                                         495
                                     490
                 485
Cys Gln Lys Leu Met Gln Val Val Pro Gln Glu Thr
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<210> 39 <211> 504 <212> PRT <213> Homo sapiens

<300>
<308> GenBank ID No: g1840045

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Leu Gln Asn Ile Gly Ala Met Ser Ser Tyr Leu Tyr Ile Ile Lys
 Ser Glu Leu Pro Leu Val Ile Gln Thr Phe Leu Asn Leu Glu Glu
                 170
                                      175
 Lys Thr Ser Asp Trp Tyr Met Asn Gly Asn Tyr Leu Val Ile Leu
                 185
                                      190
 Val Ser Val Thr Ile Ile Leu Pro Leu Ala Leu Met Arg Gln Leu
                                      205
 Gly Tyr Leu Gly Tyr Ser Ser Gly Phe Ser Leu Ser Cys Met Val
                 215
                                     220
 Phe Phe Leu Ile Ala Val Ile Tyr Lys Lys Phe His Val Pro Cys
                 230
                                     235
 Pro Leu Pro Pro Asn Phe Asn Asn Thr Thr Gly Asn Phe Ser His
                 245
                                     250
 Val Glu Ile Val Lys Glu Lys Val Gln Leu Gln Val Glu Pro Glu
 Ala Ser Ala Phe Cys Thr Pro Ser Tyr Phe Thr Leu Asn Ser Gln
                 275
                                     280
 Thr Ala Tyr Thr Ile Pro Ile Met Ala Phe Ala Phe Val Cys His
                                     295
Pro Glu Val Leu Pro Ile Tyr Thr Glu Leu Lys Asp Pro Ser Lys
                                     310
Lys Lys Met Gln His Ile Ser Asn Leu Ser Ile Ala Val Met Tyr
                                     325
Ile Met Tyr Phe Leu Ala Ala Leu Phe Gly Tyr Leu Thr Phe Tyr
                335
                                     340
Asn Gly Val Glu Ser Glu Leu Leu His Thr Tyr Ser Lys Val Asp
                350
                                     355
Pro Phe Asp Val Leu Ile Leu Cys Val Arg Val Ala Val Leu Thr
                                     370
Ala Val Thr Leu Thr Val Pro Ile Val Leu Phe Pro Val Arg Arg
                380
                                     385
Ala Ile Gln Gln Met Leu Phe Pro Asn Gln Glu Phe Ser Trp Leu
                395
                                     400
Arg His Val Leu Ile Ala Val Gly Leu Leu Thr Cys Ile Asn Leu
                                     415
Leu Val Ile Phe Ala Pro Asn Ile Leu Gly Ile Phe Gly Val Ile
                                    430
Gly Ala Thr Ser Ala Pro Phe Leu Ile Phe Ile Phe Pro Ala Ile
                440
                                    445
Phe Tyr Phe Arg Ile Met Pro Thr Glu Lys Glu Pro Ala Arg Ser
                455
                                    460
Thr Pro Lys Ile Leu Ala Leu Cys Phe Ala Met Leu Gly Phe Leu
                470
                                    475
Leu Met Thr Met Ser Leu Ser Phe Ile Ile Ile Asp Trp Ala Ser
                485
                                    490
Gly Thr Ser Arg His Gly Gly Asn His
                500
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<210> 40

<211> 393

<212> PRT

<213> Homo sapiens

<300>

<308> GenBank ID No: g1526438

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380

390

385

Leu Val Lys

<210> 41 <211> 893 <212> PRT

<213> Homo sapiens

<308> GenBank ID No: g3335175

<400> 41

His Val Gln Asp Phe Thr Ala Phe Trp Asp Lys Ala Ser Glu Thr Pro Thr Leu Gln Gly Leu Ser Phe Thr Val Arg Pro Gly Glu Leu Leu Ala Val Val Gly Pro Val Gly Ala Gly Lys Ser Ser Leu Leu Ser Ala Val Leu Gly Glu Leu Ala Pro Ser His Gly Leu Val Ser 55 Val His Gly Arg Ile Ala Tyr Val Ser Gln Gln Pro Trp Val Phe Ser Gly Thr Leu Arg Ser Asn Ile Leu Phe Gly Lys Lys Tyr Glu 85 Lys Glu Arg Tyr Glu Lys Val Ile Lys Ala Cys Ala Leu Lys Lys 95 100 Asp Leu Gln Leu Leu Glu Asp Gly Asp Leu Thr Val Ile Gly Asp 110 Arg Gly Thr Thr Leu Ser Gly Gly Gln Lys Ala Arg Val Asn Leu 130 Ala Arg Ala Val Tyr Gln Asp Ala Asp Ile Tyr Leu Leu Asp Asp 140 145 Pro Leu Ser Ala Val Asp Ala Glu Val Ser Arg His Leu Phe Glu 160 Leu Cys Ile Cys Gln Ile Leu His Glu Lys Ile Thr Ile Leu Val 175 Thr His Gln Leu Gln Tyr Leu Lys Ala Ala Ser Gln Ile Leu Ile 185 190 Leu Lys Asp Gly Lys Met Val Gln Lys Gly Thr Tyr Thr Glu Phe 200 205 Leu Lys Ser Gly Ile Asp Phe Gly Ser Leu Leu Lys Lys Asp Asn 215 225 Glu Glu Ser Glu Gln Pro Pro Val Pro Gly Thr Pro Thr Leu Arg Asn Arg Thr Phe Ser Glu Ser Ser Val Trp Ser Gln Gln Ser Ser 250 Arg Pro Ser Leu Lys Asp Gly Ala Leu Glu Ser Gln Asp Thr Glu 260 265 Asn Val Pro Val Thr Leu Ser Glu Glu Asn Arg Ser Glu Gly Lys Val Gly Phe Gln Ala Tyr Lys Asn Tyr Phe Arg Ala Gly Ala His 295 Trp Ile Val Phe Ile Phe Leu Ile Leu Leu Asn Thr Ala Ala Gln

WU	00/20	243												
				305					310					315
Val	Ala	Tvr	Val	Leu	Gln	Asp	Trp	Trp	Leu	Sér	Tyr	Trp	Ala	Asn
				320					325					330
Lvs	Gln	Ser	Met	Leu	Asn	Val	Thr	Val	Asn	Gly	Gly	Gly	Asn	Val
				335					340					343
Thr	Glu	Lys	Leu	Asp	Leu	Asn	Trp	Tyr	Leu	Gly	Ile	Tyr	Ser	Gly
				350					355					360
Leu	Thr	Val	Ala	Thr	Val	Leu	Phe	Gly	Ile	Ala	Arg	Ser	Leu	Leu
				365			· .		370					3/5
Val	Phe	Tyr	Val	Leu	Val	Asn	Ser	Ser	Gln	Thr	Leu	His	Asn	Lys
				380					385					390
Met	Phe	Glu	Ser	Ile	Leu	Lys	Ala	Pro	Val	Leu	Phe	Phe	Asp	Arg
				395					400					405
Asn	Pro	Ile	Gly	Arg	Ile	Leu	Asn	Arg	Phe	Ser	Lys	Asp	Ile	GIY
				410					415					420
His	Leu	Asp	Asp	Leu	Leu	Pro	Leu	Thr	Phe	Leu	Asp	Phe	Ile	GID
				425					430					433
Thr	Leu	Leu	Gln	Val	Val	Gly	Val	Val	Ser	Val	Ala	val	Ala	vai
				440					445					450
Ile	Pro	Trp	Ile	Ala	Ile	Pro	Leu	Val	Pro	Leu	GIA	TTE	Ile	ACE
				455				_	460	_	•	7	77-7	465
Ile	Phe	Leu	Arg	Arg	Tyr	Phe	Leu	Glu	Thr	Ser	Arg	Asp	Val	480
				470					475			***	T 011	
Arg	Leu	Glu	Ser		Thr	Arg	Ser	Pro	vai	Pne	Ser	птр	Leu	495
				485		_	_,	-1.	490	210	Ф. т.	Larc	λla	
Ser	Ser	Leu	Gln		Leu	Trp	Thr	IIe	Arg	Ala	TYL	пур	Ala	510
				500		_,	_		505	~1-	7 cn	Len	Hic	
Glu	Arg	Cys	Gln		Leu	Phe	Asp	Ala	520	GIII	Asp	, LICU	His	525
				515	_,		<b></b>	mb		71	Trn	Phe	Δla	
Glu	Ala	Trp	Phe		Pne	Leu	THE	TILL	535	Arg	TIP	1110	Ala	540
				530	<b>a</b>	77.	Mot	Dha		Tle	Tle	Val	Ala	
Arg	Leu	Asp	Ala		Cys	Ald	Mec	PHE	550	110			Ala	555
	_	_	-1-	545	71 T	T	Thr	T.e.11		Δla	Glv	Gln	Val	
Gly	Ser	Leu	TTE		Ala	пåр	1111	Пец	565		1			570
_	- 3		0	560	777	T.011	Thr	Leu		Glv	Met	Phe	Gln	Trp
Leu	Ala	Leu		575		neu	1111		580	1				585
<b>~</b>	**- 1	7	~1 m	2/2	712	Glu	Val	Glu			Met	Ile	Ser	Val
Cys	vaı	Arg	GIII	590		GIU	Vul	014	595					600
a1	7	110 1	т1.	230	ጥኒታን	Thr	Asp	Leu		Lvs	Glu	Ala	Pro	Trp
GIU	Arg	vai	TTE	605					610	•				615
<b>a</b> 1	Шт гэс	Cl n	Tare	220	Pro	Pro	Pro	Ala	Trp	Pro	His	Glu	Gly	Val
GIU	ıyı	GIII	пуз	620					625					630
T1.0	710	Dho	λen	Den	Val	Asn	Phe	Met		Ser	Pro	Gly	Gly	Pro
TTE	TTE	Pne	Азр	635					640					645
	7727	T 011	Tare	Uic	T.e.11	Thr	Ala	Leu			Ser	Gln	Glu	Lys
ьeu	vai	ьeu	пуз	650					655	•				660
770]	Glv	710	Val	Glv	Ara	Thr	Glv	Ala			Ser	Ser	Leu	Ile
Vai	Gry	110	val	665			1		670	-				675
202	Ale	t.e.r	Pha	Ara	I.e.1	Ser	Glu	Pro	Glu	Gly	Lys	Ile	Trp	Ile
261	AT a	<u></u> u		680					685	-	-			690
Δen	Live	Tle	Leu	Thr	Thr	Glu	Ile	Gly	Leu	His	Asp	Leu	Arg	Lys
				695					700					/05
Taye	Met	Ser	Ile	Ile	Pro	Gln	Glu	Pro	Val	Leu	Phe	Thr	Gly	Thr
درد				710					715					720

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Met Arg Lys Asn Leu Asp Pro Phe Lys Glu His Thr Asp Glu Glu
Leu Trp Asn Ala Leu Gln Glu Val Gln Leu Lys Glu Thr Ile Glu
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                                    745
Asp Leu Pro Gly Lys Met Asp Thr Glu Leu Ala Glu Ser Gly Ser
                755
                                    760
Asn Phe Ser Val Gly Gln Arg Gln Leu Val Cys Leu Ala Arg Ala
                770
                                    775
Ile Leu Arg Lys Asn Gln Ile Leu Ile Ile Asp Glu Ala Thr Ala
                                    790
                                                         795
                785
Asn Val Asp Pro Arg Thr Asp Glu Leu Ile Gln Lys Lys Ile Arg
                800
                                    805
Glu Lys Phe Ala His Cys Thr Val Leu Thr Ile Ala His Arg Leu
                                    820
                815
Asn Thr Ile Ile Asp Ser Asp Lys Ile Met Val Leu Asp Ser Gly
Arg Leu Lys Glu Tyr Asp Glu Pro Tyr Val Leu Leu Gln Asn Lys
                                    850
Glu Ser Leu Phe Tyr Lys Met Val Gln Gln Leu Gly Lys Ala Glu
                860
                                    865
Ala Ala Ala Leu Thr Glu Thr Ala Lys Gln Val Ile Leu Gln Lys
                875
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<300>

<308> GenBank ID No: g1279457

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Leu	Val	Gly	Phe	Ser		Ser	Ala	Ala	Leu 160	Glņ	Val	Thr	Gln	Thr 165
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Gln	Gly	Gly	Gln		Glu	Phe	Arg	Asp	Phe	Gly	Leu	Arg	Tyr	
Pro	Glu	Leu	Pro		Ala	Val	Gln	Gly		Ser	Phe	Lys	Ile	His
Ala	Gly	Glu	Lys		Gly	Ile	Val	Gly		Thr	Gly	Ala	Gly	
Ser	Ser	Leu	Ala		Gly	Leu	Leu	Arg		Gln	Glu	Ala	Ala	
Gly	Gly	Ile	Trp		Asp	Gly	Val	Pro		Ala	His	Val	Gly	
His	Thr	Leu	Arg	275 Ser	Arg	Ile	Ser	Ile		Pro	Gln	Asp	Pro	
Leu	Phe	Pro	Gly	290 Ser	Leu	Arg	Met	Asn		Asp	Leu	Leu	Gln	
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Asp	Glu	Ala	Thr	365 Ala	Ala	Val	Asp	Pro	370 Gly	Thr	Glu	Leu	Gln	375 Met
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- T J														

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IN LEKNATONAL SEARCH REP PCT/US 99/26048 CLASSIFICATION OF SUBJECT MATTER PC 7 C12N15/12 C07k A61K38/17 C07K16/18 C12Q1/68 CO7K14/47 C12N1/21 According to International Patent Classification (IPC) or to both national classification and IPC B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) CO7K C12N A61K C12Q Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) C. DOCUMENTS CONSIDERED TO BE RELEVANT Citation of document, with indication, where appropriate, of the relevant passages Relevant to claim No. 1-16,19, DATABASE GENEMBL [Online] X 20 20 February 1998 (1998-02-20) HALLECK M.S. ET AL: "Homo sapiens putative ATPase mRNA, partial cds." XP002132693 Accession No. U78978 -& HALLECK, M.S. ET AL.: "Multiple А members of a third subfamily of P-type ATPases identified by genomic sequences and ESTs." GENOME RESEARCH, vol. 8, no. 4, April 1998 (1998-04), pages 354-361, XP002132690 figure 2B page 357 -/--Patent family members are listed in annex. Further documents are listed in the continuation of box C. Х Special categories of cited documents: T later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the "A" document defining the general state of the art which is not considered to be of particular relevance invention "X" document of particular relevance; the claimed invention *E* earlier document but published on or after the international cannot be considered novel or cannot be considered to filing date involve an inventive step when the document is taken alone "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another "Y" document of particular relevance; the claimed invention citation or other special reason (as specified) cannot be considered to involve an inventive step when the document is combined with one or more other such docu- O document referring to an oral disclosure, use, exhibition or ments, such combination being obvious to a person skilled other means in the art. *P* document published prior to the international filing date but "&" document member of the same patent family later than the priority date claimed Date of mailing of the international search report Date of the actual completion of the international search 2 O. JUNI. 2000 15 March 2000 Authorized officer Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentiaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,

ALCONADA RODRIG.., A

Fax: (+31-70) 340-3016

## INTERNATIONAL SEARCH REPORT

PCT/US 99/26048

C.(Continua	ation) DOCUMENTS CONSIDERED TO BE RELEVANT	1/03 33/20048
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DATABASE GENEMBL [Online] 27 May 1998 (1998-05-27) STANCHI,F.: "Homo sapiens mRNA for putative ATPase, partial" XP002132694 Accession AJ006268	1-16,19,
A	ALLIKMETS R ET AL: "Characterization of the human ABC superfamily: isolation and mapping of 21 new genes using the expressed sequence tags database." HUMAN MOLECULAR GENETICS, (1996 OCT) 5 (10) 1649-55., XP002132691 figures 2,3; table I	·
A	MASTROBERARDINO L ET AL: "Amino-acid transport by heterodimers of 4F2hc/CD98 and members of a permease family." NATURE, (1998 SEP 17) 395 (6699) 288-91., XP002132692 the whole document	
A	SARDET C ET AL: "Molecular cloning, primary structure, and expression of the human growth factor-activatable Na+/H+ antiporter." CELL, (1989 JAN 27) 56 (2) 271-80., XP000876824 the whole document	
	*****	

### INTERNATIONAL SEARCH REPORT

onal application No. PCT/US 99/26048

BoxI	Observations where certain claims were found unsearchable (Continuation of Item 1 of first sheet)
This Inte	ernational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1 X	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
	Although claims 19 and 20 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2. X	Claims Nos.: 17, 18 because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
	see FURTHER INFORMATION sheet PCT/ISA/210
з. 🗌	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of Item 2 of first sheet)
This Inte	mational Searching Authority found multiple inventions in this international application, as follows:
1.	As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2.	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. X	No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:  1-16,19-20 (partially)
Remark	The additional search fees were accompanied by the applicant's protest.  No protest accompanied the payment of additional search fees.

## FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

1. Claims: 1-16,19-20 (partially)

Human polypeptide comprising the amino acid sequence of SEQ ID NO:1 or any variant having at least 90% amino acid identity with said sequence, the polynucleotide sequence encoding said polypeptide (SEQ ID NO:18) and any variant having at least 90% identity with said polynucleotide sequence; a polynucleotide that hybridizes with said polynucleotide; methods for detection of the polynucleotide; expression vectors and hosts for the recombinant expression of said polypeptide; method for the production of said polypeptide; antibodies against said polypeptide, agonists, antagonists, pharmaceutical compositions containing said polypeptide and uses thereof for the treatment or prevention of a disorder.

2. Claims: 1-16,19-20 (partially)

As subject 1, but comprising the polypeptide sequence of SEQ ID NO:2 and the polynucleotide sequence of SEQ ID NO:19.

3. Claims: 1-16,19-20 (partially)

As subject 1, but comprising the polypeptide sequence of SEQ ID NO:3 and the polynucleotide sequence of SEQ ID NO:20.

4. Claims: 1-16,19-20 (partially)

As subject 1, but comprising the polypeptide sequence of SEQ ID NO:4 and the polynucleotide sequence of SEQ ID NO:21.

5. Claims: 1-16,19-20 (partially)

As subject 1, but comprising the polypeptide sequence of SEQ ID NO:5 and the polynucleotide sequence of SEQ ID NO:22.

6. Claims: 1-16,19-20 (partially)

As subject 1, but comprising the polypeptide sequence of SEQ ID NO:6 and the polynucleotide sequence of SEQ ID NO:23.

7. Claims: 1-16,19-20 (partially)

As subject 1, but comprising the polypeptide sequences of SEQ ID NO:7 and 11 and the respective polynucleotide sequences of SEQ ID NO:24 and 28.

8. Claims: 1-16,19-20 (partially)

### FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

As subject 1, but comprising the polypeptide sequence of SEQ ID NO:8 and the polynucleotide sequence of SEQ ID NO:25.

9. Claims: 1-16,19-20 (partially)

As subject 1, but comprising the polypeptide sequence of SEQ ID NO:9 and the polypucleotide sequence of SEQ ID NO:26.

10. Claims: 1-16,19-20 (partially)

As subject 1, but comprising the polypeptide sequence of SEQ ID NO:10 and the polynucleotide sequence of SEQ ID NO:27.

11. Claims: 1-16,19-20 (partially)

As subject 1, but comprising the polypeptide sequence of SEQ ID NO:12 and the polynucleotide sequence of SEQ ID NO:29.

12. Claims: 1-16,19-20 (partially)

As subject 1, but comprising the polypeptide sequence of SEQ ID NO:13 and the polynucleotide sequence of SEQ ID NO:30.

13. Claims: 1-16,19-20 (partially)

As subject 1, but comprising the polypeptide sequence of SEQ ID NO:14 and the polynucleotide sequence of SEQ ID NO:31.

14. Claims: 1-16,19-20 (partially)

As subject 1, but comprising the polypeptide sequence of SEQ ID NO:15 and the polynucleotide sequence of SEQ ID NO:32.

15. Claims: 1-16,19-20 (partially)

As subject 1, but comprising the polypeptide sequence of SEQ ID NO:16 and the polynucleotide sequence of SEQ ID NO:33.

16. Claims: 1-16,19-20 (partially)

As subject 1, but comprising the polypeptide sequence of SEQ ID NO:17 and the polynucleotide sequence of SEQ ID NO:34.

### FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Claims Nos.: 17,18

Present claims 17 and 18 relate to an extremely large number of possible compounds. Support within the meaning of Article 6 PCT and/or disclosure within the meaning of Article 5 PCT is not to be found, however, for any of the compounds claimed. In the present case, the claims so lack support, and the application so lacks disclosure, that a meaningful search over the whole of the claimed scope is impossible. Consequently, the search has not been carried out for those claims which do not appear to be supported and disclosed, namely those parts relating to the agonists and antagonists of the polypeptides of the invention.

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.